

## Research



**Cite this article:** Dupoué A, Brischoux F, Lourdaï O. 2017 Climate and foraging mode explain interspecific variation in snake metabolic rates. *Proc. R. Soc. B* **284**: 20172108. <http://dx.doi.org/10.1098/rspb.2017.2108>

Received: 20 September 2017

Accepted: 16 October 2017

**Subject Category:**

Evolution

**Subject Areas:**

ecology, evolution

**Keywords:**

climate, ectotherm, lifestyle, metabolic cold adaptation, metabolic rate

**Author for correspondence:**

Andréaz Dupoué

e-mail: [andreaz.dupoue@gmail.com](mailto:andreaz.dupoue@gmail.com)

Electronic supplementary material is available online at <https://dx.doi.org/10.6084/m9.figshare.c.3923218>.

# Climate and foraging mode explain interspecific variation in snake metabolic rates

Andréaz Dupoué<sup>1</sup>, François Brischoux<sup>2</sup> and Olivier Lourdaï<sup>2,3</sup>

<sup>1</sup>CNRS UPMC, UMR 7618, iEES Paris, Université Pierre et Marie Curie, Tours 44-45, 4 Place Jussieu, 75005 Paris, France

<sup>2</sup>CEBC-CNRS, UMR 7372, 79360, Villiers en Bois, France

<sup>3</sup>School of Life Sciences, Arizona State University, Tempe, AZ 85287-4501, USA

AD, 0000-0002-2501-464X; FB, 0000-0002-5788-1326

The energy cost of self-maintenance is a critical facet of life-history strategies. Clarifying the determinant of interspecific variation in metabolic rate (MR) at rest is important to understand and predict ecological patterns such as species distributions or responses to climatic changes. We examined variation of MR in snakes, a group characterized by a remarkable diversity of activity rates and a wide distribution. We collated previously published MR data ( $n = 491$  observations) measured in 90 snake species at different trial temperatures. We tested for the effects of metabolic state (standard MR (SMR) versus resting MR (RMR)), foraging mode (active versus ambush foragers) and climate (temperature and precipitation) while accounting for non-independence owing to phylogeny, body mass and thermal dependence. We found that RMR was 40% higher than SMR, and that active foragers have higher MR than species that ambush their prey. We found that MR was higher in cold environments, supporting the metabolic cold adaptation hypothesis. We also found an additive and positive effect of precipitation on MR suggesting that lower MR in arid environments may decrease dehydration and energetic costs. Altogether, our findings underline the complex influences of climate and foraging mode on MR and emphasize the relevance of these facets to understand the physiological impact of climate change.

## 1. Introduction

Understanding metabolic adaptation has attracted considerable scientific interest [1,2]. Whole organism metabolic rate (MR) is an integrative parameter of biochemical kinetics that can be related to major traits such as growth, survival and reproduction [3–5]. In this context, energy requirement for self-maintenance (i.e. either standard or resting MR, designated hereafter as SMR and RMR [6]) is a crucial facet of life-history variation both at the individual and species level [2,4]. Variations in MR are primarily driven by allometric factors (body size) and temperature dependence (Arrhenius relationship) [7] but are also functionally related to life-history strategies (fast versus slow pace of life [8]) and to climatic conditions [9,10]. Therefore, MR ultimately correlates with major ecological traits such as species distribution and responses to climatic change [5,9,11].

Important interspecific variation exists in resting energy requirements. For example, endothermic vertebrates (i.e. birds and mammals) have an elevated metabolism at rest which is essential to support high maximal MR and sustain prolonged activity (i.e. aerobic scope model [12]). This metabolic specialization has profound implications from subcellular (e.g. ATP production by mitochondria) to the whole organism level (e.g. muscle performance, activity level) [13]. Substantial variation in MR also exists depending on ecological traits such as foraging mode, energy intake capacity or diet [14]. For instance, among ectotherms, sustained locomotor performances in active foragers are usually associated

with higher MR compared with sit-and-wait species as demonstrated in arthropods [15], fishes [16], amphibians [17] and reptiles [6,18].

Interspecific variation in MR can also be related to environmental gradients and reflect climatic adaptation [9,19]. Notably, when measured at the same body temperature, species from cold climates (high altitude or latitude) usually display higher MR than those from warmer climates. The slope between MR and temperature can also be steeper under cold climates [20]. This difference in MR-temperature relationship is described as the metabolic cold adaptation (MCA [21]), and appears essential to complete the life cycle and perform critical activities despite thermal constraints. Although this hypothesis is still debated [22,23], it has received substantial empirical support in most taxa (i.e. negative relationships between MR and environmental temperature) such as in insects [10], fishes [22], amphibians [24], reptiles [25], birds [26] and mammals [27].

Conversely, the determinants of MR in warm climates are relatively rather overlooked [26,28]. Lower MR in warm climates can be beneficial, for example, in limiting energy expenditure but also dehydration through lowered rates of ventilatory water loss [29]. The influence of water availability on MR variation is crucial in the context of climate change that involves both changes in temperature and precipitation regimes. Several studies previously examined the influence of global warming on species functional traits [30,31], but the impacts of aridity changes are currently overlooked [32].

Snakes (more than 3000 species [33]) represent particularly relevant models to clarify the influence of climatic conditions and foraging modes on MR. Snakes typically have limited energy requirements associated with important fasting capacities. Their body temperature is closely influenced by environmental thermal conditions and also behavioural thermoregulation. As ectotherms, there is an exponential increase in MR with body temperature [34]. Snakes are widely distributed from deserts and tropics to boreal climates, thereby allowing the exploration of climatic influences [25,35]. Finally, their diversity in life-history strategies provides a unique opportunity to investigate variation in MR notably according to foraging modes that range from sit-and-wait to active predation [18].

Previous studies have investigated the determinant of MRs in squamates focusing notably on the influences of lifestyle [6,18]. Yet, we emphasize that additive influences of climatic conditions and phylogeny on MR have not been considered to date. In this study, we investigated the effects of foraging mode combined with climate (temperature and precipitation) on MR in 90 species of the main snake clades. We constituted a dataset based on MR at rest (SMR and RMR) measured at different trial temperatures ( $T_{\text{trial}}$ ) including phylogenetic proximity among species [36]. In this context, we hypothesized that snake MR should be explained by the combined influences of lifestyle (foraging mode) and climate and we tested the following predictions:

- (i) to sustain higher aerobic activity, species that actively hunt their prey (active foragers) should have higher MR than species that wait in ambush [6,18];
- (ii) according to the MCA hypothesis, MR (corrected for body mass and  $T_{\text{trial}}$ ) should negatively correlate with environmental temperature because higher MR in cold climates compensates for lower average body temperature [21]; and
- (iii) because low MR helps to limit the rate of water loss, snake MR (corrected for body mass and  $T_{\text{trial}}$ ) should

be positively correlated with precipitation, so that species from arid climates should display lower rates of water loss than species from humid climates [26,37].

## 2. Material and methods

### (a) Dataset

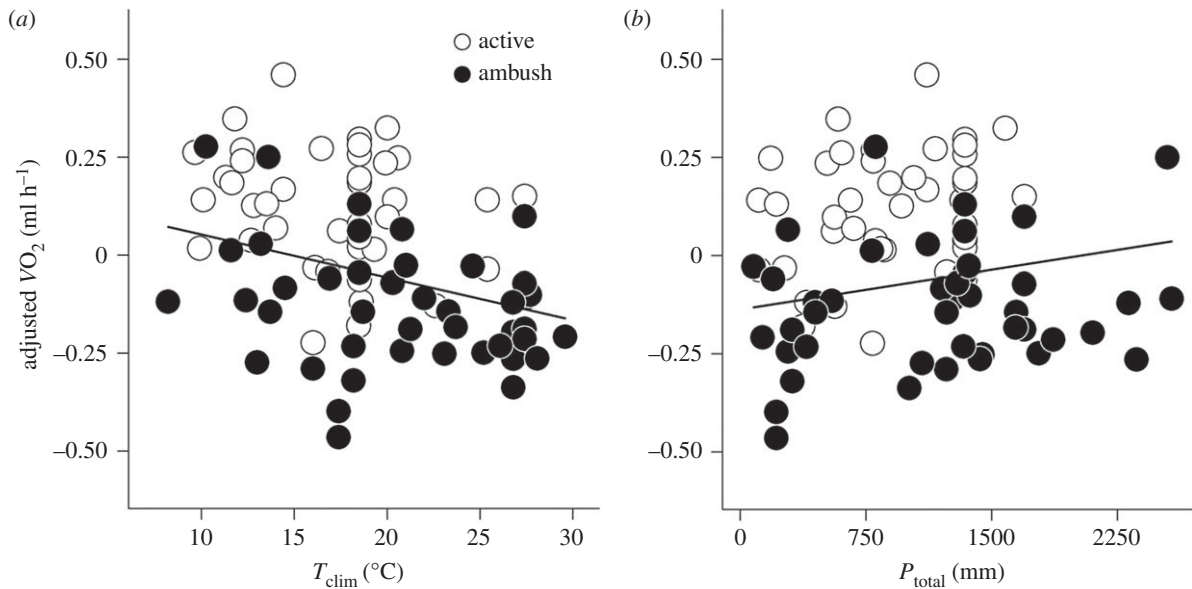
We reviewed the published literature on snake oxygen consumption ( $VO_2$ ), which is an indirect measure of whole organism MR [38] to compile the dataset (90 species from 76 studies; electronic supplementary material, table S1). We only extracted  $VO_2$  values collected when individuals were post-absorptive and inactive and we assessed metabolic state (i.e. SMR or RMR) following the previously established classification [6]. That is, RMR is measured during the active period of the daily cycle (daytime for most snake species), while SMR is measured during the inactive period of the daily cycle (night for most snake species) [6].  $VO_2$  values were either given in the text, derived from allometric equations ( $VO_2 = a \times \text{Mass}^b$ ) provided within each study, or extracted from figures using a specific software (PLOTDIGITIZER v. 2.6.6) when neither values nor equations per species were provided in text [39–41], because these three methods did not affect snake  $VO_2$  (all  $p > 0.360$ ). When authors measured carbon dioxide production ( $VCO_2$ ), we used the respiratory quotient of post-absorptive uricotelic carnivores ( $RQ = 0.72$ ), to estimate snake  $VO_2$ :  $VO_2 = VCO_2/RQ$ . Data were collected at various  $T_{\text{trial}}$  ranging from 5 to 35°C leading to a total of 491 observations (electronic supplementary material, table S1). We also included data obtained from eight species (electronic supplementary material, table S1), for which we used a closed-system respirometry to measure snake SMR at three  $T_{\text{trial}}$  (10°C, 20°C and 30°C) (for details in methods, see [25]).

### (b) Foraging mode assignment

Snakes are characterized by a continuum in foraging mode ranging from sit-and-wait to active predation. For simplicity, we used a categorical approach and assigned to each species a foraging mode. We used supplementary information from [42] as a basis and we completed the dataset with published data and personal experience. Species that are known to rely mainly on sit-and-wait strategies (e.g. most viperids or boids) were classified as ‘ambush’ predators, whereas species relying on important movement for resource acquisition (e.g. most colubrids, natricines or lamprophids) were classified as ‘active’ foragers (electronic supplementary material, table S1).

### (c) Climatic conditions

We used the mean annual temperature and total annual precipitation (obtained from <http://en.climate-data.org>) to approximate thermal conditions ( $T_{\text{clim}}$ ) and total precipitation ( $P_{\text{total}}$ ) experienced at the locality of origin described in the study. When locality was lacking, (e.g. not provided or captive animals), we randomly chose a locality within species geographical range (see the electronic supplementary material, table S1 for all localities) because this procedure did not impact the following results. Although large-scale climatic variables do not permit us to address microclimatic variation and actual body temperature, they reflect contrasted thermal environment that can correlate with physiological adaptations [43]. Preliminary analyses showed that  $T_{\text{trial}}$  used in the different studies were related to local  $T_{\text{clim}}$  because MR was usually measured in an ecologically relevant thermal range. That is, species from cold climates were measured in relatively cold  $T_{\text{trial}}$  as compared to species from warm climates (generalized linear models,  $T_{\text{clim}} \sim T_{\text{trial}}$ :  $F_{5,387} = 7.1$ ,  $p < 0.001$ ).



**Figure 1.** Effects of climatic conditions on  $VO_2$  adjusted for body mass and  $T_{\text{trial}}$  (species  $VO_2$  coefficients derived from a mixed model including fixed effects of body mass and  $T_{\text{trial}}$ , random effect of species as a random factor, and phylogenetic signal set to  $\lambda = 0.2$ ). (a) Negative relationship between  $VO_2$  and habitat temperature  $T_{\text{clim}}$  and (b) positive relationship between  $VO_2$  and total precipitation  $P_{\text{total}}$ .

#### (d) Statistical analysis

All statistics were performed with R software (R Development Core Team, v. 3.2.0, <http://cran.r-project.org/>).  $VO_2$  and mass were  $\log_{10}$  transformed to achieve normal distributions. We built a mixed effect model (package *coxme* [44]) initially designed from pedigree analyses to test for the effects of body mass,  $T_{\text{trial}}$ , respirometry method (closed versus open-flow system), metabolic state (RMR or SMR), foraging mode, climatic conditions ( $T_{\text{clim}}$  and  $P_{\text{total}}$  at the locality) on snake  $VO_2$ . We accounted for multiple  $VO_2$  measures by treating species as random factor. Phylogenetic relationships between species were included in this model using a recent molecular-based study [36] (electronic supplementary material, figure S1). We excluded four species that were not in the phylogeny from [36]: *Dasypeltis inornata*, *Helicops modestus*, *Pseudonaja nuchalis*, and *Philodryas chamissonis* (electronic supplementary material, table S1). To determine the influence of phylogenetic relationships on  $VO_2$ , we adjusted the scaling-factor for the correlation of the residual error ( $\lambda$ , the coefficient that applies on branch length) (package *ape*, [45]). For  $\lambda = 1$ , traits follow a constant-variance model of evolution equivalent to a Brownian motion model, while for  $\lambda = 0$ , trait evolution is supposedly independent between species corresponding to an ordinary least square model [46]. For  $0 < \lambda < 1$ , phylogeny covariance among species is overestimated and supposes different levels of phylogenetic signal [46]. Figures were obtained after extracting  $VO_2$  coefficients for each species in the global model.

### 3. Results

#### (a) Influence of phylogenetical signal, body mass, $T_{\text{trial}}$ , metabolic state and respirometry method

Log likelihood ratio tests showed that the final model was obtained for  $\lambda = 0.2$  (see the electronic supplementary material, table S2).

$VO_2$  was significantly positively influenced by mass ( $\beta = 0.86 \pm 0.02$ ,  $z_{1,490} = 45.2$ ,  $p < 0.001$ ) and  $T_{\text{trial}}$  ( $\beta = 0.04 \pm 0.00$ ,  $z_{1,490} = 36.7$ ,  $p < 0.001$ ). We found a significant influence of metabolic state ( $\beta = 0.06 \pm 0.02$ ,  $z_{1,490} = 3.1$ ,  $p = 0.002$ ) with

RMR being on average 39.87% higher than SMR. In turn, MR was not influenced by respirometry method ( $\beta = 0.05 \pm 0.02$ ,  $z_{1,490} = 1.9$ ,  $p = 0.058$ ).

#### (b) Influence of foraging mode and climatic conditions

We found a significant influence of foraging mode ( $\beta = 0.31 \pm 0.04$ ,  $z_{1,490} = 7.7$ ,  $p < 0.001$ ) with species that actively hunt their prey having  $VO_2$  (adjusted for body mass and  $T_{\text{trial}}$ ) 28% higher than sit-and-wait foragers. Finally, we found that  $VO_2$  (adjusted for body mass and  $T_{\text{trial}}$ ) was negatively influenced by  $T_{\text{clim}}$  ( $\beta = -8.03 \times 10^{-3} \pm 2.57 \times 10^{-3}$ ,  $z_{1,490} = -3.1$ ,  $p = 0.002$ ; figure 1a) and positively influenced by  $P_{\text{total}}$  ( $\beta = 6.7 \times 10^{-5} \pm 2.97 \times 10^{-5}$ ,  $z_{1,490} = 2.2$ ,  $p = 0.024$ ; figure 1b).

### 4. Discussion

We demonstrated that foraging mode, climate (temperature and precipitation) independently contribute to explain interspecific variation in snake MR. Our results show that active foragers have higher maintenance requirements than species relying on sit-and-wait strategies, and that MR is negatively correlated to temperature but positively correlated to precipitation. To our knowledge, this is the first empirical demonstration of combined effects of foraging mode and climate on MRs in snakes.

We found a weak phylogenetic signal ( $\lambda = 0.2$ ) suggesting that the phylogeny describing residual covariance was nearly a star with little hierarchical structure. This results contrast with others studies on ecological correlates of MRs in ectotherms that have detected a more important contribution of phylogeny [47]. Although our finding may be owing to a strong covariation pattern between phylogeny and foraging mode in snakes, we emphasize that other phylogenetically informed analyses in this group have led to similar weak phylogenetic signals [42,48]. Such contrasts among lineages clearly warrant further explorations [49].

Factors affecting MR are multiple, including allometry, thermal dependence but also life-history strategies and environmental gradients. Coefficients for mass-scale and  $T_{\text{trial}}$  were highly similar with those previously reported [6]. The approximately 40% difference between RMR and SMR is also consistent with these previous findings. This considerable source of variation underlines the critical importance of reporting the metabolic state to permit comparative studies. In support of our first prediction, we found that active foragers have higher MR (either standard or resting) than species relying on a sit-and-wait strategy. Similar effects of foraging mode on MR have been reported in several species [6,18,50]. A close relationship between resting and maximal MR is expected [51]. While high MR is costly [4], active species benefit from high MR at rest owing to enhanced maximal MR and sustained activity capacities. A previous meta-analysis in snakes also showed that active foragers have a higher proportion of red blood cells to optimize oxygen delivery [42]. Our results also provided support to the MCA hypothesis [21], with a counter gradient variation between MR and environmental temperature. Therefore, species from cold climates have higher MR than species from warm ones when accounting for  $T_{\text{trial}}$ . Ectotherms primarily rely on behavioural means to regulate body temperature and optimize performance (e.g. locomotion, digestion, immunity, reproduction) [34]. Under cold climates, elevated MR provides a positive compensation to efficiently perform these physiological functions despite low ambient temperatures and reduced thermal opportunities [34]. Higher MR can result from both adaptation and physiological plasticity such as thermal acclimation to local conditions [52]. While our study does not permit us to separate their relative contribution the trends detected cannot be solely attributed to acclimation and probably reflect climatic adaptations [20,25].

The negative relationship between MR and temperature may also involve a specific response to warm climates for energy saving strategies. That is, higher ambient temperatures probably translate in higher mean body temperature and lower MR may contribute to minimize energy costs of self-maintenance [21]. Concomitantly, a lower MR may limit the production of reactive oxygen species and oxidative damage [53], and the rate of dehydration through lower respiratory water loss [29]. In this perspective, our results support a positive effect of precipitation on snake MR at rest which is consistent with previous studies in vertebrates [26,37,54]. Arid or xeric environments are typically characterized by limited access to energy and water resources [55]. At the individual level, diverse responses help in limiting the dehydration rate when access to water is limited, which include physiological (e.g. metabolic depression [56]) or behavioural adjustments (e.g. shift in thermoregulation [57]; change in activity level [58]). Therefore, species exposed to arid conditions might benefit from lower MR to limit the risk of dehydration through lower pulmo-cutaneous water loss [29,59]. Aridity probably correlates with other parameters such as primary productivity and reduced MR at rest may therefore also reflect limiting energy availability. However, a recent comparative analysis in birds showed that precipitation *per se* and not primary production drive metabolic adaptations [26]. Further studies are therefore required to understand how precipitation shapes metabolic adaptations independently of ecosystem productivity. Notably, while water constraints strongly affect species

persistence [60], precipitation is usually not included in predictive models on climatic changes. General trends predict that annual rainfall should decrease in subtropical regions while it should increase in temperate areas [61]. Water requirements are, therefore, critical to integrate in order to understand and predict species responses to climate change.

Our study focuses on large-scale climatic gradients and their influence on physiological trends [43]. Yet, it would be important to account for variation in thermoregulation strategies. For instance, sympatric species can display contrasted thermal preference, microhabitat selection [62]. Such differences can directly contribute to energy and water budget and sensitivity to climatic factors [62]. Similarly, average temperatures at the locality do not provide information on thermal heterogeneity of the habitat which can strongly influence achieved body temperature [63]. While detailed information on species trait and thermal variation are often lacking they should contribute to refine the general trend observed in our model.

Overall, previous studies focusing on the determinants of MR at rest may have overlooked the additive effects of environmental temperature, precipitation and foraging modes. Our study provides, to our knowledge, for the first time clear evidence that MR at rest is better explained by the combined effects of lifestyle and climate. Previous studies suggest that tropical squamates are vulnerable to climate change owing to narrower thermal limits [64]. Yet maintenance requirements are also crucial to consider and will be profoundly affected by temperature increase. We posit that active foraging and/or cold adapted species will be particularly sensitive to climate change owing to their elevated MR at rest. Our results support lower MRs in arid environments as an energy and water saving mechanism. Projected changes in rainfall patterns may influence water loss and compromise water balance notably in tropical species with elevated MRs. Behavioural thermoregulation also has strong potential for buffering the physiological constraints induced by climate change [65]. Therefore, further work is required to better understand the short- (plastic) and long-term (evolutionary) physiological and behavioural responses to complex environmental changes.

**Ethics.** All experiments were performed in accordance with laws related to the capture, experiments and welfare of the animals (Dérogation du ministère de l'écologie, de l'énergie, du développement durable et de la mer no. 09/346/DROG; permit no. 792, Direction service vétérinaire des Deux-Sèvres).

**Data accessibility.** The dataset is available online in the electronic supplementary material, table S1.

**Authors' contributions.** All authors conceived the ideas and methodology; A.D. collected and analysed the data; A.D. led the writing of the manuscript. All authors contributed critically to the writing and gave final approval for publication.

**Competing interests.** We declare that we have no competing interests.

**Funding.** Financial support was provided by the CNRS, the ANR JJC Ectoclim, the Conseil Départemental des Deux-Sèvres and the Conseil Régional Poitou Charentes.

**Acknowledgements.** Authors thank the following persons who helped with capturing snakes in the field: Jean-Pierre Vacher, Gaëtan Guiller, Mickaël Ricordel, Anthony Olivier, Matthieu Berroneau, Yannig Bernard and Benoit Heulin. We are also grateful to Bernard Ragot and the Zoodyssée for providing access to captive individuals. We thank Tom Van Dooren and Jean François Le Galliard for their precious help in analyses. We finally thank Pauline Blaimont for English revision.

## References

- Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL. 2001 Effects of size and temperature on metabolic rate. *Science* **293**, 2248–2251. (doi:10.1126/science.1061967)
- White CR, Kearney MR. 2013 Determinants of interspecific variation in basal metabolic rate. *J. Comp. Physiol. B* **183**, 1–26. (doi:10.1007/s00360-012-0676-5)
- Hochachka PW, Somero GN. 2002 *Biochemical adaptation*. Princeton, NJ: Princeton University Press.
- Burton T, Killen SS, Armstrong JD, Metcalfe NB. 2011 What causes intraspecific variation in resting metabolic rate and what are its ecological consequences? *Proc. R. Soc. B* **278**, 3465–3473. (doi:10.1098/rspb.2011.1778)
- Kearney M. 2012 Metabolic theory, life history and the distribution of a terrestrial ectotherm. *Funct. Ecol.* **26**, 167–179. (doi:10.1111/j.1365-2435.2011.01917.x)
- Andrews RM, Pough HF. 1985 Metabolism of squamate reptiles: allometric and ecological relationships. *Physiol. Zool.* **58**, 214–231. (doi:10.1086/physzool.58.2.30158569)
- Brown JH, Gillooly JF, Allen AP, Savage VM, West GB. 2004 Toward a metabolic theory of ecology. *Ecology* **85**, 1771–1789. (doi:10.1890/03-9000)
- Wiersma P, Muñoz-García A, Walker A, Williams JB. 2007 Tropical birds have a slow pace of life. *Proc. Natl Acad. Sci. USA* **104**, 9340–9345. (doi:10.1073/pnas.0702212104)
- Chown SL, Gaston KJ. 1999 Exploring links between physiology and ecology at macro-scales: the role of respiratory metabolism in insects. *Biol. Rev. Camb. Philos. Soc.* **74**, 87–120. (doi:10.1017/S000632319800526X)
- Addo-Bediako A, Chown SL, Gaston KJ. 2002 Metabolic cold adaptation in insects: a large-scale perspective. *Funct. Ecol.* **16**, 332–338. (doi:10.1046/j.1365-2435.2002.00634.x)
- Dillon ME, Wang G, Huey RB. 2010 Global metabolic impacts of recent climate warming. *Nature* **467**, 704–706. (doi:10.1038/nature09407)
- Hayes JP, Garland T. 1995 The evolution of endothermy: testing the aerobic capacity model. *Evolution* **49**, 836–847. (doi:10.1111/j.1558-5646.1995.tb02320.x)
- Hulbert AJ. 2008 The links between membrane composition, metabolic rate and lifespan. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* **150**, 196–203. (doi:10.1016/j.cbpa.2006.05.014)
- McNab BK. 1986 The influence of food habits on the energetics of eutherian mammals. *Ecol. Monogr.* **56**, 1–19. (doi:10.2307/2937268)
- Greenstone MH, Bennett AF. 1980 Foraging strategy and metabolic rate in spiders. *Ecology* **61**, 1255–1259. (doi:10.2307/1936843)
- Goolish EM. 1991 Anaerobic swimming metabolism of fish: sit-and-wait versus active forager. *Physiol. Zool.* **64**, 485–501. (doi:10.1086/physzool.64.2.30158187)
- Taigen TL, Pough FH. 1983 Prey preference, foraging behavior, and metabolic characteristics of frogs. *Am. Nat.* **122**, 509–520. (doi:10.1086/284152)
- Beaupre SJ, Montgomery CE. 2007 The meaning and consequences of foraging mode in snakes. In *Lizard ecology: the evolutionary consequences of foraging mode* (eds SM Reilly, LD McBrayer, DB Miles), pp. 334–367. New York, NY: Cambridge University Press.
- Gaston KJ *et al.* 2009 Macrophysiology: a conceptual reunification. *Am. Nat.* **174**, 595–612. (doi:10.1086/605982)
- Terblanche JS, Clusella-Trullas S, Deere JA, Van Vuuren BJ, Chown SL. 2009 Directional evolution of the slope of the metabolic rate-temperature relationship is correlated with climate. *Physiol. Biochem. Zool.* **82**, 495–503. (doi:10.1086/605361)
- Clarke A. 2006 Temperature and the metabolic theory of ecology. *Funct. Ecol.* **20**, 405–412. (doi:10.1111/j.1365-2435.2006.01109.x)
- White CR, Alton LA, Frappell PB. 2011 Metabolic cold adaptation in fishes occurs at the level of whole animal, mitochondria and enzyme. *Proc. R. Soc. B* **279**, 1740–1747. (doi:10.1098/rspb.2011.2060)
- Gaitan-Espitia JD, Nespolo R. 2014 Is there metabolic cold adaptation in terrestrial ectotherms? Exploring latitudinal compensation in the invasive snail *Cornu aspersum*. *J. Exp. Biol.* **217**, 2261–2267. (doi:10.1242/jeb.101261)
- Navas CA, Gomes FR, Carvalho JE. 2008 Thermal relationships and exercise physiology in anuran amphibians: integration and evolutionary implications. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* **151**, 344–362. (doi:10.1016/j.cbpa.2007.07.003)
- Lourdais O, Guillon M, DeNardo DF, Blouin-Demers G. 2013 Cold climate specialization: adaptive covariation between metabolic rate and thermoregulation in pregnant vipers. *Physiol. Behav.* **119**, 149–155. (doi:10.1016/j.physbeh.2013.05.041)
- White CR, Blackburn TM, Martin GR, Butler PJ. 2007 Basal metabolic rate of birds is associated with habitat temperature and precipitation, not primary productivity. *Proc. R. Soc. B* **274**, 287–293. (doi:10.1098/rspb.2006.3727)
- Raichlen DA, Gordon AD, Muchlinski MN, Snodgrass JJ. 2010 Causes and significance of variation in mammalian basal metabolism. *J. Comp. Physiol. B* **180**, 301–311. (doi:10.1007/s00360-009-0399-4)
- Chown SL. 2002 Respiratory water loss in insects. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* **133**, 791–804. (doi:10.1016/S1095-6433(02)00200-3)
- Woods HA, Smith JN. 2010 Universal model for water costs of gas exchange by animals and plants. *Proc. Natl Acad. Sci. USA* **107**, 8469–8474. (doi:10.1073/pnas.0905185107)
- Chamaillé-Jammes S, Massot M, Aragon P, Clobert J. 2006 Global warming and positive fitness response in mountain populations of common lizards *Lacerta vivipara*. *Glob. Chang. Biol.* **12**, 392–402. (doi:10.1111/j.1365-2486.2005.01088.x)
- Buckley LB, Kingsolver JG. 2012 Functional and phylogenetic approaches to forecasting species' responses to climate change. *Annu. Rev. Ecol. Syst.* **43**, 205–226. (doi:10.1146/annurev-ecolsys-110411-160516)
- McKechnie AE, Wolf BO. 2010 Climate change increases the likelihood of catastrophic avian mortality events during extreme heat waves. *Biol. Lett.* **6**, 253–256. (doi:10.1098/rsbl.2009.0702)
- Uetz P, Hosek J. 2013 The reptile database. See <http://www.reptile-database.org>.
- Angilletta MJ. 2009 *Thermal adaptation: a theoretical and empirical synthesis*. Oxford, UK: Oxford University Press.
- Bradshaw S. 1997 *Homeostasis in desert reptiles*. Berlin, Germany: Springer Verlag.
- Pyron RA, Burbrink FT, Wiens JJ. 2013 A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol. Biol.* **13**, 93–106. (doi:10.1186/1471-2148-13-93)
- Tieleman BI, Williams JB, Bloomer P. 2003 Adaptation of metabolism and evaporative water loss along an aridity gradient. *Proc. R. Soc. Lond. B* **270**, 207–214. (doi:10.1098/rspb.2002.2205)
- Lighton JRB. 2008 *Measuring metabolic rates: a manual for scientists*. Oxford, UK: Oxford University Press.
- Aleksziuk M. 1971 Temperature-dependent shifts in the metabolism of a cool temperate reptile, *Thamnophis sirtalis parietalis*. *Comp. Biochem. Physiol.* **39**, 495–503. (doi:10.1016/0300-9629(71)90313-6)
- Dmi'el R. 1972 Effect of activity and temperature on metabolism and water loss in snakes. *Am. J. Physiol.* **223**, 510–516.
- McCue MD. 2006 Cost of producing venom in three North American pitviper species. *Copeia* **2006**, 818–825. (doi:10.1643/0045-8511(2006)6[818:COPVIT]2.0.CO;2)
- Lourdais O, Gartner GEA, Brischoux F. 2014 Ambush or active life: foraging mode influences haematocrit levels in snakes. *Biol. J. Linn. Soc.* **111**, 636–645. (doi:10.1111/bij.12223)
- Chown SL, Addo-Bediako A, Gaston KJ. 2003 Physiological diversity: listening to the large-scale signal. *Funct. Ecol.* **17**, 568–572. (doi:10.1046/j.1365-2435.2003.07431.x)
- Therneau TM. 2015 coxme: mixed effects cox models. See <http://cran.r-project.org/package=coxme>.
- Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)

46. Hernández CE, Rodríguez-Serrano E, Avaria-Llautureo J, Inostroza-Michael O, Morales-Pallero B, Boric-Bargetto D, Canales-Aguirre CB, Marquet PA, Meade A. 2013 Using phylogenetic information and the comparative method to evaluate hypotheses in macroecology. *Methods Ecol. Evol.* **4**, 401–415. (doi:10.1111/2041-210X.12033)
47. Killen SS, Glazier DS, Rezende EL, Clark TD, Atkinson D, Willener AST, Halsey LG. 2016 Ecological influences and morphological correlates of resting and maximal metabolic rates across Teleost fish species. *Am. Nat.* **187**, 592–606. (doi:10.1086/685893)
48. Brischox F, Gartner GEA, Garland T, Bonnet X. 2011 Is aquatic life correlated with an increased hematocrit in snakes? *PLoS ONE* **6**, e17077. (doi:10.1371/journal.pone.0017077)
49. Ives AR, Midford PE, Garland T. 2007 Within-species variation and measurement error in phylogenetic comparative methods. *Syst. Biol.* **56**, 252–270. (doi:10.1080/10635150701313830)
50. Wall M, Thompson MB, Shine R. 2013 Does foraging mode affect metabolic responses to feeding? A study of pygopodid lizards. *Curr. Zool.* **59**, 618–625. (doi:10.1093/czoolo/59.5.618)
51. Pough FH. 1980 The advantages of ectothermy for tetrapods. *Am. Nat.* **115**, 92–112. (doi:10.1086/283547)
52. Guderley H, Seebacher F. 2011 Thermal acclimation, mitochondrial capacities and organ metabolic profiles in a reptile (*Alligator mississippiensis*). *J. Comp. Physiol. B* **181**, 53–64. (doi:10.1007/s00360-010-0499-1)
53. Finkel T, Holbrook NJ. 2000 Oxidants, oxidative stress and the biology of aging. *Nature* **408**, 239–247. (doi:10.1038/1781154a0)
54. Cox CL, Cox RM. 2015 Evolutionary shifts in habitat aridity predict evaporative water loss across squamate reptiles. *Evolution* **69**, 2507–2516. (doi:10.1111/evo.12742)
55. Webb W, Szarek S, Lauenroth W, Kinerson R, Smith M. 1978 Primary productivity and water use in native forest, grassland, and desert ecosystems. *Ecology* **59**, 1239–1247. (doi:10.2307/1938237)
56. Muir TJ, Costanzo JP, Lee RE. 2007 Osmotic and metabolic responses to dehydration and urea-loading in a dormant, terrestrially hibernating frog. *J. Comp. Physiol. B* **177**, 917–926. (doi:10.1007/s00360-007-0190-3)
57. Köhler A, Sadowska J, Olszewska J, Trzeciak P, Berger-tal O, Tracy CR. 2011 Staying warm or moist? Operative temperature and thermal preferences of common frogs (*Rana temporaria*) and effects on locomotion. *Herpetol. J.* **21**, 17–26.
58. Davis JR, DeNardo DF. 2009 Water supplementation affects the behavioral and physiological ecology of gila monsters (*Heloderma suspectum*) in the Sonoran Desert. *Physiol. Biochem. Zool.* **82**, 739–748. (doi:10.1086/605933)
59. Dupoué A, Stahlschmidt ZR, Michaud B, Lourdais O. 2015 Physiological state influences evaporative water loss and microclimate preference in the snake *Vipera aspis*. *Physiol. Behav.* **144**, 82–89. (doi:10.1016/j.physbeh.2015.02.042)
60. McCain CM, Colwell RK. 2011 Assessing the threat to montane biodiversity from discordant shifts in temperature and precipitation in a changing climate. *Ecol. Lett.* **14**, 1236–1245. (doi:10.1111/j.1461-0248.2011.01695.x)
61. IPCC. 2014 *Climate change 2014: synthesis report. Contribution of working groups I, II and III to the fifth assessment report of the intergovernmental panel on climate change*. Geneva, Switzerland: IPCC.
62. Lelièvre H, Le Henanff M, Blouin-Demers G, Naulleau G, Lourdais O. 2010 Thermal strategies and energetics in two sympatric colubrid snakes with contrasted exposure. *J. Comp. Physiol. B* **180**, 415–425. (doi:10.1007/s00360-009-0423-8)
63. Basson CH, Levy O, Angilletta MJ, Clusella-Trullas S. 2017 Lizards paid a greater opportunity cost to thermoregulate in a less heterogeneous environment. *Funct. Ecol.* **31**, 856–865. (doi:10.1111/1365-2435.12795)
64. Huey RB, Deutsch CA, Tewksbury JJ, Vitt LJ, Hertz PE, Alvarez Pérez HJ, Garland T. 2009 Why tropical forest lizards are vulnerable to climate warming. *Proc. R. Soc. B* **276**, 1939–1948. (doi:10.1098/rspb.2008.1957)
65. Kearney M, Shine R, Porter WP. 2009 The potential for behavioral thermoregulation to buffer 'cold-blooded' animals against climate warming. *Proc. Natl Acad. Sci. USA* **106**, 3835–3840. (doi:10.1073/pnas.0808913106)