

Population expansion, current and past gene flow in Gould's petrel: implications for conservation

A. Iglesias-Vasquez¹ · B. Gangloff^{1,5} · S. Ruault¹ · C. Ribout¹ · D. Priddel² · N. Carlile² · V. L. Friesen³ · A. Cibois⁴ · V. Bretagnolle¹

Received: 4 November 2015 / Accepted: 13 September 2016 / Published online: 21 September 2016
© Springer Science+Business Media Dordrecht 2016

Abstract Seabird life-history traits such as long generation time, low annual fecundity and delayed sexual maturation make them more prone to population loss and consequently to extinction; petrels are indeed amongst the most threatened birds. Based on coalescence and multiloci inference this study examines the extent of genetic differentiation of a vulnerable New Caledonia (*caledonica*) and Australia (*leucoptera*) subspecies of *Pterodroma leucoptera* (Gould's Petrel), and whether the genetic relationship between them results from the influence of past events like variation in sea level, or is dominated by contemporary gene flow. Sequences of two mitochondrial genes and five nuclear introns were obtained from 86 individuals from both populations. Haplotype networks were used to infer relationships between the haplotypes of both populations. The demographic history of the *P. leucoptera* complex was studied using neutrality tests and Extended Bayesian Skyline Plots. A weak population differentiation was revealed.

The Extended Bayesian Skyline plot suggested a population expansion approximately 80,000 years before present (bp) for *caledonica* and 30,000 years bp for *leucoptera*. The split was dated to 30,000 years bp by means of multilocus inference through *BEAST. Despite genetic similarity of the two taxa, we advocate to consider them as independent units for conservation management, given their strong ecological distinctiveness (foraging distribution, winter distribution, breeding phenology and breeding distribution).

Keywords Phylogeography · Extended Bayesian Skyline plot · Procellariiformes · Seabird · Isolation with Migration · Conservation genetics

Introduction

Anthropogenic impacts on islands, in particular biological invasion and habitat destruction, have been identified as the most important causes of genetic diversity loss, population extinction and, island fauna depletion (Lande 1998; Steadman 2006; Illera et al. 2012). Population extinction is (usually) a long process through which the population first declines in size, then experiences demographic stochasticity and finally becomes extinct (Caughley 1994). Indeed, population genetic theory predicts that when a population declines, genetic diversity is lost as a result of genetic drift and inbreeding depression (Frankham et al. 2002; Allendorf and Luikart 2009). However, disentangling the relative roles of contemporary and historical processes on the overall genetic diversity and population differentiation, and ultimately, survival of populations, is notoriously difficult (Chiucchi and Gibbs 2010; Henriques et al. 2014).

Electronic supplementary material The online version of this article (doi:10.1007/s10592-016-0886-6) contains supplementary material, which is available to authorized users.

✉ A. Iglesias-Vasquez
adriana.iglesias@outlook.fr

¹ Centre d'Etudes Biologiques de Chizé, UMR 7372, CNRS & Université de La Rochelle, 79360 Villiers en Bois, France

² Office of Environment and Heritage,
PO Box 1967, Hurstville, NSW 2220, Australia

³ Department of Biology, Queen's University, Kingston,
ON K7L3N6, Canada

⁴ Department of Mammalogy and Ornithology, Natural History
Museum of Geneva, 1211 Geneva 6, Geneva, Switzerland

⁵ Service de Systématique Moléculaire, UMS 2700 OMSI,
MNHN – CNRS, 75005 Paris, France

Maximum likelihood and Bayesian analyses based on coalescent theory, when applied to DNA (site and length polymorphism), may provide insights into demographic history and genetic structure (Rocha et al. 2011; Brown et al. 2012; Zieliński et al. 2013). Coalescent theory assumes that for any two genetic sequences drawn from a population at random, the probability that they coalesce at a given time is a function of population size at that time (Kingman 1982). Coalescent theory also allows estimation of historic gene flow (Beerli and Felsenstein 1999; Hey and Nielsen 2004). This approach has proved useful in conservation genetics to understand patterns of intraspecific diversity, especially when population size and gene flow are key factors for the long-term survival of conservation units (Hey 2005; Rocha et al. 2011; Zieliński et al. 2013).

Pleistocene climatic oscillations have influenced population distribution, gene flow and genetic variability through changes in habitat availability in various marine organisms (e.g. Pielou 2008; Cheang et al. 2012). In particular, Pleistocene glacial/interglacial cycles seems to have had marked effects on seabirds due to their current extreme characteristics in life-history traits (Gómez-Díaz et al. 2006; Mareile Techow et al. 2009; Techow et al. 2010; Gangloff et al. 2012; Silva et al. 2015) that make them particularly suitable for investigating past and recent population changes using genetic tools. Among seabirds, petrels (Procellariidae) are the most extreme in regard to demographic parameters (as reviewed in Dobson and Jouventin 2007). They usually show large distributions and have very high dispersal abilities (Shaffer et al. 2006). In addition, they breed on remote oceanic islands and exhibit high natal philopatry, often returning to breed within a few meters of their natal nest (Rabouam et al. 1998; Huyvaert and Anderson 2004). High dispersal ability and philopatry act in opposition: strong dispersal capacities should promote gene flow (van Bekkum et al. 2006), while their strong philopatry should promote genetic differentiation (Burg and Croxall 2001; Dearborn et al. 2003).

Within the petrels, the genus *Pterodroma* (c.35 species) alone accounts for 10 % of all seabird species. Gould's Petrel *Pterodroma leucoptera* is a small pelagic gadfly petrel (200–250 g), breeding only on two sites separated by c.1500 km in the Southwest Pacific Ocean (Fig. 1). Each population has been treated as an endemic subspecies because of subtle differences in morphology and coloration (Imber and Jenkins 1981; Bretagnolle and Shirihai 2010). The Australian subspecies, *P. l. leucoptera* (hereafter *leucoptera*), now breeds only on two small islands in New South Wales (Cabbage Tree Island and Boondelbah Island separated by 1.6 km: Carlile et al. 2003), while the New Caledonian subspecies (*P. l. caledonica*, hereafter *caledonica*) is restricted to the south central chain of New Caledonia (Naurois and Rancurel 1978; Bretagnolle and

Shirihai 2010). Several thousands of birds have been ringed but no exchange has ever been documented; however long-term ringing has been done only on Cabbage Tree Island (Priddel et al. 2014). Subspecies also differ in breeding behavior and habitats (*leucoptera* nests in natural cavities among rock scree close to sea level, *caledonica* excavates soil burrows high in the mountains), breeding phenology (a lag of one month; Priddel et al. 2014), foraging zones while breeding, and migration and non-breeding areas (Priddel et al. 2014). However, the genetic relationship between the two taxa remains mostly unknown. Both taxa experienced recent fluctuations in population sizes: *leucoptera* was numerous when discovered in the eighteenth century (Gould 1865), but decreased to fewer than 1500 individuals in 1992 (c. 200 pairs). Classified as *vulnerable* (IUCN 2015), its population recovered to 1000 pairs thanks to a recent restoration program (Priddel and Carlile 1995; 2009). Although no precise information on population trend exists for *caledonica*, its numbers decreased following the introduction of predators (black rats *Rattus rattus*, cats *Felis catus* and pigs *Sus scrofa*) with European settlement approximately 190 years ago (Miller and Mullette 1985; Armstrong 1992; IUCN 2015).

In this study, seven molecular markers were used (two mitochondrial genes and five nuclear introns), (i) to clarify whether taxonomic treatment of the two subspecies is supported by molecular data, (ii) to test whether present genetic structure results from demographic fluctuations (expansions and bottlenecks) due to Pleistocene climatic oscillations, or from more recent changes likely related to anthropogenic pressure; and (iii) to estimate time of divergence of the two populations and their past effective population sizes.

Methods

Sampling, DNA purification, gene amplification, and sequencing

A total of 86 samples of *Pterodroma leucoptera*, consisting of blood (45) and feathers (41), were collected from the two known breeding localities (Fig. 1). Feathers were sampled from live adult birds from the Mt. Dzumac colony, New Caledonia, during the breeding seasons of 2005–2006 and 2008–2009. Similarly, blood samples were collected from Australian birds at the Cabbage Tree Island colony from 2005 to 2008. Blood was collected from veins on the leg or wing using microcapillaries. Samples were stored until analysis in 70 % ethanol at -20°C . Total genomic DNA was isolated from blood samples using Qia-genDNeasy Blood and Tissue extraction kits (Qiagen, Inc., Valencia, CA). Polymerase Chain Reaction (PCR) was

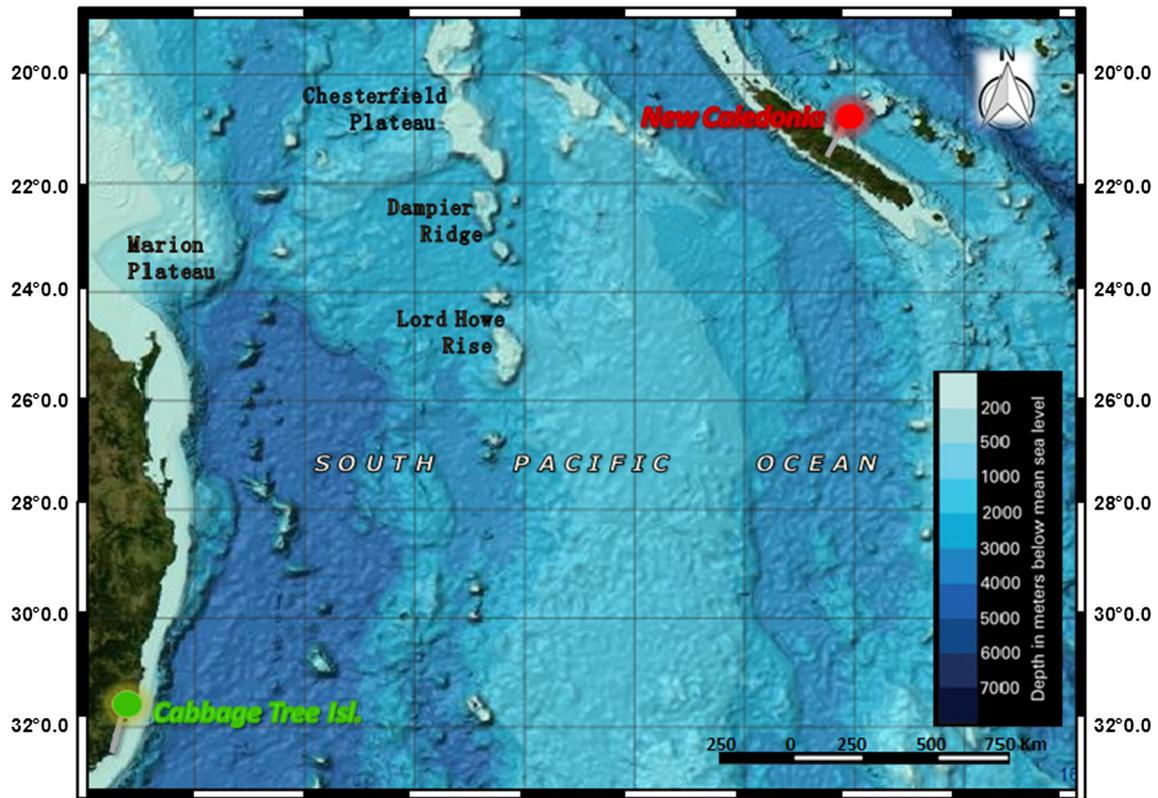


Fig. 1 Sampled breeding localities of *Pterodroma leucoptera*: Dzumac, New Caledonia; Cabbage Tree Island, Australia

used to amplify two mitochondrial DNA (mtDNA) loci and five nuclear introns using previously published primers (Supplementary Table 1). Amplifications were performed in 25 μ l reactions containing 1.5 μ l (30–100 ng/ μ l) of the DNA template, 1X QIAGEN Multiplex PCR Master Mix (Qiagen, UK) and 0.8 μ M of each forward and reverse primer (Supplementary Table 1) PCR products were purified and sequenced using the same PCR primers by Eurofins Scientific (France) and Genome Québec Innovation Centre (McGill University, Montreal, QC, Canada).

Mitochondrial origin and intralocus recombination

Mitochondrial origin of the concatenated mtDNA gene sequences was confirmed by translating DNA sequences to check for stop codons and other potential indications of nuclear origin (Ibarguchi et al. 2006) in BioEdit Sequence alignment Editor v. 7.2.5 (Hall 1999). Sequences were checked visually and aligned with CLUSTAL X (Thompson et al. 1997), implemented in BioEdit Sequence alignment Editor. Recombination in nuclear loci was tested with the four gamete test (Hudson and Kaplan 1985) implemented in DnaSP v.5.10.01 (Librado and Rozas 2009). When the test suggested intralocus recombination, we retained the longest contiguous unrecombined sequence for subsequent analyses. Because the Isolation with Migration

and BEAST analysis requires having known phase for nuclear sequences, we determined the gametic phase using the program PHASE (Stephens et al. 2001) implemented in DnaSP with default parameters and a threshold value of 0.90. Phased nuclear data were then used in all analyses, individuals thus being represented by two sequences.

Evolutionary relationships, genetic diversity and population differentiation

Haplotype frequencies were inferred with DnaSP. Genealogical relationships among haplotypes were reconstructed using a median-joining network and default parameters (Bandelt et al. 1999) in NETWORK v.4.6.0.0 (<http://www.fluxus-engineering.com>). Concatenated mitochondrial and nuclear sequences were used to estimate general statistics of genetic diversity, including number of polymorphic sites (N_p), number of haplotypes (H), number of private haplotypes (PH), haplotype diversity (h ; Nei 1987) and nucleotide diversity (π ; Nei and Tajima 1983), for each subspecies using DnaSP and Arlequin v. 3.5.1 (Excoffier and Lischer 2010). The proportion of genetic variance accounted for within and between subspecies was estimated using an analysis of molecular variance, AMOVA (Excoffier et al. 1992) in Arlequin, and tested for statistical significance using 10,000 permutations. We

calculated pairwise differentiation between subspecies using Φ_{ST} (with Tamura-Nei substitution model), a direct analogue of Wright's F_{st} for nucleotide sequence divergence.

Population size fluctuations through time

Two tests were used to assess if genetic variation deviated from neutral expectations due to either a recent population expansion or selection: Tajima's D (Tajima 1989) and Fu's F_s (Fu 1997) both implemented in Arlequin. Tajima's D values >0 suggest either a recent population bottleneck or balancing selection, while $D < 0$ indicates a population expansion or directional selection. These values are considered significant when $p < 0.05$. Fu's F_s tends to be negative when there is an excess of recent mutations (therefore an overabundance of rare alleles), characteristic of a recent population expansion. Positive Fu's F_s values indicate a deficiency of rare alleles, suggesting a population bottleneck or overdominant selection. Following (Fu 1997) Fu's F_s values are regarded as significant if $p < 0.02$, which corresponds to the conventional significant $p < 0.05$ for Tajima's D. Significant negative Tajima's D and Fu's F_s indices may also indicate selection and genetic hitchhiking associated with selective sweeps. These analyses were conducted for all genes.

A coalescent-based graphical method, the Extended Bayesian Skyline Plot (EBSP) was carried out in BEAST v. 2.1.3 (Drummond and Rambaut 2007) for both populations independently and pooled together to infer potential historical fluctuations in effective population size (N_e). The EBSP allows simultaneous analysis of data from multiple unlinked loci, taking into account their specific mode of inheritance, thus significantly improving the reliability of demographic inferences over single-locus analyses (Heled and Drummond 2008; Shapiro and Ho 2011). EBSP analysis was run using strict clock models as it is considered a good approximation for intra-population level analyses, and it simplifies the coalescent model, helping convergence. Per lineage mutation rate (μ) was calculated by using $\mu = d_{xy}/2T$ (Nei 1987), where d_{xy} stands for inter-lineage divergence and T is the divergence time since two unique lineages diverged (See Supplementary Table 3). For concatenated mtDNA, $d_{xy} = 1.89 \times 10^{-8}$ s/s/y (substitution/site/years) (Weir and Schluter 2008) and for nuclear introns $d_{xy} = 3.6 \times 10^{-9}$ s/s/y (Axelsson et al. 2004); All analyses were run three times to check for convergence with 7×10^7 generations, and sampling every 2×10^3 generations. The first 30 % of the genealogies were discarded as burn-in. Convergence, stationarity, effective sample size for each parameter of interest and the appropriate burn-in were evaluated using the software TRACER v.1.6 (Rambaut et al. 2014). The uncorrected

median population size obtained by EBSP, which is the product $N_e g$ where N_e is the effective population size and g generation time (expressed in the same units as the divergence times) was rescaled and expressed as N_e by dividing by g (Drummond and Bouckaert 2015). A generation time of 20 years as indicated by (Gangloff et al. 2013) was used to rescale the uncorrected population size.

Estimation of gene flow and population connectivity

Patterns of historical and contemporary connectivity between the two subspecies were disentangled through a coalescent inference using the Isolation with Migration under Changing Population Size model (IM) (Hey and Nielsen 2004; Hey 2005). The following parameters were calculated between *leucoptera* and *caledonica*: effective population sizes (N_e *caledonica*; N_e *leucoptera*; N_e *ancestral*), population divergence time (T), the splitting parameter (S) allowing for population size change through time, and migration rates ($M1$, $M2$), where $M1$ indicates the probability of migration per generation migrating from *caledonica* to *leucoptera* forwards in time, and $M2$ indicates migration in the other direction. Concatenated mtDNA and multilocus nuclear DNA were used both jointly and separately to perform the IM analysis. Several preliminary runs were conducted to optimize priors (looking for posterior density curves that rise from zero, peak and then fall to zero within the range for each of the required demographic parameters) following Hey 2009. The final analysis was carried out with the HKY mutation model (Hasegawa et al. 1985) for both nuclear introns and mtDNA, a geometric heating scheme ($g1 = 0.96$ and $g2 = 0.9$), 10 chains, and a chain length of 2 million steps following a 1 million step burn-in. To assess convergence, three separate runs were conducted with different random seed numbers. Effective sample size (ESS) values were monitored to ensure proper mixing of the Markov chain. To convert raw parameter estimates into demographic values, we used the per-locus mutation rates (substitution/year) obtained by multiplying per lineage mutation rate (as used in EBSP) by the number of base pairs of each sequence. The geometric mean of the per-locus mutation rates (μ) was calculated and then used to compute the divergence time by using $T = t/\mu$, expressed in years before present (t , is the maximum likelihood estimate of the parameter T). To calculate effective population size (N_e), we used $N_e = \theta/(4G \mu)$, with a generation time (G) of 20 years (Gangloff et al. 2013). To estimate the population migration rate (M), we used $2N_e M = N_e m/2$, where m stands for the maximum likelihood estimate of the parameter M . The number of migrants from the ancestral to the *leucoptera* population was calculated as $(1-s)\theta a$ (where $(1-s)$ represents the size of the *P. leucoptera* population and θa

stands for the effective size of the ancestral population (Hey 2005).

As the divergence time obtained with IM was unreliable (Supplementary Table 5; Fig. 1), we also used *Beast (Heled and Drummond 2010), implemented in BEAST v.1.6.1 (Drummond and Rambaut 2007), which provides simultaneously phylogenies and divergence time estimates. To root the tree four related species were used as outgroups, two of them, *P. brevipes* and *P. oculata*, were amplified and sequenced in this study while sequences from *P. feae* and *P. madeira* were obtained from GeneBank (Supplementary Table 4). Three runs of 5×10^7 generations, sampling every 1000 generations with a burn in of 2000 trees were performed and then combined in LogCombiner v.1.6.1 (Heled and Drummond 2010). HKY nucleotide substitution model and a strict clock model with an uncorrelated lognormal distribution were used. Per lineage mutation rate of 4.87×10^{-3} s/s/My was used. For the species tree, a Yule process speciation under a coalescent model assuming a constant population over the time period was chosen. Finally, Tracer v.1.6 (Drummond and Rambaut 2007) was used to visualize the results of the runs and to check the effective sample size of each parameter.

Results

We sequenced 1327 base pairs for concatenated CO1 and Cytb (see Table 1 for exact numbers of individuals sequenced for each gene) and 500, 481, 924, 637, and 452 base pairs for the introns PAXIPI, CSDE1, β fibint7, IRF2F1 and TPM1 respectively. Of the five nuclear introns, Bfibint7 was the only one presenting signals of recombination, so we kept the longest possible contiguous unrecombined sequence (918 base pairs) for subsequent analyses. As TPM1 did not present any variation, it was withdrawn from any further analysis. The concatenated mtDNA data did not display insertions or deletions, and after translation, no nonsense or stop codons were found. No ambiguous sites were detected, and true mitochondrial origin of obtained sequences was therefore very likely. DNA was amplified with specific primers designed for *Pterodroma* species (Primmer et al. 2002; Kimball et al. 2009; Jesus et al. 2009; Gangloff et al. 2013) rather than universal primers, which is supposed to reduce the risk of the coamplification of nuclear copies of mitochondrial genes (*numts*) amplification (Sorenson and Quinn 1998; Ibaraguchi et al. 2006).

Table 1 Diversity indices and results of tests for deviations from neutrality for two subspecies of *Pterodroma leucoptera*

Gene	Subspecies	N	Np	Nh	ph	Diversity indices		Neutrality tests	
						Hd [SD ($\times 10^{-2}$)]	π % (SD)	Tajima's D	Fu's F_s
<i>COI-Cytb</i>									
	<i>caledonica</i>	17	13	11	9	0.93 (0.05)	0.16 (0.09)	-1.04	-5.19
	<i>leucoptera</i>	20	7	8	6	0.79 (0.07)	0.063 (0.05)	-1.40	-4.46
<i>PAXIPI</i>									
	<i>caledonica</i>	26	2	4	2	0.28 (7.0)	0.04 (0.06)	-0.64	-2.12
	<i>leucoptera</i>	56	5	7	5	0.21 (5.2)	0.05 (0.06)	-1.41	-5.81
<i>CSDE1</i>									
	<i>caledonica</i>	16	10	8	6	0.68 (5.6)	0.08 (0.06)	-1.62	-3.08
	<i>leucoptera</i>	50	5	5	3	0.38(5.0)	0.03 (0.03)	-1.16	-1.83
<i>βfibint7</i>									
	<i>caledonica</i>	15	19	17	13	0.88 (5.1)	0.09 (0.06)	-1.87	-12.9
	<i>leucoptera</i>	35	7	8	4	0.56 (4.8)	0.031 (0.03)	-1.25	-3.91
<i>IRF2F1</i>									
	<i>caledonica</i>	26	4	5	3	0.68 (6.0)	0.25 (0.18)	0.47	-0.24
	<i>leucoptera</i>	45	3	3	1	0.46 (6.0)	0.19 (0.15)	0.86	1.89
<i>TPM1</i>									
	<i>caledonica</i>	15	0	1	0	0	0		
	<i>leucoptera</i>	18	0	1	0	0	0		

Number of individuals sequenced (N), number of polymorphic sites (Np); number of haplotypes (Nh); number of private haplotypes (ph); haplotype diversity (Hd), nucleotide diversity (π , in %), average number of nucleotide differences between haplotypes (K). Significant values for tests of neutrality: $p < 0.05$ for Tajima's D and $p < 0.02$ for Fu's F_s are shown in bold

Molecular variability

Concatenated mitochondrial data contained 18 polymorphic sites with a total of 17 different haplotypes (13 for *caledonica* and 7 for *leucoptera*; Table 1). Nuclear data exhibited a total of 55 variable sites (7 in PAXIP1, 15 in CSDE1, 26 in β fibint7 and 7 in IRF2F1), leading to 7, 9, 17 and 4 haplotypes respectively (after phasing the nuclear intron sequences). Haplotype and nucleotide diversities tended to be lower in *leucoptera* than in *caledonica* (Table 1).

Population structure and evolutionary relationships

Pairwise single locus estimates of population (subspecies) differentiation for mtDNA and nuDNA were low but significantly different from 0 between the two subspecies taken together ($p < 0.05$ for all global mtDNA and nuDNA except PAXIP1 ($p > 0.05$)) (Supplementary Table 2). Indeed, >90 % of genetic variation was detected at the intra-population level for all loci. No pronounced phylogeographic structure could be detected in the haplotype networks, either for mitochondrial or nuclear markers (Fig. 2). Indeed, all networks were characterized by one or more dominant haplotypes shared by the two subspecies. Similarly, most networks showed star-like topologies, with

one central prevalent haplotype, and other haplotypes having much lower frequencies, suggesting possible past and rapid population expansion (Slatkin and Hudson 1991; Kulikova et al. 2005). *Pterodroma l. caledonica* showed a higher number of private haplotypes despite a lower sample size (Table 1; Fig. 2).

Past population history

Most estimates of Tajima's D and Fu's F_s were negative, suggesting a recent population expansion for either *caledonica* or *leucoptera* (see Table 1 for all values and associated statistics). Reconstruction of the population size history by means of EBSP suggested an expansion episode for both *caledonica* and *leucoptera* (Fig. 3). EBSP further underlined an earlier population increase in *caledonica* (c. 60,000 years bp) than in *leucoptera* (c. 15,000 years bp).

Estimation of gene flow and population connectivity

Migration rates, divergence time and present effective population sizes were obtained with IM analysis only for the two mitochondrial loci since our data failed to find convergence when using all loci. Posterior density curves were acceptable (see Hey 2007) for all but three parameters (Supplementary Table 5; Fig. 2), and therefore, estimates

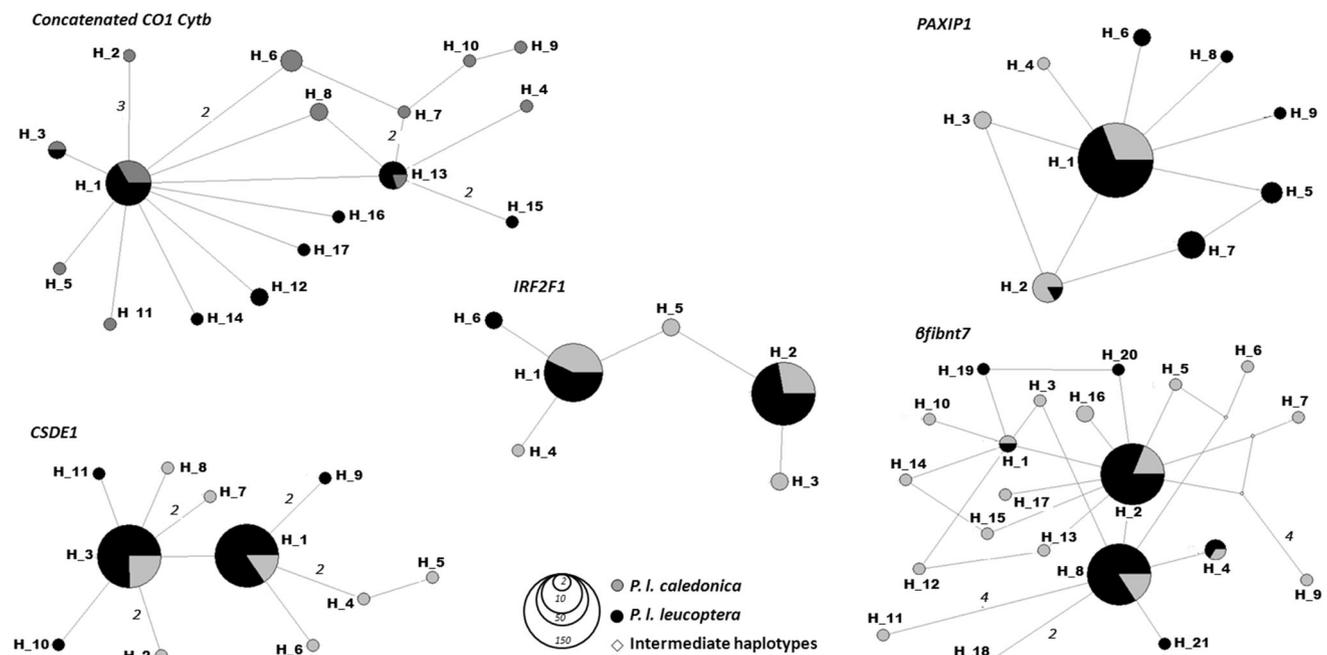
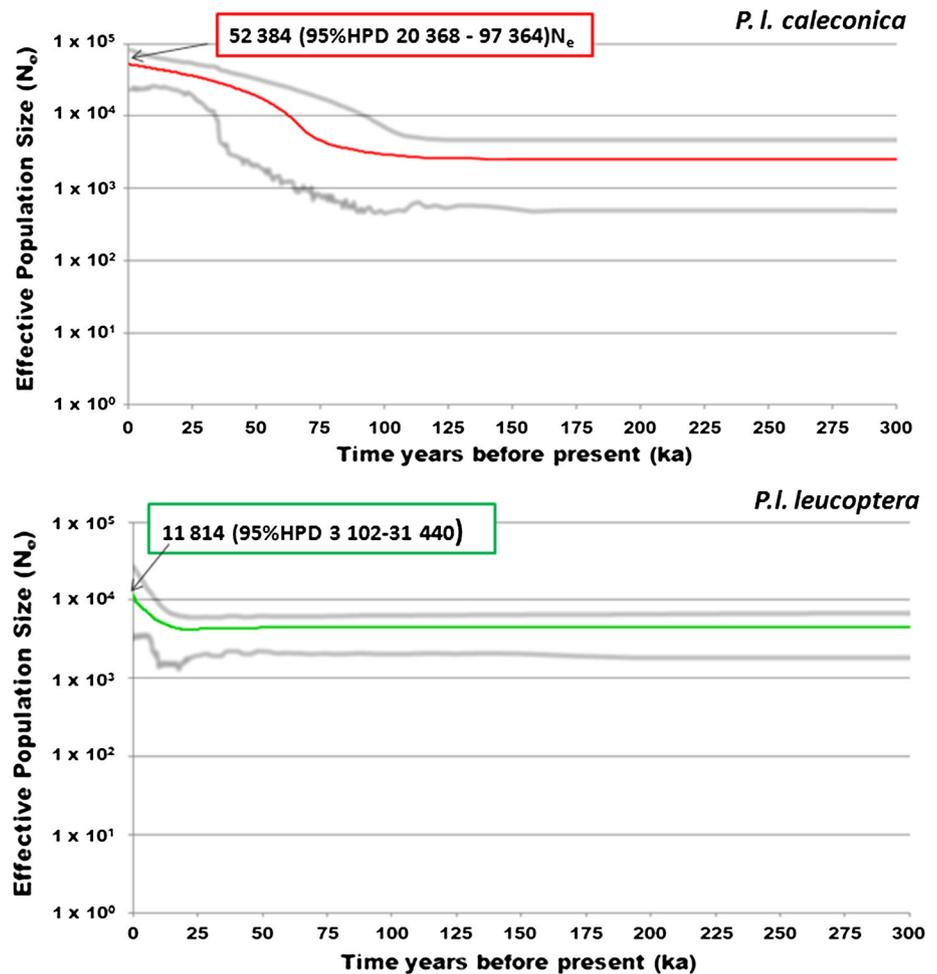


Fig. 2 Haplotype networks obtained with concatenated mtDNA COI and Cytb genes, and five nuclear introns, PAXIP1, β Fibint7, CSDE1 and IRF2F1 within *Pterodroma leucoptera* (TPM1 not shown since there was no variation). Node sizes are proportional to number of individuals found with that haplotype, while color codes refer to

sampling site (see Fig. 1 for sample site locations). Polygons correspond to intermediate (non-sampled) haplotypes. Branch lengths are not scale to improve visualization. Numbers on branches show the number of mutations between alleles. When no number is indicated, only one mutation step occurred

Fig. 3 Extended Bayesian Skyline Plots of *P. l. caledonica* and *P. l. leucoptera* subspecies based on the CO1, Cytb, PAXP1, CSDE1, β Fibint7 and IRF21 data sets. The *green* and *red* lines represent the median of the parameter N_e , proportional to the effective population size in thousands, while the *grey* lines are the 95 % CI



of θa , T (divergence time) and f (number of founder individuals of *leucoptera*) were unreliable and not presented in the results. Estimates of divergence time obtained by BEAST analysis indicated a split event of 30,000 years ago [95 % HPD = 0–0.058] (Fig. 4) between *P. l. caledonica* and *P. l. leucoptera*.

Discussion

The two *P. leucoptera* subspecies show evidence of recent divergence. Results are inconclusive in regard to whether the populations are still connected by gene flow, possibly indicated by high numbers of shared haplotypes, few private haplotypes, and low indices of differentiation. In the absence of population bottlenecks, higher haplotype and nucleotide diversities are expected in ancestral populations compared to more recent ones (Bisconti et al. 2011). Here, we found greater nucleotide and haplotype diversity in *caledonica*, for five out of the six loci under study, suggesting that *caledonica* may be the ancestral form.

A proposed historical reconstruction

Pterodroma l. caledonica was apparently stable in numbers and remained at low population size during the climate cooling period that extended from 130,000 to 70,000 years bp (associated lowering sea levels (Lambeck et al. 2002). From 70,000 years bp, a very smooth increase in population size is however apparent, possibly in relation to further cooling of water temperature and a presumable increase in ocean productivity: today, the two populations feed in southern seas during breeding, and this is especially marked in *caledonica* (Priddel et al. 2014). We thus suggest that colder, more productive waters, moving slowly northward and therefore closer to breeding localities in New Caledonia, may have allowed this population increase. The two lineages apparently split during the Last glacial cycles of the pleistocene epoch (starting 70,000 years bp). Between 70,000 and 20,000 years bp sea level dropped from c. –80 to –120 m, while sea surface temperature decreased sharply (Rohling et al. 1998; Barrows et al. 2007). Several ridges between New Caledonia

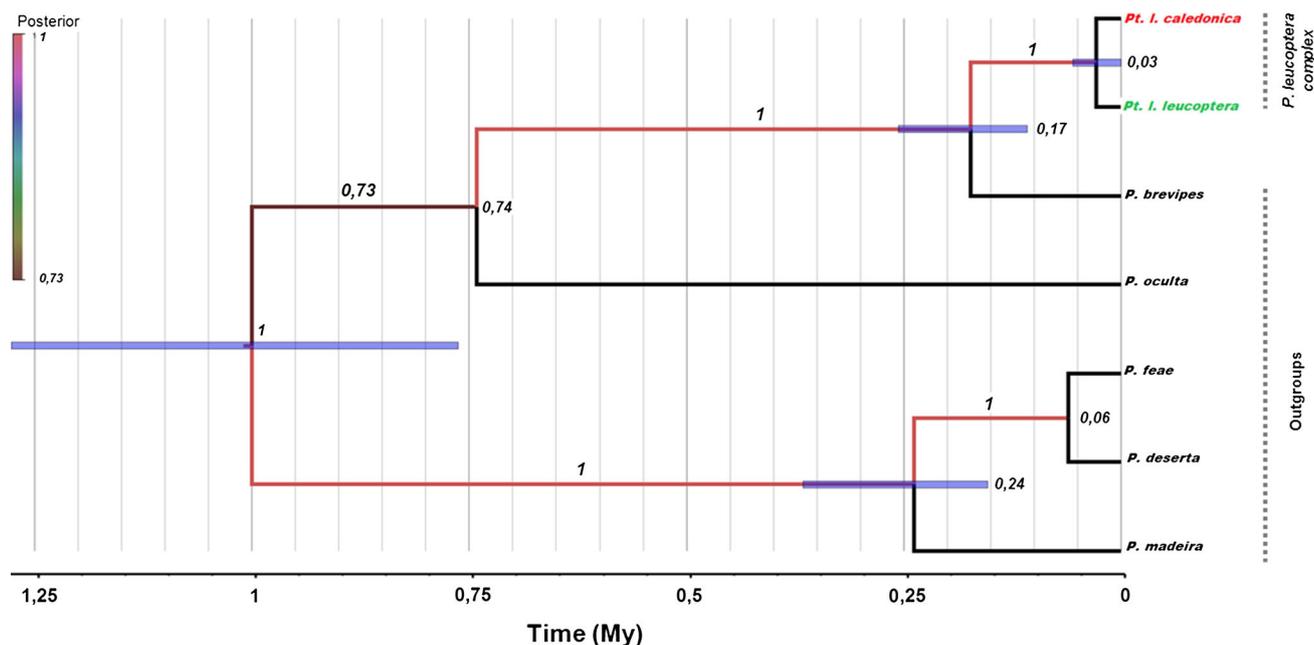


Fig. 4 *Beast tree using two mitochondrial genes (concatenated COI and Cytb) and four nuclear intron (PAXIP1, CSDE1, β fibin7, IRF2F1) showing simultaneously the phylogeny and divergence time of *P. l. caledonica* and *P. l. leucoptera* sub-species under a relaxed

clock assumption. The divergence times expressed in Million years are displayed at each node. Purple bars represent the 95 %HPD interval of the node ages. Numbers on branches represent the posterior probability of each clade

and Australia, currently underwater (e.g. Chesterfield Ridge; Fig. 1), were presumably above sea level, providing many lowland islets. *P. l. caledonica* may have colonized one or several islets halfway between New Caledonia and Australia at this time (see Fig. 1). Divergence hypothetically occurred between the populations, leading to the *leucoptera* lineage. Interestingly, these new islands were lowland islets, a character now found in the breeding habitat of *leucoptera*. Around 18,000 years bp, sea level and sea surface temperatures started to rise. Breeding sites became unavailable for *leucoptera* and the population then presumably colonized or survived only on the few islands off the Australian coast, where they are breeding today.

This proposed historical scenario, however, relies on an accurate estimate of time of divergence as well as population size estimates. However, our IM simulations did not fully converge for parameters such as divergence time and long-term effective size. Indeed similar lack of convergence was already found in other petrel studies (Welch et al. 2011), with the posterior distribution of the curves showing two distinct peaks, hence very wide HPD. Low resolution in parameter estimates with IM can result from sensitivity to inadequate sampling of target populations (Jacobsen and Omland 2012) since parameter estimates will also depend on the level of gene flow between unsampled populations. Besides, low numbers of loci coverage was suggested to promote failure convergence of the IM parameters estimates (Hey 2005; Jacobsen and Omland 2012). In our study we also used multi locus

coalescence-based approaches implemented in EBS and *BEAST rather than concatenated sequences analysis (Degnan and Rosenberg 2006; Kubatko and Degnan 2007), the latter accounting for discrepancies between gene and species trees history when studying closely related species (Zink and Barrowclough 2008). Indeed *BEAST accepts shared polymorphisms originated from incomplete lineage sorting but excludes introgression, treating the latter by conducting separate analyses for each marker (Heled and Drummond 2010; Drummond and Bouckaert 2015; Meyer et al. 2016). To conclude, polyphyly and dissimilarities in gene tree topologies were expected since we analysed closely related species or subspecies, as was appointed by Maddison and Knowles 2006.

The accuracy of estimates of divergence times based on evolutionary substitution rates is also increasingly debated (Garcia-Moreno 2004; Lovette 2004; Ho et al. 2007; Shapiro and Ho 2011), since they are affected by base composition, calibration point sensitivity, generation time, metabolic rates and population size (Lovette 2004; Ho et al. 2005). For instance, Nunn and Stanley 1998 used a calibration point older than 12 Myr, causing saturation of mutations in their cytb sequences and consequently underestimating sequence divergence. It is indeed recommended to use calibrations derived from lineages as close as possible to the organism under study (Lovette 2004; Peterson 2006). For these reasons, we used the interlineage molecular rate derived from (Weir and Schluter 2008) and converted it to a per lineage mutation rate. There was some

discrepancy between our estimate of $\sim 30,000$ years bp divergence time as obtained with BEAST and the fact that IM analysis suggested that one of the taxon started to increase in size earlier, 60,000 years bp in *caledonica*. It should however be noted that both estimates stay within respective confidence intervals, so discrepancy may result from large uncertainty in estimated times.

Contemporary differentiation and taxonomic consequences

These two populations are currently classified as separate subspecies based on slight morphological and color differentiation (Imber and Jenkins 1981). Some researchers even suggested species status (e.g. Onley and Scofield (2007) despite strong overlap in measurements and coloration (Bretagnolle and Shirihai 2010). Our haplotype networks, based on seven loci including both nuclear introns and mitochondrial DNA, revealed very low population differentiation between the two taxa based on Φ_{ST} statistics (mitochondrial $\Phi_{ST} = 0.01$, nuclear Φ_{ST} ranging 0.0049–0.0876) and no phylogeographic differentiation in haplotype networks. Several petrel studies have now used both nuclear and mtDNA, revealing, in general, greater resolution in mitochondrial loci and deeper levels of genetic divergence than for nuclear loci (Silva et al. 2011; Welch et al. 2011; Gangloff et al. 2012; Silva et al. 2015). Unlike other petrels (Ovenden et al. 1991; Friesen et al. 2006), our two taxa showed few locally restricted haplotypes. Conversely, these populations differ in their breeding as well as non-breeding ecology (Priddel et al. 2014). Differences in ecological traits despite similarity at neutral molecular markers and few locally restricted haplotypes may suggest recent divergence with on-going gene flow between *caledonica* and *leucoptera*, with lineages currently unsorted but likely in the process of divergence, rather than a remnant of a large ancestral population.

Conservation implications

Conservation management should target demographically independent populations whose population dynamics depend largely on local birth and death rates rather than on migration (Palsbøll et al. 2007). Conservation genetics can help decide whether subspecies or populations within a species should be managed as separate units (Moritz 2002). Furthermore, independently evolving populations are arguably worth conserving even if they are not different species or taxa (Tobias et al. 2010). The present study provides an interesting case, where neutral molecular markers failed to find strong differences between two populations that are traditionally considered separate conservation units given their different ecological requirements and breeding

habitats. In addition, these populations currently have different potential fates: *leucoptera* shows a very small but increasing breeding population on safe islands, while *caledonica* shows a much bigger but currently declining population on an unsafe island where predation by invasive species occurs. Overall, *Pterodroma leucoptera* is considered vulnerable (Birdlife International 2015). Given differences in ecology and conservation status, and despite the weak neutral genetic and morphological differentiation, we thus recommend that both populations should be protected to preserve the evolutionary potential of these lineages. In particular, as a source of variability able to maintain the genetic diversity of this species, taxon *caledonica* warrants more conservation effort.

Acknowledgments The authors would like to thank T.P. Birt for alignment advice and primer design. We also thank to the Genome Québec Innovation Centre (McGill University Montreal, QC, Canada) for facilitating part of the sequencing of this work. We are also grateful to A. Gotscho and S.P. Tseng for discussions that improved the IM and EBSP analyses. For field work in New Caledonia, VB thanks I. Brun, M. Pandolfi and especially L. Renaudet and P. Villard. We thank A. Welch who helped to improve the manuscript, and P. Duncan for English corrections on a first draft. Thanks also go to the editors and referees for commenting on and correcting the manuscript. We also thank M. Guichard and E. Pante who provided critical assistance with the SuperMachine YMIR- *Université de la Rochelle*. Finally this work was supported by the Région Poitou–Charentes and Deux-Sèvres department, who funded AI-V PHD grant.

Authors contributions VB, DP and NC collected the samples. VB, BG and AI-V conceived the research. SR, CR, BG and AI-V performed the laboratory work. AI-V analyzed the data, with the help of AC and BG. AI-V and VB wrote the manuscript. All the authors read and approved the final manuscript.

References

- Allendorf FW, Luikart G (2009) Conservation and the genetics of populations. Wiley
- Armstrong PH (1992) Human impacts on Australia's Indian ocean tropical island ecosystems: a review. *Environmentalist* 12: 191–206
- Axelsson E, Smith NGC, Sundström H et al (2004) Male-biased mutation rate and divergence in autosomal, z-linked and w-linked introns of chicken and Turkey. *Mol Biol Evol* 21: 1538–1547
- Bandelt HJ, Forster P, Röhl A (1999) Median-joining networks for inferring intraspecific phylogenies. *Mol Biol Evol* 16:37–48
- Barrows TT, Juggins S, De Deckker P et al (2007) Long-term sea surface temperature and climate change in the Australian-New Zealand region. *Paleoceanography* 22:1–17
- Beerli P, Felsenstein J (1999) Maximum-likelihood estimation of migration rates and effective population numbers in two populations using a coalescent approach. *Genetics* 152:763–773
- Birdlife International (2015) Species fact sheet: *Pterodroma leucoptera*. <http://www.birdlife.org/datazone/speciesfactsheet.php?id=3887>
- Bisconti R, Canestrelli D, Colangelo P, Nascetti G (2011) Multiple lines of evidence for demographic and range expansion of a

- temperate species (*Hyla sarda*) during the last glaciation. *Mol Ecol* 20:5313–5327
- Bretagnolle V, Shirihai H (2010) A new taxon of collared petrel *Pterodroma brevipes* from the Banks Islands. *Vanuatu* 130:286–301
- Brown RP, Tejangkura T, El Mouden EH et al (2012) Species delimitation and digit number in a North African skink. *Ecol Evol* 2:2962–2973
- Burg TM, Croxall JP (2001) Global relationships amongst black-browed and grey-headed albatrosses: analysis of population structure using mitochondrial DNA and microsatellites. *Mol Ecol* 10:2647–2660
- Carlile N, Priddel D, Zino F et al (2003) A review of four successful recovery programmes for threatened sub-tropical petrels. *Mar Ornithol* 31:185–192
- Caughley G (1994) Directions in conservation biology. *J Anim Ecol* 63:215–244
- Cheang CC, Tsang LM, Ng WC et al (2012) Phylogeography of the cold-water barnacle *Chthamalus challengerii* in the north-western Pacific: effect of past population expansion and contemporary gene flow. *J Biogeogr* 39:1819–1835
- Chiucchi JE, Gibbs HL (2010) Similarity of contemporary and historical gene flow among highly fragmented populations of an endangered rattlesnake. *Mol Ecol* 19:5345–5358
- Dearborn DC, Anders AD, Schreiber EA et al (2003) Inter-island movements and population differentiation in a pelagic seabird. *Mol Ecol* 12:2835–2843
- Degnan JH, Rosenberg NA (2006) Discordance of species trees with their most likely gene trees. *PLoS Genet* 2:e68
- Dobson FS, Jouventin P (2007) How slow breeding can be selected in seabirds: testing lack's hypothesis. *Proc R Soc B* 274:275–279
- Drummond A, Bouckaert R (2015) Bayesian evolutionary analysis with BEAST. Cambridge University press, Cambridge
- Drummond AJ, Rambaut A (2007) BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol Biol* 7:214
- Excoffier L, Lischer HEL (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol Ecol Resour* 10:564–567
- Excoffier L, Smouse PE, Quattro JM (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131:479–491
- Frankham R, Briscoe DA, Ballou JD (2002) Introduction to conservation genetics. Cambridge University Press
- Friesen VL, González JA, Cruz-Delgado F (2006) Population genetic structure and conservation of the Galápagos petrel (*Pterodroma phaeopygia*). *Conserv Genet* 7:105–115
- Fu YX (1997) Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. *Genetics* 14:915–925
- Gangloff B, Shirihai H, Watling D et al (2012) The complete phylogeny of *Pseudobulweria*, the most endangered seabird genus: systematics, species status and conservation implications. *Conserv Genet* 13:39–52
- Gangloff B, Zino F, Shirihai H et al (2013) The evolution of north-east Atlantic gadfly petrels using statistical phylogeography. *Mol Ecol* 22:495–507
- García-Moreno J (2004) Is there a universal molecular clock for birds? *J Avian Biol* 35:465–468
- Gómez-Díaz E, González-Solís J, Peinado MA, Page RDM (2006) Phylogeography of the Calonectris shearwaters using molecular and morphometric data. *Mol Phylogenet Evol* 41:322–332
- Gould J (1865) Handbook to the birds of Australia, vol Vol 1. Cambridge University, London
- Hall T (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser* 41:95–98
- Hasegawa M, Kishino H, Yano TA (1985) Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J Mol Evol* 22:160–174
- Heled J, Drummond AJ (2008) Bayesian inference of population size history from multiple loci. *BMC Evol Biol* 15:1–15
- Heled J, Drummond AJ (2010) Bayesian inference of species trees from multilocus data. *Mol Biol Evol* 27:570–580
- Henriques R, Potts WM, Santos CV et al (2014) Population connectivity and phylogeography of a coastal fish, *Atractoscion aequidens* (Sciaenidae), across the Benguela current region: evidence of an ancient vicariant event. *PLoS One* 9:e87907
- Hey J (2005) On the number of new world founders: a population genetic portrait of the peopling of the Americas. *PLoS Biol* 3:0965–0975
- Hey J. (2007). Introduction to the IM and IMA computer programs. <http://lifesci.rutgers.edu/~hey/Programsand-Data/Programs/IM>
- Hey J (2009) Using IM documentation. <https://bio.cst.temple.edu/~hey/software/software.htm>
- Hey J, Nielsen R (2004) Multilocus methods for estimating population sizes, migration rates and divergence time, with applications to the divergence of *Drosophila pseudoobscura* and *D. persimilis*. *Genetics* 167:747–760
- Ho SYW, Phillips MJ, Cooper A, Drummond AJ (2005) Time dependency of molecular rate estimates and systematic overestimation of recent divergence times. *Mol Biol Evol* 22:1561–1568
- Ho SYW, Heupink TH, Rambaut A, Shapiro B (2007) Bayesian estimation of sequence damage in ancient DNA. *Mol Biol Evol* 24:1416–1422
- Hudson RR, Kaplan NL (1985) Statistical properties of the number of recombination events in the history of a sample of DNA sequences. *Genetics* 111:147–164
- Huyvaert K, Anderson D (2004) Limited dispersal by Nazca boobies *Sula grantii*. *J Avian Biol* 35:46–53
- Ibarguchi G, Friesen VL, Loughheed SC (2006) Defeating numts: Semi-pure mitochondrial DNA from eggs and simple purification methods for field-collected wildlife tissues. *Genome* 49(11):1438–1450
- Illera JC, Rando JC, Richardson DS, Emerson BC (2012) Age, origins and extinctions of the avifauna of Macaronesia: a synthesis of phylogenetic and fossil information. *Quat Sci Rev* 50:14–22
- Imber MJ, Jenkins JAF (1981) The New Caledonian Petrel. Department of Internal Affairs
- IUCN (2015) The IUCN Red List of Threatened Species. Version 2015.1. <http://www.iucnredlist.org>. Downloaded on 1 Feb 2015
- Jacobsen F, Omland KE (2012) Extensive introgressive hybridization within the northern oriole group (genus *icterus*) revealed by three-species isolation with migration analysis. *Ecol Evol* 2:2413–2429
- Jesus J, Menezes D, Gomes S et al (2009) Phylogenetic relationships of gadfly petrels *Pterodroma* spp. from the Northeastern Atlantic Ocean: molecular evidence for specific status of Bugio and Cape Verde petrels and implications for conservation. *Bird Conserv Int* 19(3):199–214
- Kimball RT, Braun EL, Barker FK et al (2009) A well-tested set of primers to amplify regions spread across the avian genome. *Mol Phylogenet Evol* 50:654–660
- Kingman JFC (1982) The coalescent. *Stoch Process Appl* 13:235–248
- Kubatko LS, Degnan JH (2007) Inconsistency of phylogenetic estimates from concatenated data under coalescence. *Syst Biol* 56:17–24

- Kulikova IV, Drovetski SV, Gibson DD et al (2005) Phylogeography of the Mallard (*Anas platyrhynchos*): hybridization, dispersal, and lineage sorting contribute to complex geographic structure. *Auk* 122:949–965
- Lambeck K, Esat TM, Potter E-K (2002) Links between climate and sea levels for the past three million years. *Nature* 419:199–206
- Lande R (1998) Anthropogenic, ecological and genetic factors in extinction and conservation. *Res. Popul. Ecol. (Kyoto)* 40: 259–269
- Librado P, Rozas J (2009) DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25:1451–1452
- Lovette I (2004) Mitochondrial dating and mixed support for the “2% rule” in birds. *Auk* 121:1–6
- Maddison WP, Knowles LL (2006) Inferring phylogeny despite incomplete lineage sorting. *Syst Biol* 55:21–30
- Mareile Techow N, Ryan P, O’Ryan C (2009) Phylogeography and taxonomy of white-chinned and spectacled petrels. *Mol Phylogenet Evol* 52:25–33
- Meyer BS, Matschiner M, Salzburger W (2016) Disentangling incomplete lineage sorting and introgression to refine species-tree estimates for Lake Tanganyika cichlid fishes. *BioRxiv*. doi:10.1093/sysbio/syw069
- Miller B, Mullette KJ (1985) Rehabilitation of an endangered Australian bird: the Lord Howe Island woodhen *Tricholimnas sylvestris* (Sclater). *Biol Conserv* 34:55–95
- Moritz C (2002) Strategies to protect biological diversity and the evolutionary. *Syst Biol* 51:238–254
- Naurois R, Rancurel P (1978) Observations nouvelles sur les Laridae reproducteurs en Nouvelle-Calédonie. *Sci Nat* 287:495–498
- Nei M (1987) *Molecular evolutionary genetics*. Columbia University Press, New York
- Nei M, Tajima F (1983) Maximum likelihood estimation of the number of nucleotide substitutions from restriction sites data. *Genetics* 105:207–217
- Nunn G, Stanley S (1998) Corrigendum for: body size effects and rates of cytochrome-b evolution in tube-nosed seabirds. *Mol Biol Evol* 17:1774
- Onley D, Scofield P (2007) *Field Guide to the albatrosses, petrels and shearwaters of the World* Christopher Helm Publishers Ltd
- Ovenden J, Wust-Saucy A, Bywater R et al (1991) Genetic evidence for philopatry in a colonially nesting seabird, the Fairy Prion (*Pachyptila turtur*). *Auk* 108:688–694
- Palsbøll PJ, Bérubé M, Allendorf FW (2007) Identification of management units using population genetic data. *Trends Ecol Evol* 22:11–16
- Peterson AT (2006) Application of molecular clocks in ornithology revisited. *J Avian Biol* 37:541–544
- Pielou E (2008) *After the ice age: the return of life to glaciated North America*. University of Chicago Press, Chicago
- Priddel D, Carlile N (1995) Mortality of adult Gould’s petrels *Pterodroma leucoptera leucoptera* at the nesting site on Cabbage Tree Island, New South Wales. *Emu* 95:259–264
- Priddel D, Carlile N (2009) Key elements in achieving a successful recovery programme: a discussion illustrated by the Gould’s Petrel case study. *Ecol Manag Restor* 10:97–102
- Priddel D, Carlile N, Portelli D et al (2014) Pelagic distribution of Gould’s petrel (*Pterodroma leucoptera*): linking shipboard and onshore observations with remote-tracking data. *Emu* 114(4): 360–370
- Primmer CR, Borge T, Lindell J, Sætre GP (2002) Single-nucleotide polymorphism characterization in species with limited available sequence information: high nucleotide diversity revealed in the avian genome. *Mol Ecol* 11:603–612
- Rabouam C, Thibault J, Bretagnolle V (1998) Natal philopatry and close inbreeding in Cory’s shearwater (*Calonectris diomedea*). *Auk* 115:483–486
- Rambaut A, Suchard M, Xie D, Drummond A (2014) *Tracer v1.6*. Available from <http://beast.bio.ed.ac.uk/Tracer>
- Rocha S, Harris DJ, Posada D (2011) Cryptic diversity within the endemic prehensile-tailed gecko *Urocytyledon inexpectata* across the Seychelles Islands: patterns of phylogeographical structure and isolation at the multilocus level. *Biol J Linn Soc* 104:177–191
- Rohling EJ, Fenton M, Jorinsen FJ et al (1998) Magnitudes of sea-level lowstands of the past 500,000 years. *Nature* 394(6689): 162–165
- Shaffer SA, Tremblay Y, Weimerskirch H et al (2006) Migratory shearwaters integrate oceanic resources across the Pacific Ocean in an endless summer. *Proc Natl Acad Sci USA* 103:12799–12802
- Shapiro B, Ho S (2011) Skyline-plot methods for estimating demographic history from nucleotide sequences. *Mol Ecol Resour* 11(3):423–434
- Silva MC, Duarte MA, Coelho MM (2011) Anonymous nuclear loci in the white-faced storm-petrel *Pelagodroma marina* and their applicability to other Procellariiform seabirds. *J Hered* 102:362–365
- Silva MC, Matias R, Wanless RM et al (2015) Understanding the mechanisms of anti-tropical divergence in the seabird white-faced Storm-petrel (Procellariiformes: *pelagodroma marina*) using a multi-locus approach. *Mol Ecol* 24:3122–3137
- Slatkin M, Hudson RR (1991) Pairwise comparisons of mitochondrial DNA sequences in stable and exponentially growing populations. *Genetics* 129:555–562
- Sorenson MD, Quinn TW (1998) Numts: a challenge for avian systematics and population biology. *Auk* 115:214–221
- Steadman DW (2006) *Extinction and biogeography of tropical Pacific birds*. University of Chicago Press
- Stephens M, Smith NJ, Donnelly P (2001) A new statistical method for haplotype reconstruction from population data. *Am J Hum Genet* 68:978–989
- Tajima F (1989) The effect of change in population size on DNA polymorphism. *Genetics* 123:597–601
- Techow NM, Ryan PG, O’Ryan C (2009) Phylogeography and taxonomy of white-chinned and Spectacled Petrels. *Mol Phylogenet Evol* 52:25–33
- Techow NMSM, O’Ryan C, Phillips RA et al (2010) Speciation and phylogeography of giant petrels *Macronectes*. *Mol Phylogenet Evol* 54:472–487
- Thompson JD, Gibson TJ, Plewniak F et al (1997) The CLUSTAL X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* 25:4876–4882
- Tobias JA, Seddon N, Spottiswoode CN et al (2010) Quantitative criteria for species delimitation. *Ibis* 152(4):724–746
- van Bekkum M, Sagar PM, Stahl J-C, Chambers GK (2006) Natal philopatry does not lead to population genetic differentiation in Buller’s albatross (*Thalassarche bulleri bulleri*). *Mol Ecol* 15:73–79
- Weir JT, Schluter D (2008) Calibrating the avian molecular clock. *Mol Ecol* 17:2321–2328
- Welch AJ, Yoshida AA, Fleischer RC (2011) Mitochondrial and nuclear DNA sequences reveal recent divergence in morphologically indistinguishable petrels. *Mol Ecol* 20:1364–1377
- Zieliński P, Nadachowska-Brzyska K, Wielstra B et al (2013) No evidence for nuclear introgression despite complete mtDNA replacement in the Carpathian newt (*Lissotriton montandoni*). *Mol Ecol* 22:1884–1903
- Zink RM, Barrowclough GF (2008) Mitochondrial DNA under siege in avian phylogeography. *Mol Ecol* 17:2107–2121