

Glyphosate, AMPA and glufosinate in soils and earthworms in a French arable landscape.

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1 **Abstract**

2 Although Glyphosate-based herbicides are often marketed as environmentally friendly and easily  
3 biodegradable, its bioavailability and risks to wildlife raise significant concerns. Among non-target  
4 organisms, earthworms which live in close contact with the soil can be directly exposed to pesticides  
5 and harmed. We investigated soil contamination and the exposure of earthworms to glyphosate, its  
6 metabolite AMPA, and glufosinate in an arable landscape in France, both in treated (i.e. temporary  
7 grasslands and cereal fields under conventional farming), and nontreated habitats (i.e. hedgerows,  
8 permanent grasslands and cereal fields under organic farming) (n=120 sampling sites in total).  
9 Glyphosate, AMPA and glufosinate were detected in 88%, 58% and 35% of the soil samples, and in  
10 74%, 38% and 12% of the earthworm samples, respectively. For both glyphosate and AMPA,  
11 concentrations in soils were at least 10 times lower than predicted environmental concentrations.  
12 However, the maximum glyphosate soil concentration measured (i.e., 0.598 mg kg<sup>-1</sup>) was only 2 to 3  
13 times lower than the concentrations revealed to affect earthworms (survival and avoidance) in the  
14 literature. These compounds were found both in conventional and organic farming fields, thus  
15 supporting a recent study, and for the first time they were detected in hedgerows and grasslands.  
16 However, glyphosate and AMPA were more frequently detected in soils from cereal fields and  
17 hedgerows than in grasslands, and median concentrations measured in soils from cereal fields were  
18 significantly higher than in the two other habitats. Bioaccumulation of glyphosate and AMPA in  
19 earthworms was higher than expected according to the properties of the molecules. Our findings raised  
20 issues about the high occurrence of glyphosate and AMPA in soils from cropped and more natural  
21 areas in arable landscapes. They also highlight the potential for transfer of these molecules in  
22 terrestrial food webs as earthworms are prey for numerous animals.

23

24 **Keywords:** habitats; agriculture; exposure; terrestrial invertebrates; accumulation

25

## 26 **1. Introduction**

27 Since its introduction on the market in 1974 (Richmond 2018), the global use of glyphosate (N-  
28 (phosphonomethyl)glycine) has increased (Székács and Darvas 2018), making it the most widely used  
29 herbicide in the world. Worldwide, more than 800,000 tons are applied each year and more than 750  
30 commercial formulations containing the molecule are authorized (Benbrook 2016; Kniss 2017),  
31 environmental and health impacts of which have raised numerous questions this past decade (Mesnage  
32 et al., 2015; EPC EU 2017). Whereas the renewal of the marketing authorization for glyphosate  
33 (European Commission, 2017b) evaluated by the European Commission does not find a consensus, the  
34 evaluation reports emphasize the lack of knowledge and data on the concentrations of glyphosate and  
35 metabolites in environmental matrices (in particular animal matrices), and on the assessment of the  
36 risks to biodiversity. This lack of fundamental knowledge and field data on the ecotoxicology of  
37 glyphosate hamper decisions on its regulation and constitutes a limit to the objectives of assessing  
38 risks and impacts of this herbicide. The glufosinate, which is mostly used under the glufosinate-  
39 ammonium form, was considered by the European commission (2017a) as “one of the very few  
40 alternatives to glyphosate”. Regarding the similarities in the chemical structure and properties of the  
41 two herbicides (PPDB, 2021), we can expect a similar behavior of them in soils and earthworms.

42 Despite its theoretical short persistence in soil (half-life DT50 field of 6.45 days; Pesticide Properties  
43 Database (PPDB) 2021), glyphosate and its main degradation product, AMPA  
44 (aminomethylphosphonic acid) (DT50 field of 32 days; Simonsen et al., 2008), are the two most  
45 commonly found pesticides in rivers in France and other European countries (Carles et al. 2019;  
46 Medalie et al. 2020). In the USA and Argentina, glyphosate and AMPA are usually detected together,  
47 occurring widely in sediments, ditches and drains, precipitation, rivers and streams (Aparicio et al.,  
48 2013; Battaglin et al., 2014; Medalie et al., 2020). Data are however scarcer regarding their  
49 concentrations in other environmental compartments such as soil and even more rare in living  
50 organisms. The few studies that measured glyphosate and AMPA in natural soils reported that they  
51 were among the active substances the most frequently found and at the highest concentration in  
52 agricultural and urban soils (Karasali et al., 2019; Geissen et al., 2021; Silva et al., 2019). Geissen et  
53 al. (2021) found AMPA in 96 and 83% of the topsoil samples from conventional and organic fields,

54 respectively. As far as we know, no data exist on the contamination of semi-natural landscape  
55 elements such as hedgerows or grasslands by glyphosate, AMPA and glufosinate. Yet, in farmland,  
56 these habitats are refuges for biodiversity (Geiger et al., 2010) and the occurrence of these compounds  
57 can be a threat for living organisms. Thus, assessing the contamination by glyphosate, its metabolite  
58 AMPA and glufosinate would bring knowledge on biodiversity exposure in these areas.

59 Among non-target organisms, animals living in close contact with the soil can be directly exposed to  
60 pesticides and harmed (Gill et al., 2018; Gunstone et al., 2021). Earthworms are key soil organisms for  
61 their role in soil structure, organic matter dynamics, productivity, and more generally for their  
62 contribution to a number of ecosystem services (Schon and Dominati, 2020). Numerous studies have  
63 shown negative impacts of glyphosate and glyphosate-based herbicides at recommended rates on  
64 earthworms, their activity and the soil functions to which they contribute, with potential implications  
65 on growth, yield and quality performance of plants (Owagboriaye et al., 2020a; Zaller et al., 2021). To  
66 date, only two studies were found on glyphosate bioaccumulation in earthworms, both under  
67 laboratory conditions, and both showing that earthworms bioaccumulated glyphosate proportionally to  
68 the contact period (bioaccumulation factor  $>1$ ) (Andréa et al., 2004; Owagboriaye et al., 2020b). No  
69 data are available on the accumulation of glyphosate under natural conditions, or on AMPA and  
70 glufosinate concentration in earthworms, either in laboratory or under field conditions. Characterizing  
71 the exposure and potential bioaccumulation under natural conditions could help for a better  
72 understanding of the link between agricultural practices and impacts on soil non-target organisms as  
73 well as assess the potential for transfer of the molecules in food webs as earthworms are prey for  
74 numerous animals.

75 The aim of this study was to assess the concentrations of glyphosate, its metabolite AMPA and  
76 glufosinate in soils and earthworms in 120 pesticide-treated and nontreated habitats of a cereal plain.  
77 We hypothesized that the concentrations of glyphosate, AMPA, and glufosinate in soils and  
78 earthworms would be higher in treated habitats than in seminatural habitats and organic fields which  
79 are not directly targeted by pesticides. We also hypothesized, according to literature, that  
80 bioaccumulation of the molecules would occur in soil organisms. Based on the concentrations we  
81 measured in soils and earthworms sampled within the same habitats, bioaccumulation factors were

82 calculated to assess the potential for accumulation in organisms and in food webs of the 3 compounds.  
83 Measured concentrations in soils were compared to predicted environmental concentrations of the  
84 studied pesticides in soils (i.e. PEC<sub>s</sub> provided in risk assessment documents according to the European  
85 regulation) and to toxic thresholds for earthworm to assess potential environmental risk.

86

## 87 **2. Materials and methods**

### 88 **2.1. Sampling area and design**

89 Sampling was conducted during one week in March 2018, in the Long-Term Socio-Ecological  
90 Research Site (LTSER) “Zone Atelier Plaine & Val de Sèvre” (ZAPVS; Bretagnolle et al., 2018),  
91 where forty sampling squares of 1 km<sup>2</sup> were selected (Figure 1). Spring has been chosen as it is a  
92 period of great activity for earthworms (Bouché, 1972).

93 In the sampling area, glyphosate applications occurred at two periods during the cropping season  
94 2017-2018, in autumn (early September 2017) before winter crops sowing, and in spring (April to  
95 mid-May 2018) before spring crops sowing. Glyphosate is never applied during the crop growing  
96 season, being rather mainly applied for weed control before sowing. Thus, it was not applied in all the  
97 sampled cereal fields. It was applied only in one of the sampled cereal fields as a pre-sowing treatment  
98 (i.e., Sampling location 18 C in Table S1). Based on a National survey conducted in agricultural areas  
99 all over France in 2018-2019, glyphosate in air in France was detected in every sample and with  
100 concentrations among the highest (0.05 to 0.2 ng m<sup>-3</sup>) in May and June, while the less likely to be  
101 detected (17-43%) with concentrations being the lowest (around 0.01 ng m<sup>-3</sup>) from November to  
102 February (LCSQA/Ineris, 2020). During the period March-April, the frequency of detection in air  
103 varied around 62 to 75% and the concentrations varied around ranged between 0.025 ng m<sup>-3</sup>  
104 (LCSQA/Ineris, 2020).

105

### 106 **2.2. Collection of soils and earthworms**

107 In each of the forty sampling squares of 1 km<sup>2</sup>, soils and earthworms were sampled in three habitats:  
108 arable fields sown with winter cereals, grasslands and hedgerows (as close as possible to the cereal  
109 field) (Figure 1), for a total of 120 sampling plot locations. Fifteen grassland soils were considered

110 nontreated by pesticides, which came from either temporary grasslands under organic farming ( $n = 10$ )  
111 or permanent grasslands ( $n = 5$ ). Eleven winter cereal soils under organic farming conditions were  
112 collected, and were thus considered nontreated. The farming practices in the organic cereal fields and  
113 grasslands respected the rules of the AB France label and were under organic farming for at least three  
114 years at the time of sampling.

115 At each sampling plot, three soil cores were sampled using a 5 cm  $\varnothing$  soil auger at a 0–5 cm depth.  
116 They were bulked to obtain one composite sample per plot. The 120 soil samples were frozen at  $-20^{\circ}\text{C}$   
117 before being analyzed (Table S1). Soil properties were measured at the Soil Analysis Laboratory of  
118 INRAE (LAS Arras – “Institut National de Recherche pour l’Agriculture, l’Alimentation et  
119 l’Environnement”, France), which benefits from the COFRAC (French accreditation committee)  
120 accreditation of its analytical quality regarding soil characteristics. Briefly, soils were dried at room  
121 temperature and then disaggregated and homogenized before being sieved at 2 mm. The following soil  
122 characteristics were measured, according to international standard methods (for individual references  
123 for the cited standard methods please see AFNOR 2004): pH (water suspension, NF ISO 10390),  
124 organic matter and nitrogen contents (dry combustion in a CHN autoanalyzer Carlo Erba NA 1500, in  
125  $\text{g kg}^{-1}$ ), grain size distribution (NF X 31-107) (clay  $< 2 \mu\text{m}$ , silt 2-20  $\mu\text{m}$ , and sand  $> 20 \mu\text{m}$ , in  $\text{g kg}^{-1}$ ),  
126 total calcium carbonate  $\text{CaCO}_3$  (in  $\text{g kg}^{-1}$ ), and total phosphorus  $\text{P}_2\text{O}_5$  (by ICP-MS spectrometry, in  $\text{g}$   
127  $\text{kg}^{-1}$ , NF ISO 22036, NF X31-147).

128 Between two and five earthworm individuals were also collected at each sampling plot. We chose the  
129 earthworm species *Allolobophora chlorotica* which is well represented in the different sampled  
130 habitats in the ZAPVS. Moreover, this endogeic species lives within the top centimeters of the soil.  
131 Because pesticides generally accumulate at the soil surface, species living in contact with the soil  
132 surface will potentially be more strongly exposed than those living deeper (Pelosi et al., 2013a). The  
133 earthworm individuals were sampled by superficially digging the soil. Before being weighed and  
134 frozen at  $-80^{\circ}\text{C}$ , earthworms were individually placed in petri dishes on damp filter paper for at least  
135 48 h to void their gut contents.

136

### 137 **2.3. Analysis of the pesticide residues**

138 The analysis of glyphosate, aminomethylphosphonic acid (AMPA), and glufosinate in both the 120  
139 soils and 120 earthworm samples (*A. chlorotica*) was performed according to a new sensitive and  
140 selective method (Delhomme et al., 2021) (Table S1). Briefly, the samples (15 g for soils, and between  
141 one to three earthworm individuals pooled according to their mass to attain approximately 1 g) were  
142 extracted with a borate buffer and derivatized with 9-Fluorenylmethyl chloroformate (FMOC-Cl). The  
143 excess FMOC-Cl was removed by liquid-liquid extraction with diethyl ether. The purification of  
144 derivatized extracts was carried out by solid phase extraction (SPE) on Chromabond XLB cartridges  
145 (Macherey-Nagel, France) before internal standard quantification by liquid chromatography coupled to  
146 tandem mass spectrometry (LC/MSMS). The elution step was performed with acidic methanol (1%  
147 formic acid). The extraction and purification method followed by analysis of the two herbicides and  
148 AMPA in soils using LC/MSMS determined limits of detection (LOD) and quantification (LOQ) that  
149 are described in Table 1. Concentrations in soils are expressed in mg kg<sup>-1</sup> dry weight (DW), and  
150 concentrations in earthworms are expressed in mg kg<sup>-1</sup> fresh weight (FW). For quality control, soil and  
151 earthworm samples were spiked with two internal standards before the extraction and derivation  
152 phase. One of the two internal standards was used for the calibration and the second one was used to  
153 evaluate the effectiveness of the derivatization step (see Delhomme et al. (2021) for more details).  
154 Recoveries of glufosinate, glyphosate and AMPA obtained during the optimization steps are shown in  
155 Delhomme et al. (2021).

156 Glyphosate and glufosinate concentrations in soil samples were compared to the recommended field  
157 rate for application, converted in mg kg<sup>-1</sup> soil. For that, we considered that the average glyphosate use  
158 rate ranged from 0.40 to 4.32 kg active ingredient a.i. ha<sup>-1</sup> in cereal crops (Antier et al., 2020; EFSA,  
159 2015). For most glufosinate-based herbicides, the upper limit of recommended application rate is 5 L  
160 ha<sup>-1</sup> with an active concentration in commercial herbicide of 200 g L<sup>-1</sup> (e.g., Dennis et al., 2018 for  
161 Basta®). Considering a penetration of 5 cm depth and a soil bulk density of 1.5 (EFSA, 2017), the  
162 recommended dose of glufosinate is considered as 1.33 mg kg<sup>-1</sup> soil DW and it ranges from 0.53 to  
163 5.76 mg kg<sup>-1</sup> soil DW for glyphosate.

164 To assess pesticides' bioaccumulation in earthworms (i.e. the net uptake of a pesticide from the  
165 environment by all possible exposure routes, e.g., respiration, diet, dermal), a bioaccumulation factor

166 (BAF) was calculated as the ratio of the chemical concentration in earthworm (in mg kg<sup>-1</sup>) fresh weight  
167 (FW) according to OECD (2010)) to the soil concentration (in mg kg<sup>-1</sup> DW). When the soil  
168 concentrations were < LOD, the value LOD/2 was attributed (in order to be able to calculate a ratio).  
169 The ratio with the concentration in earthworm dry weight (DW) was also calculated in order to be able  
170 to compare with the few available data of the literature. As earthworms are composed of 65 to 90%  
171 water depending on the state of hydration (Lee, 1985), we considered 80% and calculated the  
172 concentration in earthworm DW as concentration in earthworm FW/0.2.

173

#### 174 **2.4 Risk assessment for earthworms**

175 The predicted environmental concentrations of glyphosate, AMPA and glufosinate in soils (PECs;  
176 concentrations at recommended application rates obtained from calculation, modelling and/or  
177 measured concentrations in trials), and acute (LC<sub>50</sub>) or chronic (NOEC reproduction) toxicity  
178 thresholds for earthworms (*Eisenia fetida*) were collected from evaluation reports or reports of risk  
179 assessments (i.e. (European Commission, 2015; EFSA, 2005, 2015) and/or scientific publications. *E.*  
180 *fetida* is the species recommended in risk assessment procedures before pesticide marketing  
181 authorization at the European level (ISO 11268-1, 1993). Although it has been shown to be less  
182 sensitive to pesticides and metabolites than earthworm species present in cultivated fields, it was used  
183 in this study because data were available for the three studied chemicals. The risks for earthworms  
184 calculated with *E. fetida* can thus be underestimated.

185 In order to provide quantitative data about the general patterns of contamination with regards to  
186 expected levels in the environment, the measured concentrations in soils (MECs) in our study for  
187 glyphosate, AMPA and glufosinate were compared to the predicted environmental concentrations (i.e.  
188 PECs initial after treatment, long term PECs and maximum PECs calculated for winter wheat when  
189 available, or for other cereals). MECs are hypothesized to be equal to or lower than the maximum  
190 PECs. In order to evaluate the potential ecotoxicity of MECs to earthworms, MECs in soils were also  
191 compared to the values of LC<sub>50</sub> or NOEC for earthworms. This was conducted following the classical  
192 risk assessment method for pesticide regulation defined by European legislation by assessing  
193 toxicity/exposure ratio for earthworms (i.e toxicological benchmarks divided by MECs) with trigger



194 values of 10 for acute toxicity and of 5 for chronic toxicity set as risk limits (TERs above the trigger  
195 limits indicate a negligible risk probability) (European Commission, 2003; EPC EU, 2009).

196

## 197 **2.5. Statistics**

198 Except for the calculation of the BAF (see section 2.3), when a pesticide was not detected in a sample  
199 (value < LOD), the concentration value was set at 0 to perform statistical analyses. When a pesticide  
200 was detected at a level below the LOQ but above the LOD, the LOD value was attributed. ANOVA  
201 was used to compare the pesticide variables (i.e., number of molecules or concentrations) in  
202 earthworms or soils between the three habitats (i.e., cereal fields, grasslands, hedgerows). When  
203 assumptions regarding the normality and homoscedasticity of variances were not respected, we used  
204 Kruskal-Wallis tests. The paired samples t-test was used to compare earthworm and soil pesticide  
205 variables between the two modalities of pesticide use (treated/nontreated) at plot scale. When  
206 assumptions regarding the normality and homoscedasticity of variances were not respected, we used  
207 the Wilcoxon tests. For the differences between conventional and organic systems (t-test or Wilcoxon  
208 test), the data from the hedgerows were removed from the dataset, considering only cereal fields and  
209 grasslands in the analysis. All statistical analyses were performed in RStudio version 3.5.1 using the  
210 packages pgirmess (Giraudoux, 2018) and car (Fox et al., 2018).

211

## 212 **3. Results and discussion**

### 213 **3.1. Occurrence and concentrations in soils**

214 Glyphosate, AMPA and glufosinate were detected in 88%, 58% and 35% of the soil samples  
215 respectively (Table 2a; Table S1). The high frequency of glyphosate and AMPA in the soils is probably  
216 due to the popularity of glyphosate-based herbicides in current agricultural practices (Gill et al., 2018).  
217 Moreover, although it is expected to be biodegradable (Kissane and Shephard, 2017) and non-  
218 persistent in soils, the DT90 lab and field of glyphosate are 297 and 170 days respectively (PPDB,  
219 2021), suggesting that residues could be detected in soils for at least 6 to 10 months after a treatment.  
220 These findings on occurrence are slightly higher for glyphosate than in Geissen et al. (2021) who  
221 found a frequency of 78% in vegetables, orange, grape and potatoes cropping systems, while we

222 detected AMPA less often than in the cited study, which reported a frequency of 83%. The frequencies  
223 of detection of both glyphosate and AMPA were higher here than in Silva et al (2019) who showed  
224 21% and 42% of occurrence, respectively, in 317 EU agricultural topsoils (from conventional crops  
225 including cereals, orchards and vineyards, oilseed rape and sunflower, potatoes and sugar beet, other  
226 vegetables, and dry pulses, flowers and fodder crops such as temporary grasslands, alfalfa, clovers and  
227 strawberries). Despite differences between studies that can be due to cultivated crops, sampling date  
228 with respect to pesticide applications, differences in application rates, climate, depth of soils sampling,  
229 soil properties such as texture, pH, CEC (cation exchange capacity) or organic carbon (Dollinger et al.,  
230 2015; Nguyen et al., 2018), our results are in line with them by revealing residues of glyphosate and  
231 AMPA in a great proportion of arable soils. Moreover, limits of quantification were higher in Geissen  
232 et al. (2021) and Silva et al. (2019) (i.e., 0.05 mg kg<sup>-1</sup> for both glyphosate and AMPA) than in our  
233 study (0.030 mg kg<sup>-1</sup> and 0.025 mg kg<sup>-1</sup> for glyphosate and AMPA, respectively), which can explain  
234 that these authors had lower detection frequencies.

235 All the 58% of the soil samples containing AMPA also contained glyphosate, and in the 12% of cases  
236 where no glyphosate was found, the AMPA was concomitantly undetected. The co-occurrence of  
237 glyphosate and AMPA and the significant correlation between the soil concentrations of glyphosate  
238 and AMPA (Pearson coefficient of 0.37, see Table S2) are coherent considering that AMPA is a  
239 glyphosate's transformation product, thus the parent and the by-product compounds are associated.  
240 However, glyphosate was detected without AMPA in 29% of the samples. AMPA is the main but not  
241 the sole metabolite as an alternative pathway may conduct to sarcosine and glycine (Singh et al.,  
242 2019), which were not investigated in this study. However, unlike glyphosate, AMPA has been  
243 classified as persistent in soils, with a typical half-life of 151 days, but varying from 76 to 240 days  
244 depending on field conditions (Lewis et al., 2016). Studying agricultural topsoils from six different  
245 crop types (i.e. cereals, root crops, non-permanent industrial crops, dry pulses and fodder crops,  
246 permanent crops, vegetables and others) from eleven countries across Europe, Silva et al. (2019)  
247 showed that AMPA was the prominent form, as it occurred in 42% of soils whereas glyphosate was  
248 detected in 21%, and both compounds were present in 18% of the soil samples. Differences with this  
249 study regarding AMPA could be explained by the fate of glyphosate which depends on the considered

250 matrix (e.g., water, different types of soils), sampling date with respect to pesticide applications  
251 (higher glyphosate:AMPA ratio if samples were collected close to the time of application), the  
252 frequency and spatial extent of herbicide-treatments over the studied areas, as well as exogeneous  
253 inputs from wind and water transport (Dollinger et al., 2015; Silva et al., 2019). Moreover, AMPA  
254 may come from other sources than glyphosate degradation (e.g., adjuvants, detergents, plastics), which  
255 may have been less preponderant in the case of the ZAPVS cereal plain.

256 The maximum contents of glyphosate, AMPA and glufosinate in soils were 0.598 (in a grassland),  
257 0.135 and 0.041 mg kg<sup>-1</sup> (in cereal fields for both) respectively (Table 2.a). For glufosinate, this value  
258 is more than 30 times lower than the upper limit of recommended application rate (see section 2.3 for  
259 the calculation). For glyphosate, this value is very close to the calculated lowest value of the  
260 recommended dose (i.e., 0.53 mg kg<sup>-1</sup> in cereal crops, see section 2.3). The measured maximum  
261 contents are much lower than those reported by Silva et al. (2019) (i.e. 2.05 and 1.92 mg kg<sup>-1</sup> for  
262 glyphosate and AMPA respectively) or Laitanen et al. (2006) (i.e., 2.06 and 0.30 mg kg<sup>-1</sup>,  
263 respectively). Similarly, median values were, in our study, 0.088, 0.007 (i.e. < LOQ) mg kg<sup>-1</sup> and <  
264 LOD (0.006 mg kg<sup>-1</sup>) for glyphosate, AMPA and glufosinate, respectively, whereas Silva et al. (2019)  
265 reported 0.14 and 0.15 mg kg<sup>-1</sup> for glyphosate and AMPA, respectively. Some of the differences might  
266 be explained by the higher number of soils sampled investigated in Silva et al. (2019) in a larger  
267 geographical zone, involving a larger range of agricultural practices and climate (i.e., 317 samples,  
268 with southern parts of the EU showing the highest concentrations of glyphosate and AMPA in  
269 topsoils), or by the higher number of nontreated habitats included in our study (i.e. hedgerows,  
270 permanent grasslands and fields under organic farming, versus conventional crops only in Silva et al.  
271 2019). Moreover, analytically speaking, extraction yields for glyphosate and AMPA were highly  
272 variable for two different soil types (Laitinen et al., 2006). Finally, this result could be partly due to  
273 the depth of the soil samples, being 0-5 cm in our study and 0-15/20 cm in Silva et al. (2019).  
274 Although glyphosate and AMPA were found to be mainly retained in the upper soil surface layer (in  
275 the first cm of soil according to Yang et al., 2019), residues can be also found down to 1 m soil depth  
276 (Lupi et al., 2019; Laitinen et al., 2006). For instance, Lupi et al. (2019) mainly found glyphosate and

277 AMPA in soil at a depth of 0-5 cm but they also reported smaller concentrations of residues at 5-9 cm  
278 and deeper (45-60 cm and 130-140 cm).

279

### 280 **3.2. Occurrence and concentrations in earthworms**

281 For earthworms, when considering all samples (i.e., from cereal fields, hedgerows and grasslands), the  
282 frequency of the three studied molecules were lower than in soils (Table 2.b, Table S1). Glyphosate,  
283 AMPA and glufosinate were detected in 74%, 38% and 12% of the samples, respectively (on average  
284 over the three habitats, see Table 2.b). In cereal fields, the frequency of glyphosate is the same for  
285 soils and earthworms (i.e., 95%, see Table 2). In 36% of the sampled earthworms, glyphosate and  
286 AMPA were detected together. In 23% of cases, no glyphosate and no AMPA were found. A  
287 significant correlation was found between the concentrations of glyphosate and AMPA in earthworms  
288 (Pearson coefficient of 0.47, see Table S2). AMPA was detected without glyphosate in 3% of the  
289 earthworm samples, whereas glyphosate was detected without AMPA in 38% of the samples. Thus, as  
290 for soil, AMPA was almost never found without the parent molecule in earthworms. Conversely, more  
291 than a third of samples contained detectable glyphosate but no its metabolite. This might be considered  
292 as surprising given the expected biodegradability of glyphosate, and might suggest a higher  
293 persistence and/or bioavailability of the parent molecule than anticipated from the literature as  
294 emphasized by Kissane and Shephard (2017).

295 The maximum concentrations of glyphosate, AMPA and glufosinate in earthworms were 0.395, 0.247  
296 and 0.04 (< LOQ) mg kg<sup>-1</sup> FW, respectively, being all from cereal fields (Table 2.b, Table 5). Median  
297 values in earthworms were 0.07 (<LOQ) mg kg<sup>-1</sup> FW for glyphosate and < LOD for AMPA and  
298 glufosinate. To our knowledge, this is the first study which quantified the amount of these three  
299 chemicals in earthworms under natural conditions, making it impossible to compare with previous  
300 data.

301 The median value for the calculated BAF (FW/DW) for glyphosate was around 1 (maximum 15.56,  
302 measured four times in nontreated habitats: two organic cereal fields, one hedgerow and one organic  
303 temporary grassland), and around 5 on DW basis (maximum 78) (Table 3). This is in line with the  
304 results reported by Owagboriaye et al. (2020b) under laboratory conditions on two earthworm tropical

305 species which were found to be bioaccumulators and biomagnifiers of glyphosate (BAF > 1 after 8<sup>th</sup>  
306 week post glyphosate application). Similarly, Contardo-Jara et al. (2009) reported that the glyphosate  
307 bioaccumulation factor for *Lumbriculus variegatus*, a sediment dwelling invertebrate, varied between  
308 1.4 and 5.9 which was higher than estimated from chemical properties (i.e., log P<sub>ow</sub>). For AMPA, in  
309 our study, the median BAF (FW/DW) varied between zero in grasslands and hedgerows to 1.62 in  
310 cereals (maximum 67.14 in a conventional cereal field), corresponding to median BAF DW/DW  
311 around 8 in cereals (maximum values from 93 in a hedgerow, 147 in a grassland and 336 in a  
312 conventional cereal plot) (Table 3). This meant that AMPA would be less often accumulated but at  
313 higher concentration than glyphosate and/or that it would be less metabolized by earthworms. The  
314 method developed by Delhomme et al. (2021) to measure glyphosate and AMPA in earthworms  
315 should help going further on toxicokinetics of glyphosate in earthworms.

316 We found in our study that bioaccumulation was higher than expected according to the low measured  
317 or expected bio-concentration factors (BCF, Table 4), low Partition Coefficient n-Octanol/Water (log  
318 K<sub>ow</sub>, Table 4) and the high solubility of glyphosate in water. With a Partition Coefficient Octanol/Air  
319 log K<sub>oa</sub> ranging from 7.26 to 8.40 (Table 4), glyphosate, AMPA and glufosinate fall into the category  
320 of high log K<sub>oa</sub> (i.e., >6) compounds, which have a potential for bioaccumulation in air-breathing  
321 organisms and biomagnification in terrestrial food webs because of slow respiratory elimination rate  
322 (Fremlin et al., 2020; Kelly et al., 2007). Indeed, Kelly et al. (2007) evidenced that moderately  
323 lipophilic compounds having a low log K<sub>ow</sub> (between 2 and 5) but a high K<sub>oa</sub> can biomagnify in food  
324 webs containing air-breathing animals while they do not in aquatic food webs. Here we found that  
325 concentrations of glyphosate and AMPA in soils and earthworms were all highly correlated to each  
326 other (Table S2) and that glyphosate and its transformation products may bioaccumulate in  
327 earthworms, organisms that are a trophic resource for many invertebrates, birds and mammals.  
328 Altogether, this raises questions about the behaviour of glyphosate, AMPA and glufosinate in  
329 terrestrial food webs but, currently, models usable to assess both the trophic transfer and magnification  
330 potential for these types of polar ionic chemicals (i.e., hydrophilic, where lipid is a not the main  
331 storage compartment within the organism) are lacking (Gobas et al., 2016).

332 Glufosinate was found at relatively low frequency and concentrations in soils and earthworms. This is  
333 coherent with the literature which reports a quick degradation of this chemical in soils (DT50 of 7.4  
334 days for glufosinate ammonium, PPDB 2021). Furthermore, glufosinate was much less used than  
335 glyphosate. The reported sales of glyphosate in the county “Deux-Sèvres” where the site is located  
336 reached 113 and 115 tons in 2017 and 2018, respectively, while the sales of glufosinate were reported  
337 as 0.2 and 0.1 tons, respectively (BNVD, 2020). Additional analytical development to investigate the  
338 occurrence and fate of transformation products of glufosinate would be needed to get further insights  
339 into the toxicokinetics and environmental impact of this herbicide.

340

### 341 **3.3. Patterns of contamination according to habitats and agricultural management**

342 Glyphosate was more frequently detected in soils from cereal fields and hedgerows (95 and 93%,  
343 respectively) than in grasslands (75% of the samples) (Table 2a). Moreover, median concentrations of  
344 glyphosate measured in soils from cereal fields were significantly higher than in the two other habitats  
345 (Table 5). When considering only cropped fields as it is done in the rare studies on glyphosate in soils,  
346 glyphosate was more frequently detected but at slightly lower concentrations compared to Geissen et  
347 al. (2021) or Silva et al. (2019). Pesticide use (i.e. treated/nontreated by pesticides) and cropping  
348 system (i.e. field under conventional/organic farming) did not influence the mean concentration of the  
349 three studied molecules (Table 5). This means that the concentration of glyphosate, AMPA and  
350 glufosinate in soils were not influenced by the cropping system, but by the type of habitat itself. This  
351 is in accordance with Geissen et al. (2021) who showed that AMPA was the most frequent residue  
352 found in both conventional and organic fields, with a frequency of 96 and 83%, respectively. Here, the  
353 maximum concentration of glyphosate in soils was reached in a temporary grassland, in which  
354 glyphosate can be used to destroy grass before switching to an annual crop. More surprisingly, the  
355 maximum concentration of AMPA was found in a cereal field under organic farming since 2009.

356 When calculable (i.e., glyphosate concentration > LOD), the median (min value; max value) of the  
357 concentration ratio AMPA:glyphosate was 0.28 (0; 1.01) in cereal fields, 0.09 (0; 8.33) in hedgerows  
358 and 0.00 (0; 0.78) in grasslands. This suggests that the degradation of glyphosate in grasslands is  
359 lower than in the other two habitats. Soil characteristics and notably exchangeable acidity ( $H^+$  and

360  $\text{Al}^{3+}$ ), exchangeable  $\text{Ca}^{2+}$  ions and ammonium lactate extractable K were reported to be the key soil  
361 parameters governing glyphosate mineralization (Nguyen et al., 2018). These parameters were not  
362 measured in our soils but pH, which can inform on exchangeable acidity, was not different between  
363 the three habitats (mean of 8.1, 8.1 and 8.2 for cereal fields, hedgerows and grasslands, respectively,  
364  $n=40$ ). The organic matter content was 46, 88 and 53  $\text{g kg}^{-1}$  in cereal fields, hedgerows and grasslands,  
365 respectively, thus not explaining neither the potential difference in glyphosate degradation between the  
366 three habitats. Moreover, we found no significant correlation between soil concentrations of  
367 glyphosate or AMPA and soil characteristics (Table S2), except a weak one (-0.19) between sand  
368 content and soil glyphosate concentration (Figure S1). As glyphosate was less frequently found in  
369 grasslands compared to the other habitats (Table 2), microorganisms could be less adapted to  
370 glyphosate degradation in this habitat (Schlatter et al., 2017).

371 In earthworms, glyphosate and AMPA were more frequently detected (Table 2b) and at higher  
372 concentrations (Table 5) in cereal fields than in the other two habitats, and the highest glyphosate and  
373 AMPA concentrations (i.e., 0.395 and 0.247  $\text{mg kg}^{-1}$ , respectively) were measured in individuals  
374 sampled in two different conventional cereal fields. The maximum concentration measured for  
375 glyphosate in a cereal field is 50% higher than the maximum raised in hedgerows and grasslands  
376 (Table 2b). As for soils, cropping system (conventional/organic farming) did not influence the mean  
377 concentration of the three studied molecules found in earthworms (Table 5). However, a higher  
378 median concentration of glyphosate was measured in earthworms from treated habitats (0.15  $\text{mg kg}^{-1}$   
379 in temporary grasslands and fields under conventional farming) than from habitats where pesticides  
380 were not applied (<LOQ in hedgerows, permanent grasslands and fields under organic farming) (Table  
381 5). No correlation was found between the concentrations of glyphosate or AMPA in earthworms and  
382 the soil characteristics (Table S2). The BAF for glyphosate did not differ between the three habitats  
383 (Kruskal-Wallis test,  $p = 0.480$ ) but the BAF calculated for AMPA was the highest for earthworms  
384 sampled in cereal fields (BAF cereals > BAF grasslands, Kruskal-Wallis test  $p < 0.001$ ; non-  
385 significant differences between hedgerows and others) (Table 3). This finding underlines potential  
386 issues related to the contamination of an important trophic resource for many terrestrial animals.

387

### 388 3.4. Risk to earthworms

389 Considering predicted environmental concentrations in soils (PEC<sub>s</sub>), the maximum concentrations of  
390 glyphosate were about ten times lower than the maximum PEC<sub>s</sub> which were calculated at 5.974 mg kg<sup>-1</sup>  
391 <sup>1</sup> and 6.616 mg kg<sup>-1</sup> in the worst case for annual and permanent crops, respectively (European  
392 Commission, 2015; EFSA, 2015). For AMPA, measured concentrations reached maximum values that  
393 were 30- to 60-fold lower than accumulated PEC<sub>s</sub> (3.0719 mg kg<sup>-1</sup> and 6.1797 mg kg<sup>-1</sup> for annual and  
394 permanent crops, respectively). Finally, concerning glufosinate, the PEC<sub>s</sub> provided in regulatory  
395 documents reached 2.0 mg kg<sup>-1</sup> initially and 0.32 mg kg<sup>-1</sup> a hundred days after maximum, which is  
396 again several order of magnitude greater than the measured values in the sampled soils (European  
397 Food Safety Authority (EFSA), 2005).

398 Information on glufosinate effects on earthworms are almost inexistent and the concentrations  
399 obtained in this study were low. In the pesticide risk assessment of glufosinate released in 2005  
400 (EFSA, 2005), the LC<sub>50</sub> for earthworms was reported as higher than 1000 mg kg<sup>-1</sup> dry soil for the  
401 active substance as well as for several commercial formulations and the transformation product 3-  
402 methyl-phosphinico-propionic acid (MPP). For the two other transformation products of glufosinate  
403 (i.e., 2-methylphosphinico-acetic acid (MPA) and N-acetyl-glufosinate (NAG)), LC<sub>50</sub> for earthworms  
404 are far from trace levels (> 760 and > 300 mg kg<sup>-1</sup> dry soil, respectively) (EFSA, 2005).

405 One study provides a value allowing to calculate a risk of AMPA concentrations in soils to  
406 earthworms (Von Mérey et al., 2016). The reproductive no-observed-effect concentration (NOEC,  
407 after a 4-week adult exposure period) was 198.1 mg kg<sup>-1</sup> dry soil, a value close to another previous  
408 NOEC for reproduction calculated at 131.9 mg kg<sup>-1</sup> (European Commission, 2015; EFSA, 2015),  
409 which is 800 times higher than the highest AMPA concentrations measured in the sampled soils. This  
410 value was obtained through tests under laboratory conditions with *E. fetida* which is known to be less  
411 sensitive to pesticides than other earthworm species (Pelosi et al., 2013). However, our results, along  
412 with those of Von Mérey et al. (2016), suggested low likelihood of adverse effects of field  
413 concentrations of AMPA on the reproduction of earthworms.

414 The LC<sub>50</sub> for the earthworm *E. fetida* provided in regulatory documents for glyphosate was 5600 mg  
415 active substance kg<sup>-1</sup> dry soil, and the NOEC was 473 mg active ingredient kg<sup>-1</sup> dry soil (European



416 Commission, 2015; EFSA, 2015). These values are far higher than the measured concentrations in  
417 natural soils, ending up in a toxicity-exposure ratio greater than the trigger values, thus suggesting a  
418 low probability of unintentional effects based on the endpoints considered in the toxicity tests on soil  
419 fauna. However, as explained above, these values are obtained from regulatory documents with the  
420 model species *E. fetida* and considering endpoints that are not necessarily the most sensitive. For  
421 glyphosate, lower values have been reported to have detrimental effects on earthworm survival,  
422 reproduction, growth and activity. For instance, the lethal concentration LC<sub>50</sub> for Grassate®, a non-  
423 selective glyphosate-based herbicide, averaged at  $3.045 \pm 0.08$  mg kg<sup>-1</sup> (i.e. mean 1.46 a.i. mg kg<sup>-1</sup>) for  
424 the earthworm *Aporrectodea longa*, a common large species in temperate soils (Ogeleka et al., 2017).  
425 Casabé et al. (2007) found that *E. fetida* individuals avoided soils treated with glyphosate in  
426 formulation (Roundup FG®) at the manufacturers' recommended rate (i.e. 1.44 a.i. kg ha<sup>-1</sup>,  
427 corresponding to 1.92 a.i. mg kg<sup>-1</sup> considering a penetration of 5 cm depth, and a soil bulk density of  
428 1.5 (EFSA, 2017)). This concentration of glyphosate also reduced the earthworm success of  
429 reproduction in the latter study. Thus, the maximum glyphosate soil concentration measured in our  
430 study was only 2 to 3 times higher than these effect concentrations found in the scientific literature. It  
431 is noteworthy that ecotoxicity of commercial formulations of glyphosate can also be due to adjuvants  
432 (Gill et al., 2018). Several other studies have revealed the toxicity of glyphosate and glyphosate-based  
433 herbicides for soil animals, in particular earthworms, and belowground interactions even at rate lower  
434 than recommended by the manufacturer (Martin, 1982; Springett and Gray, 1992; Gill et al., 2018;  
435 Zaller et al., 2014). Our results indicate a context of chronic and low-dose exposure, involving  
436 exposure to concentrations accumulated in tissues while concentrations in soils were no longer  
437 detectable, which inadequately matches with the protocols applied in laboratory standardized tests,  
438 especially in terms of duration. Moreover, links between chronic exposure to glyphosate and nervous,  
439 immune and endocrine systems have been demonstrated (Kissane and Shephard, 2017) and several  
440 studies suggested the predominance of endocrine disrupting mechanisms caused by environmentally  
441 relevant levels of exposure (Mesnage et al., 2015). Considering the scarce data on the exposure of  
442 earthworms to glyphosate and AMPA, the lack of data linking internal concentrations and  
443 ecotoxicological effects, the importance of earthworms in soil functioning, and the socio-economic

444 and environmental issues related the use of glyphosate, further studies are needed on the exposure,  
445 bioaccumulation of glyphosate and AMPA, in relevant earthworm species, and related effects on  
446 populations, communities and related-ecosystem functions. Furthermore, recent finding obtained over  
447 the site studied here has revealed the general occurrence of mixtures of residues of currently used  
448 insecticides, fungicides and herbicides in both soils and earthworms (Pelosi et al., 2021). This  
449 highlights that earthworms may not be exposed to glyphosate and its transformation products only but  
450 to a broad spectrum of organic chemicals likely to interact in terms of toxicodynamics. This renders  
451 crucial the need to better assess and predict risks, to acquire more knowledge about both the  
452 characteristics of pesticide concentrations in the environment and biota, and the effects of exposure to  
453 such complex chemical mixtures on organisms.

454

#### 455 **4. Conclusion**

456 The present study reports for the first-time glyphosate and AMPA concentrations in soils from semi-  
457 natural habitats (hedgerows, permanent grasslands) and in one species of earthworm. We here showed  
458 a high frequency of concentrations of glyphosate and AMPA residues in agricultural soils and in  
459 earthworms, that might be considered as low with regards towards predicted environmental  
460 concentrations and relatively low regarding no effect concentrations for earthworms. In both soils and  
461 earthworms, a ubiquity of the occurrence of residues was evidenced since the compounds were  
462 detected (more frequently and at higher concentrations) in conventional cereal fields but also in  
463 nontreated fields (i.e., under organic farming) and hedgerows. However, pesticide use (i.e.  
464 treated/nontreated by pesticides) and cropping system (i.e. field under conventional/organic farming)  
465 did not influence the mean soil concentration of the three studied molecules. Surprisingly, residues of  
466 glyphosate and/or AMPA were in some cases quantified in earthworms while not detectable in soils  
467 where they were sampled, which raises questions about the inputs and fate of glyphosate-based  
468 herbicides in terrestrial habitats, together with toxicokinetics in soil biota. The bioaccumulation factors  
469 calculated from this dataset, along with previous other studies, highlight the potential of glyphosate  
470 and AMPA to bioaccumulate in terrestrial organisms, and call for further research about the transfer  
471 and trophic magnification risk of these compounds in terrestrial food webs. The situation of

472 glufosinate should be considered as well, since detected while slightly used over the area and sharing  
473 similar physico-chemical properties with glyphosate which could confer abilities for bioaccumulation  
474 in terrestrial food webs. Thus, attention should be paid to those key organisms that promote the soil  
475 functioning, to the fate of contaminants and more generally to the sustainability of agroecosystems.  
476 These concerns regarding glyphosate fate and effects can be extended to wildlife inhabiting cropland  
477 and, seven years later, we echo Battaglin et al. (2014) who concluded that effects of chronic low-level  
478 exposures to glyphosate, AMPA, associated adjuvants and mixtures on ecosystems remain to be  
479 determined.

480

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488

#### 489 **Author Contributions**

490 C.P., C.B. and C.F. coordinated the research project. C.P., C.B., C.F., S.G. and V.B. designed the  
491 experiments and organized the sampling design. C.P. carried out the sampling. C.B. managed data  
492 curation and extraction of land use metrics from maps of ZAPVS in Geographic Information Systems.  
493 C.B. and C.F. prepared the databases. C.P. analyzed the data. C.P. wrote the first draft of the  
494 manuscript. C.F. performed the risk analyses. O.D and M.M. developed the analytical method for  
495 pesticide measurements and performed the analyses. S.N. and M.D. participated to the method  
496 development and supervised the chemical consistency of the interpretations. S.G. and V.B. supervised  
497 the ZAPVS sampling area and collected data on land use and farming practices. All authors  
498 contributed to the writing of the final version of the manuscript, which was revised for English by  
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500

501 **References**

502 AFNOR, 2004. Evaluation de la qualité des sols. 1. Méthodes d'analyse chimique – AFNOR, Saint-  
503 Denis La Plaine, France.

504 Andréa, M., Papini, S., Peres, T.B., Bazarin, S., Savoy, V.L.T., Matallo, M., 2004. Glyphosate:  
505 influence on the soil bioactivity and action of earthworms on its soil dissipation. *Planta Daninha*  
506 22, 95-100.

507 Antier, C., Kudsk, P., Reboud, X., Ulber, L., Baret, P.V., Messéan, A. 2020. Glyphosate Use in the  
508 European Agricultural Sector and a Framework for Its Further Monitoring. *Sustainability*, 12, 5682.  
509 <https://doi.org/10.3390/su12145682>

510 Aparicio, V.C., De Gerónimo, E., Marino, D., Primost, J., Carriquiriborde, P., Costa, J.L., 2013.  
511 Environmental fate of glyphosate and aminomethylphosphonic acid in surface waters and soil of  
512 agricultural basins. *Chemosphere* 93, 1866-1873. DOI: 10.1016/j.chemosphere.2013.06.041

513 Battaglin, W.A., Meyer, M.T., Kuivila, K.M., and Dietze, J.E., 2014. Glyphosate and Its Degradation  
514 Product AMPA Occur Frequently and Widely in U.S. Soils, Surface Water, Groundwater, and  
515 Precipitation. *JAWRA* 50, 275- 290. DOI: 10.1111/jawr.12159

516 Benbrook, C.M., 2016. Trends in glyphosate herbicide use in the United States and globally. *Env. Sci.*  
517 *Eur.* 28, 3. doi: 10.1186/s12302-016-0070-0

518 BNVD, 2020. <https://bnvd.ineris.fr/>

519 Bouché, M.B., 1972. *Lombriciens de France : Écologie et Systématique*, INRA Ann. Zool. Ecol. Anim.  
520 Publication, France.

521 Bretagnolle, V., Berthet, E., Gross, N., Gauffre, B., Plumejeaud, C., Houte, S., Badenhauer, I.,  
522 Monceau, K., Allier, F., Monestiez, P., Gaba, S., 2018. Towards sustainable and multifunctional  
523 agriculture in farmland landscapes: Lessons from the integrative approach of a French LTSER  
524 platform. *Sci. Total Environ.* 627, 822–834. <https://doi.org/10.1016/j.scitotenv.2018.01.142>.

525 Carles, L., Gardon, H., Joseph, L., Sanchís, J., Farré, M., Artigas, J., 2019. Meta-analysis of glyphosate  
526 contamination in surface waters and dissipation by biofilms. *Environ. Int.* 124, 284-293.  
527 <https://doi.org/10.1016/j.envint.2018.12.064>

528 Casabé, N., Piola, L., Fuchs, J., Oneto, M.L., Pamparato, L., Basack, S., Gimenez, R., Massaro, R.,  
529 Papa, J.C., Kesten, E., 2007. Ecotoxicological assessment of the effects of glyphosate and  
530 chlorpyrifos in an Argentine soya field. *Journal of Soils and Sediments* 7, 232-239.  
531 DOI10.1065/jss2007.04.224

532 Contardo-Jara, V., Klingelmann, E., Wiegand, C., 2009. Bioaccumulation of glyphosate and its  
533 formulation Roundup Ultra in *Lumbriculus variegatus* and its effects on biotransformation and  
534 antioxidant enzymes. *Environ. Pollut.* 157, 57-63. doi:10.1016/j.envpol.2008.07.027.

535 Delhomme, O., Hernandez, A., Rodrigues, A., Chimjarn, S., Bertrand, C., Bourdat-Deschamps, M.,  
536 Fritsch, C., Pelosi, C., Nélieu, S., Millet, M., 2021. A method to assess glyphosate, glufosinate and  
537 aminomethylphosphonic acid (AMPA) in soil and earthworms. *J. Chromatogr. A*, 1651, Article  
538 n°462339.

539 Dollinger, J., Dages, C., Voltz, M., 2015. Glyphosate sorption to soils and sediments predicted by  
540 pedotransfer functions. *Environ. Chem. Lett.* 13, 293-307. DOI10.1007/s10311-015-0515-5

541 EFSA (European Food Safety Authority), 2005. Conclusion regarding the peer review of the pesticide  
542 risk assessment of the active substance glufosinate. EFS2 3. <https://doi.org/10.2903/j.efsa.2005.27r>

543 EFSA (European Food Safety Authority), 2015. Conclusion on the peer review of the pesticide risk  
544 assessment of the active substance glyphosate. EFS2 13. <https://doi.org/10.2903/j.efsa.2015.4302>

545 EFSA (European Food Safety Authority), 2017. EFSA Guidance Document for predicting  
546 environmental concentrations of active substances of plant protection products and transformation  
547 products of these active substances in soil. *Efsa J.* 15 (10), 115.  
548 <https://doi.org/10.2903/j.efsa.2017.4982>, 4982.

549 EPC EU, European Parliament and Council of the European Union, 2009. Directive 2009/128/EC of  
550 the European Parliament and of the Council of 21 October 2009 Establishing a Framework for  
551 Community Action to Achieve the Sustainable Use of Pesticides (Text with EEA Relevance).

552 EPC EU, European Parliament and Council of the European Union, 2017. Commission Implementing  
553 Regulation (EU) No 540/2011 as regards the list of approved active substances Text with EEA  
554 relevance - Annex I & II Glyphosate.

555 European Commission, 2003. Technical Guidance Document on Risk Assessment in Support of  
556 Commission Directive 93/67/EEC on Risk Assessment for New Notified Substances, Commission  
557 Regulation (EC) No 1488/94 on Risk Assessment for Existing Substances, Directive 98/8/EC of the  
558 European Parliament and of the Council Concerning the Placing of Biocidal Products on the  
559 Market. Part II (No. EUR 20418 EN/2).

560 European Commission, 2015. Final addendum to the Renewal Assessment Report for the active  
561 substance Glyphosate - public version. EFSA (OpenEFSA portal).

562 European Commission, 2017a. Mergers: Commission Opens In-Depth Investigation into Proposed  
563 Acquisition of Monsanto by Bayer, p. 3, [europa.eu/rapid/press-release\\_IP-17-2762\\_en.htm](http://europa.eu/rapid/press-release_IP-17-2762_en.htm).

564 European Commission, 2017b. Commission Implementing Regulation (EU) 2017/2324 of 12  
565 December 2017 renewing the approval of the active substance glyphosate in accordance with  
566 Regulation (EC) No 1107/2009 of the European Parliament and of the Council concerning the  
567 placing of plant protection products on the market, and amending the Annex to Commission  
568 Implementing Regulation (EU) No 540/2011 (Text with EEA relevance). C/2017/8419.  
569 [http://data.europa.eu/eli/reg\\_impl/2017/2324/oj](http://data.europa.eu/eli/reg_impl/2017/2324/oj)

570 Fox, J., Weisberg, S., Fox, J., 2018. An R Companion to Applied Regression, 3rd ed. SAGE  
571 Publications, Thousand Oaks, Calif.

572 Fremlin, K.M., Elliott, J.E., Green, D.J., Drouillard, K.G., Harner, T., Eng, A., Gobas, F., 2020.  
573 Trophic magnification of legacy persistent organic pollutants in an urban terrestrial food web. *Sci.*  
574 *Tot. Env.* 714. <https://doi.org/10.1016/j.scitotenv.2020.136746>

575 Geiger, F., Bengtsson, J., Berendse, F., Weisser, W.W., Emmerson, M., Morales, M.B., Ceryngier, P.,  
576 Liira, J., Tschamtkke, T., Winqvist, C., Eggers, S., Bommarco, R., Part, T., Bretagnolle, V.,  
577 Plantegenest, M., Clement, L.W., Dennis, C., Palmer, C., Onate, J.J., Guerrero, I., Hawro, V.,  
578 Aavik, T., Thies, C., Flohre, A., Hanke, S., Fischer, C., Goedhart, P.W., Inchausti, P., 2010.  
579 Persistent negative effects of pesticides on biodiversity and biological control potential on  
580 European farmland. *Basic Appl. Ecol.* 11, 97–105. <https://doi.org/10.1016/j.baae.2009.12.001>.

581 Geissen, V., Silva, V., Lwanga, E.H., Beriot, N., Oostindie, K., Bin, Z., Pyne, E., Busink, S., Zomer,  
582 P., Mol, H., Ritsema, C.J., 2021. Cocktails of pesticide residues in conventional and organic

583 farming systems in Europe – Legacy of the past and turning point for the future. *Environ. Pollut.*  
584 278, 116827. <https://doi.org/10.1016/j.envpol.2021.116827>

585 Gill, J.P.K., Sethi, N., Mohan, A., Datta, S., Girdhar, M., 2018. Glyphosate toxicity for animals.  
586 *Environ. Chem. Lett.*, 16, 2, pp. 401-426. DOI: 10.1007/s10311-017-0689-0

587 Giraudoux, P., 2018. Package ‘pgirmess’, Spatial Analysis and Data Mining for Field Ecologists. URL.

588 Gobas, F.A., Burkhard, L.P., Doucette, W.J., Sappington, K.G., Verbruggen, E.M., Hope, B.K.,  
589 Bonnell, M.A., Arnot, J.A., Tarazona, J.V., 2016. Review of existing terrestrial bioaccumulation  
590 models and terrestrial bioaccumulation modeling needs for organic chemicals. *Integr. Environ.*  
591 *Assess. Manag.* 12, 123–134. <https://doi.org/10.1002/ieam.1690>

592 Gunstone, T., Cornelisse, T., Klein, K., Dubey, A., Donley, N., 2021. Pesticides and Soil Invertebrates:  
593 A Hazard Assessment. *Front. Environ. Sci.* 9. <https://doi.org/10.3389/fenvs.2021.643847>

594 ISO (International Organisation for Standardization), 1993. Effects of pollutants on earthworms  
595 (*Eisenia fetida*). Part 1: Determination of acute toxicity using artificial soil substrate – No. 11268–1.  
596 Geneva.

597 LCSQA/Ineris, 2020. Résultats de la Campagne Nationale Exploratoire de mesure des résidus de  
598 Pesticides dans l’air ambiant (2018-2019) (No. DRC-20-172794-02007C).

599 Karasali, H., Pavlidis, G., Marousopoulou, A., 2019. Investigation of the presence of glyphosate and its  
600 major metabolite AMPA in Greek soils. *Environ. Sci. Pollut. Res.* 26, 36308-36321. doi:  
601 10.1007/s11356-019-06523-x

602 Kelly, B.C., Ikonou, M.G., Blair, J.D., Morin, A.E., Gobas, F.A.P.C., 2007. Food Web-Specific  
603 Biomagnification of Persistent Organic Pollutants. *Science* 317, 236–239.  
604 <https://doi.org/10.1126/science.1138275>

605 Kissane, Z., Shephard, J.M., 2017. The rise of glyphosate and new opportunities for biosentinel early-  
606 warning studies. *Conserv. Biol.* 31, 1293-1300. DOI: 10.1111/cobi.12955 Kniss 2017

607 Laitinen, P., Siimes, K., Eronen, L., Rämö, S., Welling, L., Oinonen, S., Mattsoff, L., Ruohonen-Lehto,  
608 M., 2006. Fate of the herbicides glyphosate, glufosinate-ammonium, phenmedipham, ethofumesate  
609 and metamitron in two Finnish arable soils. *Pest Manag. Sci.* 62, 473-491.  
610 <https://doi.org/10.1002/ps.1186>

611 Lee K., 1985. Earthworms, Their Ecology and Relationships with Soils and Land Use. Academic  
612 Press. Sydney.

613 Lewis, K.A., Tzilivakis, J., Warner, D., Green, A., 2016. An international database for pesticide risk  
614 assessments and management. HERAIJ, 22, 1050-1064. DOI: 10.1080/10807039.2015.1133242

615 Lupi, L., Bedmar, F., Puricelli, M., Marino, D., Aparicio, V.C., Wunderlin, D., Miglioranza, K.S.B.,  
616 2019. Glyphosate runoff and its occurrence in rainwater and subsurface soil in the nearby area of  
617 agricultural fields in Argentina. Chemosphere 225, 906–914.  
618 <https://doi.org/10.1016/j.chemosphere.2019.03.090>

619 Martin, N.A., 1982. The effect of herbicides used on asparagus on the growth rate of the earthworm  
620 *Allolobophora caliginosa*. Proc. New Zealand Weed Pest Control Conf. 35, 328–331.

621 Medalie, L., Baker, N.T., Shoda, M.E., Stone, W.W., Meyer, M.T., Stets, E.G., Wilson, M., 2020.  
622 Influence of land use and region on glyphosate and aminomethylphosphonic acid in streams in the  
623 USA. Sci. Total Environ. 707, 136008. <https://doi.org/10.1016/j.scitotenv.2019.136008>

624 Mesnage, R., Defarge, N., Vendômois, J., Séralini, G.-E., 2015. Potential toxic effects of glyphosate  
625 and its commercial formulations below regulatory limits. Food Chem. Toxicol. 84. DOI:  
626 10.1016/j.fct.2015.08.012

627 Nguyen N.K., Dörfler U., Welzl G., Munch J.C., Schroll R., Suhadolc M. (2018) Large variation in  
628 glyphosate mineralization in 21 different agricultural soils explained by soil properties. Sci. Tot.  
629 Environ. 627:544-552. <https://doi.org/10.1016/j.scitotenv.2018.01.204>

630 OECD, Guideline for the Testing of Chemicals, 2010. 317.

631 Ogeleka, D., Onwuemene, C., Okieimen, F., 2017. Toxicity potential of Grassate® a non-selective  
632 herbicide on snails (*Achachatina marginata*) and earthworms (*Aporrectodea longa*). Chem. Ecol.  
633 33, 447-463. DOI: 10.1080/02757540.2017.1320393

634 Owagboriaye, F., Dedeke, G., Bamidele, J., Bankole, A., Aladesida, A., Feyisola, R., Adeleke, M.,  
635 Adekunle, O., 2020a. Wormcasts produced by three earthworm species (*Alma millsoni*, *Eudrilus*  
636 *eugeniae* and *Libyodrilus violaceus*) exposed to a glyphosate-based herbicide reduce growth, fruit  
637 yield and quality of tomato (*Lycopersicon esculentum*). Chemosphere 250, 126270. DOI:  
638 10.1016/j.chemosphere.2020.126270



639 Owagboriaye, F., Dedeké, G.A., Bamidele, J.A., Aladesida, A.A., Isibor, P.O., Feyisola, R.T., Adeleke,  
640 M., 2020b. Biochemical response and vermiremediation assessment of three earthworm species  
641 (*Alma millsoni*, *Eudrilus eugeniae* and *Libyodrilus violaceus*) in soil contaminated with a  
642 glyphosate-based herbicide. *Ecol. Indic.* 108, 105678.  
643 <https://doi.org/10.1016/j.ecolind.2019.105678>.

644 Pelosi, C., Toutous, L., Chiron, F., Dubs, F., Hedde, M., Muratet, A., Ponge, J.-F., Salmon, S.,  
645 Makowski, D., 2013a. Reduction of pesticide use can increase earthworm populations in wheat  
646 crops in a European temperate region. *Agric. Ecosyst. Environ.* 181, 223–230.  
647 <https://doi.org/10.1016/j.agee.2013.10.003>.

648 Pelosi, C., Joimel, S., Makowski, D., 2013b. Searching for a more sensitive earthworm species to be  
649 used in pesticide homologation tests – a meta-analysis. *Chemosphere* 90, 895–900.  
650 <https://doi.org/10.1016/j.chemosphere.2012.09.034>.

651 Pelosi, C., Bertrand, C., Daniele, G., Coeurdassier, M., Benoit, P., Néliu, S., Lafay, F., Bretagnolle,  
652 V., Gaba, S., Vulliet, E., Fritsch, C., 2021. Residues of currently used pesticides in soils and  
653 earthworms: A silent threat? *Agric. Ecosyst. Environ.* 305, 107167.  
654 <https://doi.org/10.1016/j.agee.2020.107167>

655 PPDB—Pesticide Properties DataBase (2021) <https://sitem.herts.ac.uk/aeru/ppdb/en/Reports/246.htm>

656 Richmond, M.E., 2018. Glyphosate: A review of its global use, environmental impact, and potential  
657 health effects on humans and other species. *J. Environ. Stud. Sci.* 8, 416-434. DOI 10.1007/s13412-  
658 018-0517-2

659 Schlatter, D.C., Yin, C., Hulbert, S., Burke, I., Paulitz, T., 2017. Impacts of Repeated Glyphosate Use  
660 on Wheat-Associated Bacteria Are Small and Depend on Glyphosate Use History. *Appl. Environ.*  
661 *Microbiol.* 83. DOI: 10.1128/AEM.01354-17

662 Schon, N.L., Dominati, E.J., 2020. Valuing earthworm contribution to ecosystem services delivery.  
663 *Ecosyst. Serv.* 43, 101092. <https://doi.org/10.1016/j.ecoser.2020.101092>

664 Silva, V., Mol, H.G.J., Zomer, P., Tienstra, M., Ritsema, C.J., Geissen, V., 2019. Pesticide residues in  
665 European agricultural soils – a hidden reality unfolded. *Sci. Total Environ.* 653, 1532–1545.  
666 <https://doi.org/10.1016/j.scitotenv.2018.10.441>.

667 Simonsen, L., Fomsgaard, I.S., Svensmark, B., Spliid, N.H., 2008. Fate and availability of glyphosate  
668 and AMPA in agricultural soil. *J. Environ. Sci. Health B.* 43, 365-75. doi:  
669 10.1080/03601230802062000. PMID: 18576216.

670 Singh, K., Kumar, V., Singh, J. (2019) Kinetic study of the biodegradation of glyphosate by indigenous  
671 soil bacterial isolates in presence of humic acid, Fe(III) and Cu(II) ions. *J. Environ. Chem. Eng.*  
672 7(3):103098. <https://doi.org/10.1016/j.jece.2019.103098>.

673 Székács, A., Darvas, B., 2018. Re-registration Challenges of Glyphosate in the European Union. *Front.*  
674 *Environ. Sci.* 6. <https://doi.org/10.3389/fenvs.2018.00078>

675 Springett, J. A. & Gray, R. A. J., 1992. Effect of repeated low doses of biocides on the earthworm  
676 *Aporrectodea caliginosa* in laboratory culture. *Soil Biol. Biochem.* 24, 1739–1744.  
677 [https://doi.org/10.1016/0038-0717\(92\)90180-6](https://doi.org/10.1016/0038-0717(92)90180-6)

678 von Mérey, G., Manson, P.S., Mehrsheikh, A., Sutton, P., Levine, S.L., 2016. Glyphosate and  
679 aminomethylphosphonic acid chronic risk assessment for soil biota. *Environ. Toxicol. Chem. /*  
680 *SETAC* 35, 2742-2752. DOI: 10.1002/etc.3438

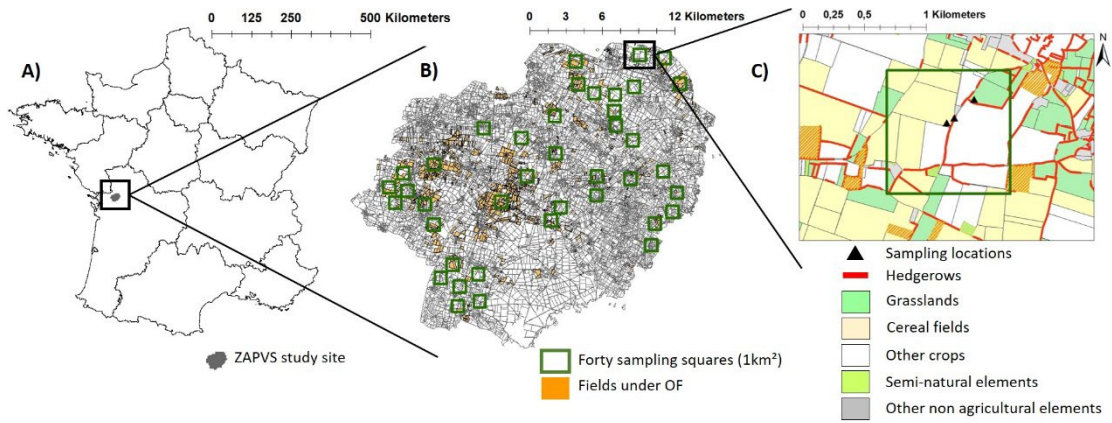
681 Yang, X., Lwanga, E.H., Bemani, A., Gertsen, H., Salanki, T., Guo, X., Fu, H., Xue, S., Ritsema, C.,  
682 Geissen, V., 2019. Biogenic transport of glyphosate in the presence of LDPE microplastics: A  
683 mesocosm experiment. *Environ. Pollut.* 245, 829-835. <https://doi.org/10.1016/j.envpol.2018.11.044>

684 Zaller, J.G., Heigl, F., Ruess, L., Grabmaier, A., 2014. Glyphosate herbicide affects belowground  
685 interactions between earthworms and symbiotic mycorrhizal fungi in a model ecosystem. *Sci. Rep.*  
686 4, 5634. <https://doi.org/10.1038/srep05634>

687 Zaller, J.G., Weber, M., Maderthaner, M., Gruber, E., Takács, E., Mörtl, M., Klátyik, S., Györi, J.,  
688 Römbke, J., Leisch, F., Spangl, B., Székács, A., 2021. Effects of glyphosate-based herbicides and  
689 their active ingredients on earthworms, water infiltration and glyphosate leaching are influenced by  
690 soil properties. *Env. Sci. Eur.* 33, 51. <https://doi.org/10.1186/s12302-021-00492-0>

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692 Figure 1. A) Map of France showing B) the area where the forty sampling squares of 1 km<sup>2</sup> were  
693 selected, and C) the three habitats where sis and earthworms were sampled in each square.



694

695 Table 1. Limits of detection (LOD) and quantification (LOQ) for glyphosate, AMPA and glufosinate  
696 in soil and earthworm matrices.

	Glyphosate		AMPA		Glufosinate	
	Soil	Earthworm	Soil	Earthworm	Soil	Earthworm
697 LOD	0.009	0.070	0.007	0.065	0.006	0.040
LOQ	0.030	0.230	0.025	0.200	0.020	0.120

698 Table 2. Residues of the three chemicals in a) the 120 soils and b) the 120 earthworms in the three  
 699 studied habitats: cereal fields, whether under conventional (treated by pesticides) or organic  
 700 (nontreated) farming, temporary (treated) or permanent (nontreated) grasslands, and hedgerows  
 701 (nontreated). nd for not detected. < LOQ lower than the limit of quantification. Q1 (the first quartile  
 702 concentrations, i.e. 25% of the values above LOQ), the maximum concentrations and the median  
 703 concentrations are expressed in mg kg<sup>-1</sup> dry weight for soil and fresh weight for earthworms. BAF for  
 704 earthworms is the bioaccumulation factor with the standard deviation between brackets.

705

706 a)

b)

	Glyphosate	AMPA	Glufosinate
<b>Cereal fields</b>			
Freq (%)	95	85	50
Q1	0.089	<LOQ	nd
Max	0.432	0.135	0.041
Median	0.142	0.038	<LOQ
<b>Hedgerows</b>			
Freq (%)	93	65	28
Q1	<LOQ	nd	nd
Max	0.322	0.075	<LOQ
Median	0.081	<LOQ	nd
<b>Grasslands</b>			
Freq (%)	75	25	28
Q1	<LOQ	nd	nd
Max	0.598	0.031	<LOQ
Median	<LOQ	nd	nd
<b>All samples</b>			
Freq (%)	88	58	35
Q1	<LOQ	nd	nd
Max	0.598	0.135	0.041
Median	0.088	<LOQ	nd

	Glyphosate	AMPA	Glufosinate
<b>Cereal fields</b>			
Freq (%)	95	68	20
Q1	<LOQ	nd	nd
Max	0.395	0.247	<LOQ
Median	0.225	<LOQ	nd
<b>Hedgerows</b>			
Freq (%)	68	35	15
Q1	nd	nd	nd
Max	0.261	0.239	<LOQ
Median	<LOQ	nd	nd
<b>Grasslands</b>			
Freq (%)	60	13	0
Q1	nd	nd	nd
Max	0.269	0.206	nd
Median	<LOQ	nd	nd
<b>All samples</b>			
Freq (%)	74	38	12
Q1	nd	nd	nd
Max	0.395	0.247	<LOQ
Median	<LOQ	nd	nd

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709

710 Table 3. Median and maximum of the bioaccumulation factor (BAF) of glyphosate and AMPA,  
 711 calculated as the ratio of the chemical concentration in earthworms (in mg kg<sup>-1</sup> fresh weight or dry  
 712 weight) to the soil concentration (in mg kg<sup>-1</sup> dry weight) in the three studied habitats: cereal fields,  
 713 whether under conventional (treated by pesticides) or organic (nontreated) farming, temporary  
 714 (treated) or permanent (nontreated) grasslands, and hedgerows (nontreated).

715

	Fresh weight			Dry weight		
	Q1	Median	Max	Q1	Median	Max
<i>Glyphosate</i>						
Cereal fields	0.78	1.09	15.56	3.90	5.44	77.78
Hedgerows	0	0.90	15.56	0	4.52	77.78
Grasslands	0	1.09	15.56	0	5.44	77.78
All habitats	0	1.00	15.56	0	5.01	77.78
<i>AMPA</i>						
Cereal fields	0	1.62	67.14	0	8.12	335.71
Hedgerows	0	0	18.57	0	0	92.86
Grasslands	0	0	29.43	0	0	147.14
All habitats	0	0	67.14	0	0	335.71

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725 Table 4. Values of Partition Coefficient Octanol/Water ( $\log K_{ow}$ ), Partition Coefficient Octanol/Air  
726 ( $\log K_{oa}$ ), and Bio-concentration Factor (BCF) of glyphosate, AMPA and glufosinate reported in  
727 chemical databases.

728

<b>Compound</b>	<b>Log <math>K_{ow}</math></b>	<b>Log <math>K_{oa}</math></b>	<b>BCF</b>
Glyphosate	-3.12 <sup>§</sup>	8.40 <sup>§</sup>	0.5 <sup>□</sup>
AMPA	-2.42 <sup>§</sup>	7.26 <sup>§</sup>	NA <sup>□</sup>
Glufosinate	-1.74 <sup>§</sup>	8.32 <sup>§</sup>	Considered as low ( $\log K_{ow} < 3$ ) <sup>□</sup>

729

730 <sup>§</sup> refers to US EPA comptox database <https://comptox.epa.gov/dashboard>

731 <sup>□</sup> refers to PPDB database <http://sitem.herts.ac.uk/aeru/ppdb/en/index.htm>

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735 Table 5. Differences in patterns of contamination (median values and maximum between brackets) by  
 736 the three chemicals according to habitats (i.e., cereal fields, whether under conventional (treated by  
 737 pesticides) or organic (nontreated) farming, temporary (treated) or permanent (nontreated) grasslands,  
 738 and hedgerows (nontreated)) and agricultural management (pesticide use and cropping system) in a)  
 739 soils and b) earthworms. For cereal fields, n = 11 under organic farming (nontreated). For grasslands,  
 740 n = 10 under organic farming, and n = 5 in permanent grasslands, for a total of 15 nontreated  
 741 grasslands. Nonparametric Kruskal-Wallis tests were used for all variables. Different letters indicate  
 742 significant differences at p = 0.05 between habitats (one analysis per chemical). For pesticide use and  
 743 cropping system analyses, NS means not significant and \*\*p < 0.01 (Wilcoxon tests).

744

745 a)

Compound				Pesticide use		Cropping system		
	Cereal fields (n=40)		Hedgerows (n=40)		Treated/untreated	Organic/conventional		
Glyphosate	0.142 (0.432)	a	0.080 (0.322)	b	<LOQ	b	NS	NS
AMPA	0.034 (0.135)	a	<LOQ	b	nd	c	NS	NS
Glufosinate	<LOQ	a	nd	a	nd	a	NS	NS

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748 b)

Compound				Pesticide use		Cropping system		
	Cereal fields (n=40)		Hedgerows (n=40)		Treated/untreated	Organic/conventional		
Glyphosate	0.225 (0.395)	a	<LOQ	b	<LOQ	b	**	NS
AMPA	<LOQ	a	nd	b	nd	b	NS	NS
Glufosinate	nd	a	nd	a	nd	a	NS	NS

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Graphical abstract

