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Colouration matters in dull toads: ultraviolet adornment for ladies and agrochemicals fading effects

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Abstract

Integument colouration can influence many aspects of fitness, and is under strong sexual selection. Amphibians often express sexual dichromatism, and ultra-violet (UV) colouration is usually biased toward males as a sexual signal. As an honest signal, colouration is related to several individual traits, but can also be related to environmental factors such as anthropogenic pollutants, to which amphibians are highly sensitive. In this study, we investigated sexual dichromatism and UV reflectance covering a large visual spectrum (wavelength ranging from 300 to 700 nm) on different body areas (throat, ventral and dorsal areas), in a widespread amphibian species, the spiny toad (*Bufo spinosus*). Then, we tested the impact of chronic exposure to two widespread herbicides (glyphosate's primary metabolite [AMPA] and Nicosulfuron) on their colouration. We found a strong but unexpected sexual dichromatism with females reflecting more in the UV spectrum (throat and ventral area) than males, suggesting these body parts might be critical in intra-specific signalling. Females with higher ventral UV reflectance were in better body condition, suggesting an honest signal role of UV reflectance which could influence male choice. Throat colouration was further differentially influenced by agrochemicals according to sexes. In AMPA-exposed males, throat was more saturated in yellow-orange than in control males, and Nicosulfuron exposure decreased the throat's reflectance hue in females, which can bear consequences on mate attractiveness. Future studies need to investigate the underlying mechanisms that are altered by agrochemical exposure.

 $\textbf{Keywords} \ \ Amphibians \cdot Honest \ signal \cdot AMPA \cdot Nicosulfuron \cdot Sexual \ dichromatism$

Introduction

Integument colouration is a major interface between individuals and their environment, as it influences many aspects of their biology including anti-predator defences through crypsis (Caro 2005), thermoregulation (Stuart-Fox and Moussalli 2009), regulation of physiological processes

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(Caro 2005), immunity (Lifshitz and St Clair 2016), parasitic exploitation (Côte et al. 2018), UV protection (Jablonski 1998; Clusella Trullas et al. 2007) and social and sexual signaling in both males and females (Bradbury and Vehrencamp 1998; Caro 2005; Delhey et al. 2007; Stuart-Fox et al. 2007; Martín and López 2009; Clutton-Brock 2009; Secondi et al. 2012; Olsson et al. 2013; Heath et al. 2013; Endler and Mappes 2017). Given the role of body colouration in such fitness-related functions, the mosaic of colours displayed by an animal can therefore convey information about individual quality and is considered to be under strong sexual selection (Hews and Moore 1995; Bradbury and Vehrencamp 1998; Ptacek 2000; Martín and López 2009; Svensson and Wong 2011; Sever and Staub 2011; Heath et al. 2013; Blévin et al. 2014; Endler and Mappes 2017; Weaver et al. 2018).

From vibrant to substrate-matching colours, amphibians show a strong diversity of colourations and contrasting patterns (Rudh and Qvarnström 2013). Some amphibian species express sexual dichromatism (i.e. colour difference between the two sexes); (Lifshitz and St Clair 2016), with adult males



being more brightly coloured than adult females (Bell and Zamudio 2012). Colour signals can facilitate gender recognition, influence courtship behaviour and mate-choice, even in nocturnal species (Gomez et al. 2009; Sztatecsny et al. 2010; Secondi et al. 2012). A chromatophore unit (i.e. pigment containing cells) of amphibians can combine properties of different chromatophores to achieve rapid colour changes (Bagnara et al. 1968; Rudh and Qvarnström 2013). Across 178 anuran species, males were shown to use rapid colour change (dynamic dichromatism) to mediate socio-sexual interactions during the breeding season (Bell et al. 2017). Dynamic dichromatism often precedes explosive breeding, and could facilitate sexual recognition by helping males distinguish each other from females (Doucet and Mennill 2010; Sztatecsny et al. 2012; Stückler et al. 2022).

Although sexual dichromatism may not be the most prevalent secondary sexual trait in anurans (Duellman and Trueb 1986; Bell and Zamudio 2012; Bell et al. 2017), features non-visible to human eye, such as ultra-violet (UV) signalling, may be much more common than previously thought. Various taxa, including amphibians, have retinal photopigments that allow for UV vision (Jacobs 1992). Most bright integument colour patterns reflect light maximally in the UV portion of the spectrum, constituting UV colour signals (Burkhardt and Finger 1991; Fleishman et al. 1993; Deutschlander and Phillips 1995; Stoehr and McGraw 2001; Hunt et al. 2001; Pérez i de Lanuza and Font 2007; Martin et al. 2013). This pattern is mainly expressed by males (Stoehr and McGraw 2001; Pérez i de Lanuza and Font 2007; Martin et al. 2013) and to a lesser extent by females (Martin et al. 2013). In ectothermic species, iridophores are chromatophores responsible for these bright reflecting colours, as they contain reflecting platelets (Rudh and Qvarnström 2013). As observed in reptiles (Pérez i de Lanuza and Font 2007) and caudates (Secondi et al. 2012), UV reflectance could increase sexual dichromatism and male conspicuousness in anurans. Additionally, skin colour is mostly influenced by pigments contained within specialized groups of cells in the upper epidermal layer of skin. Melanins are the most prevalent pigments, producing many yellow-brownish (pheomelanin) and grey-black (eumelanin) colours (McGraw 2005). Melanogenesis is controlled genetically and vertebrates synthesize melanins from internal ressources (Lin and Fisher 2007). By contrast to melanin, carotenoid pigments are acquired through the diet (Schiedt 1989). They produce many yellow, orange and red patches, which are central to ornamentation (Blount and McGraw 2008).

In amphibians, colouration has been related to several individual traits such as body size, body condition or hormone levels (Nilsson Sköld et al. 2013; Höbel et al. 2022; Barzaghi et al. 2022), and can also vary with environmental factors. For instance, temperature, habitat (e.g. site elevation,

productivity, background colour) or food availability have been related to colour variations in amphibians (Norris and Lowe 1964; Sztatecsny et al. 2010; Mack and Beaty 2021; Barzaghi et al. 2022; Mirč et al. 2023). Since colouration is thought to be an adaptive trait (Rudh and Qvarnström 2013), exogenous factors leading to colour modification could affect individual fitness.

The ubiquity and plasticity of integument colouration make it a powerful indicator of the competing costs of environmental stressors, such as anthropogenic pollutants (Lifshitz and St Clair 2016), among which pesticides are dominant in agricultural habitats where several amphibians persist (McConnell et al. 1998; LeNoir et al. 1999; Rashid et al. 2010; Guillot et al. 2016). Indeed, pesticides primarily cause substantial damages to organisms by producing free radicals that overwhelm the antioxidant system (Galván and Alonso-Alvarez 2009; Cheron et al. 2022), and the major groups of animal pigments exhibit antioxidant activity (McGraw 2005) suggesting a trade-off between the expression of colouration and resistance to pollutants (Arellano-Aguilar and Macías Garcia 2008). Pollutants have been related to enhanced traits coloured by black melanin (Lifshitz and St Clair 2016; Goiran et al. 2017), reduced carotenoid colouration (Baatrup and Junge 2001; Alonso-Alvarez and Galván 2011; Shenoy 2012), as well as modified hue and saturation of individuals (Larramendy 2017; Ujhegyi and Bókony 2020). These impairments could result from negative effects of pollutants on gonad function (McCoy et al. 2008; Hayes et al. 2010), leading to sex steroid disruption (Trudeau et al. 2020). Testosterone, for instance, can affect chromatophore expression and xanthophores dispersion in amphibian (Richards 1982; Tang et al. 2014). Despite their potential higher susceptibility to contaminants, due to a lack of protective epidermal structures and permeable skin (Wells 2007), only a handful of studies have been conducted on the effects of pollutants on colouration in amphibians (Larramendy 2017; Ujhegyi and Bókony 2020). Agrochemicals are mostly sprayed in spring, when many amphibians' species from temperate areas reproduce (Wells 2007; Berger et al. 2013; Lenhardt et al. 2015), and animals may be exposed to these compounds through inhalation, skin permeability, and consumption of contaminated food as well as water.

In this study, we investigated skin colour variations and UV signalling according to sex and considering different body parts (throat, ventral and dorsal areas) in a widespread amphibian species, the spiny toad (*Bufo spinosus*), characterized by an overall dull colouration as compared to other brightly coloured anurans (Rudh and Qvarnström 2013). We also tested the effects of exposure to two heavily used herbicides (glyphosate's primary metabolite, aminomethylphosphonic acid [AMPA] and Nicosulfuron) to which freeranging amphibians are exposed in agricultural areas (Brühl et al. 2013; Berger et al. 2013; Lenhardt et al. 2015; Adams



et al. 2021). To do so, we captured adults of both sexes in Western France out of the breeding period and examined their throat, ventral, and dorsal colouration based on spectrophotometric measures. In amphibians, ventral and throat colours are lighter than the dorsal area and show clear oxyhaemoglobin absorption peaks (Norris and Lowe 1964). This species expresses a strong sexual dimorphism (Speybroeck et al. 2018), and sexual dichromatism has been described in a closely related species, Bufo bufo, with males being yellower-greener (less red) and brighter than females (Ujhegyi and Bókony 2020). We thus expect that colouration will vary with sex in Bufo spinosus, and will be related to individual traits as an honest signal of quality, as demonstrated in males (Martín and López 2009; Heath et al. 2013). In addition, given the effects of agrochemicals on gonad function and hormone concentrations (McCoy et al. 2008; Trudeau et al. 2020), we expect that contaminant exposure may as well affect individual's colouration, increasing black melanin pigmentation (Lifshitz and St Clair 2016; Goiran et al. 2017), reducing carotenoid colouration (Baatrup and Junge 2001; Alonso-Alvarez and Galván 2011; Shenoy 2012), and modifying hue and saturation (Larramendy 2017; Ujhegyi and Bókony 2020).

Material and methods

Study species

The spiny toad is the largest toad species originating from western Europe (Speybroeck et al. 2018). The species reproduces in ponds during late winter, but is found migrating both for aestivation and hibernation, respectively in spring and autumn. Males and females are sexually dimorphic, females being larger than males, and males presenting nuptial pads on their fingers, allowing them to clasp females during reproduction (Speybroeck et al. 2018).

Animal care and housing

Between 21-Sep-2020 and 27-Oct-2020, 81 free-ranging spiny toads (32 males and 49 females) were captured by hand on the roadside to the south of the Deux-Sèvres department (western France) and brought back to the laboratory. Toads were housed in a thermally controlled (17 °C) room under natural photoperiod (12:12 h cycle), in individual plastic boxes ($80 \times 40 \times 16$ cm) lined with paper towels, with a shelter (halved PVC tubes) and a petri dish for water. Twice a week, in quantities adapted to their size, toads were fed crickets, meal worms and earth worms dusted with a mixture of CaCO₃ and multivitamin powder (Repti Calcium, Zoo Med Laboratories, Inc). Once a week, boxes were cleaned and paper towels were replaced.

Agrochemical exposure

AMPA (aminomethylphosphonic acid) is the primary metabolite of glyphosate, and is one of the main contaminants detected in surface waters worldwide (Grandcoin et al. 2017). New herbicides molecules are also regularly developed, such as Sulfonylurea Herbicides (e.g. Sulfosulfuron, Rimsulfuron, and Nicosulfuron) which are persistent in aquatic environments such as ponds (Cessna et al. 2015). We exposed toads either to AMPA or one Sulfonylurea herbicide (Nicosulfuron) and started a chronic exposition at environmentally relevant concentrations from 15-Apr-2021 to 01-Jul-2021. Individuals were exposed to one of the three treatments: Control (11 males, 20 females), AMPA (10 males, 17 females), Nicosulfuron (11 males, 12 females). The individuals in the three experimental groups did not differ in size, either considering males (linear model: Sum Sq = 12.352, F-value = 0.745, p-value = 0.484, Control males: $58.65 \text{ mm} \pm 2.08 \text{ SE}$, AMPA males: $59.93 \text{ mm} \pm 3.87$ SE, Nicosulfuron males: $59.98 \text{ mm} \pm 2.49 \text{ SE}$) or females (linear model: Sum Sq = 231.43, F-value = 1.703, p-value = 0.193, Control females: $72.81 \text{ mm} \pm 8.33 \text{ SE}$, AMPA females: $73.81 \text{ mm} \pm 8.66 \text{ SE}$, Nicosulfuron females: $78.22 \text{ mm} \pm 7.43 \text{ SE}$). AMPA was administered by dissolution in drinking water (dechlorinated tap water) at 4 μ g L⁻¹ (crystalline powders, 99% purity, ACROS ORGANICSTM), and Nicosulfuron was administered by dissolution in drinking water (dechlorinated tap water) at 1 μ g L⁻¹ (crystalline powders, 99% purity, ACROS ORGANICSTM). These concentrations correspond to the concentrations of these pollutants found in agricultural environments (Tartu et al. 2022, and data from Agence de l'Eau Loire-Bretagne). Concentrations measured in 4 samples of drinking water per treatment validated that the actual concentrations were close to the nominal concentrations (Qualyse lab, La Rochelle, France). Toads were exposed to their treatment through drinking water (petri dish) twice a week.

Colouration and morphometrics

On 01-Jul-2021, after a two-and-a-half-month exposure, we obtained reflectance spectra from the throat, the ventral, and the dorsal area of each individual (two replicates; reflectance spectrums are given in Appendix A). These measurements were done using a USB-2000 spectrophotometer (Badiane et al. 2020; Kawamoto et al. 2021). The probe was hand-held over each area, approximately perpendicular to the patch surface. We then processed spectral data in R v.4.0.5 (R Core Team 2019) using the package pavo (Maia et al. 2013). We cropped each spectrum between 300 and 700 nm, smoothed them using a loess smooth span of 0.2, and averaged the two replicates recorded for each body region. For each body part we extracted hue, UV-saturation, UV-luminance, total



brightness (B1 300–700). We then calculated yellow-orange saturation (B1 575–700/B1 300–700), violet-blue saturation (B1 400–515/B1 300–700) and green saturation (B1 495–570/B1 300–700) (Badiane et al. 2020). All individuals were weighed (electronic balance: \pm 0.1 g) and their snoutvent length (SVL) was measured with an electronic calliper (\pm 0.01 mm) on the same day. We then calculated a body condition index using residual scores from the linear regression between log (body size) and log (body mass). At the end of the experiment, individuals were all released at their site of capture.

Statistical analyses

First, we ran a principal component analysis (PCA) on the different fractions of reflectance with the ade4 package (Dray and Dufour 2007). We generated principal components (PCs) for further analyses from the first and second axis of the PCA (projected inertia > 70%) that we used as a global index of colouration. Correlations between these PCs and the different fractions of reflectance included in the PCA are presented in Appendix B. Second, we used linear models (LMs) to test 1) the effects of sex and 2) the effects of body condition on reflectance (PC 1 and PC 2 for each body part). Sex, body condition and their interaction were defined as explanatory variables and reflectance PC1 and PC2 for each body part as response variables. We tested these relationships in control individuals only, to test for this effect without the effect of contaminant exposure. Third, we tested the effects of the treatment (Control, AMPA, or Nicosulfuron) on reflectance (PC 1 and PC 2 for each body part) in each sex separately, as PC1 and PC2 values were highly different between males and females (LM models: all p-values < 0.029, except for the comparisons in PCA 2 throat coloration between sexes [p-value = 0.159], see also Fig. 1). Finally, we tested the effects of body condition, sex and their interaction on reflectance in AMPA and Nicosulfuron exposed individuals only. These variables were selected by backward stepwise selection, and only the last retained variables are presented in the final models. All analyses were performed with R v.4.0.5 (R Core Team 2019).

Results

Reflectance sexual dimorphism in control individuals

In control individuals, we observed a significant sexual dichromatism in reflectance of different body parts (Fig. 1, Table 1). Females reflected in the UV spectrum whereas males rather reflected in the yellow-orange and green spectra (Fig. 1). Ventral PC1 varied according to

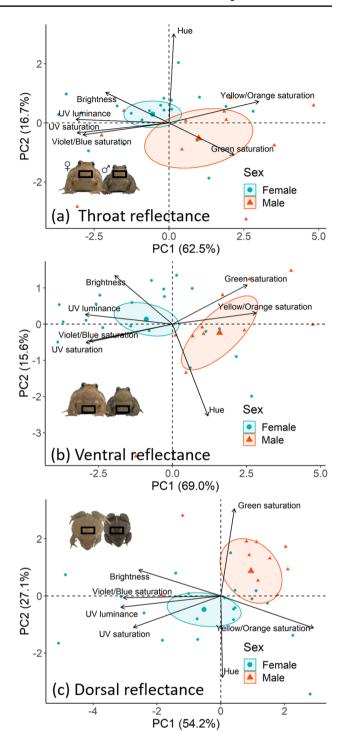


Fig. 1 Ordination plot from reflectance principal component analysis (PCA) scores grouped by sex in non-exposed spiny toads *Bufo spinosus*, for **a** throat reflectance, **b** ventral reflectance and **c** dorsal reflectance. PC1 and PC2 values show the contribution of the axes to the total variation. Each dot represents an individual. Individuals with similar reflectance (PCA scores) are near each other and individuals with dissimilar reflectance are farther from each other. Circles represent females and triangles represent males. The circle and triangle at the center of each ellipse represent the centroid of PCA scores for that group



 Table 1
 Relationships between reflectance PCA scores and sex in spiny toads (Bufo spinosus)

Response variables	Explanatory variable	Estimate ± SE	p value
Throat reflectance			
PCA 1	Sex	0.61 ± 0.28	0.036
PCA 2	Sex	-0.41 ± 0.29	0.174
Ventral reflectance			
PCA 1	Sex	1.11 ± 0.33	0.002
PCA 2	Sex	-0.18 ± 0.29	0.539
Dorsal reflectance			
PCA 1	Sex	0.69 ± 0.38	0.082
PCA 2	Sex	1.05 ± 0.3 0.002	

Values are estimates obtained from linear regressions with female as a reference level. PC scores were obtained from a PCA including hue, UV-saturation, UV-luminance, total brightness (B1 300–700), yellow-orange saturation, violet-blue saturation, and green saturation. The variables that correlated the most with axis 1 (PCA 1) and axis 2 (PCA 2) are identifiable in Fig. 1 and Appendix B. Achromatic components (UV, luminance) are mainly correlated with PCA 1, while chromatic components (e.g. hue) are mostly correlated with PCA 2. Values in bold are significant at $\alpha\!=\!0.05$

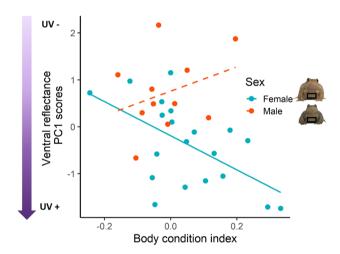


Fig. 2 Relationship between ventral reflectance and body condition index in male and female spiny toads *Bufo spinosus*. More negative values on the PC1 axis represent stronger reflectance in the UV. Females and males are represented by turquoise and orange dots, respectively. The full line represents a significant relationship, the dashed line a non-significant relationship

sex (being higher in males than females, 0.950 ± 0.301 , p = 0.004, values are LM estimates \pm standard error), body condition (-3.639 ± 1.270 , p = 0.008), and their interaction (6.237 ± 2.729 , p = 0.030). Indeed, body condition was negatively related with ventral PC1 in females (-3.64 ± 1.27 , p = 0.008, values are Linear Models estimates \pm standard error, Fig. 2), but not in males (1.53 ± 1.46 , p = 0.302, Fig. 2), and body condition did

not correlate with the reflectance of other body parts (p > 0.157) for all tests).

Effect of agrochemical exposure on reflectance and body condition

Nicosulfuron treatment decreased the hue of throat reflectance in females, whereas AMPA increased yellow-orange saturation of the throat in males (Table 2, Fig. 3A). In females, AMPA had a slight effect on ventral and dorsal reflectance, with a marginally higher ventral hue and increased yellow-orange dorsal saturation (Table 2, Fig. 3A). When testing the effects of body condition on reflectance in the exposed groups (AMPA and Nicosulfuron exposed individuals), we observed no relationship between body condition and reflectance for any body part, only the effect of sex is found significant (Appendix C).

Discussion

In this study, we highlighted a strong sexual dichromatism in spiny toads. Unexpectedly, the throat and ventral areas of females reflected more in the UV spectrum than those of males, suggesting these body parts might be critical in intra-specific signalling. In contrast, the throats and ventral areas of males were rather yellow-orange, with low UV reflectance. The dorsal area spectrum of females was dominated by yellow-orange, whereas that of males by green. Importantly, females with higher UV reflectance (ventral area only) were in better body condition. Moreover, throat colouration of both females and males was influenced by agrochemicals. The throats of AMPA-exposed males were more saturated in yellow-orange than that of control males, and Nicosulfuron exposure decreased the reflectance hue of the throat in females. We also observed a marginal decrease in the hue of the ventral area of AMPA-exposed females and a marginal increase of the yellow-orange colour of their back.

Although sexual dichromatism is not the most prevalent secondary sexual trait in amphibians (Bell and Zamudio 2012; Bell et al. 2017), adult males of several species are more brightly coloured than adult females (Bell and Zamudio 2012), this being considered as a visual signal facilitating gender recognition, enhanced by striking postures that highlight specific body parts (Sztatecsny et al. 2010). In *Bufo bufo* for instance, males are yellower-greener and brighter than females (Ujhegyi and Bókony 2020). Sexual dichromatism with colouration conspicuousness biased toward males is common among most vertebrates (Lifshitz and St Clair 2016), but some species of anurans display femalebiased colouration conspicuousness, in which females undergo colour transformation, often resulting in more



Table 2 Relationships between reflectance PCA scores and pesticide exposure according to sex in spiny toads (*Bufo spinosus*)

Response variables	Explanatory variables	Females		Males	
		Estimate ± SE	p value	Estimate ± SE	p value
Throat reflec	ctance				
PCA 1	AMPA	-0.09 ± 0.26	0.724	0.84 ± 0.38	0.036
	Nicosulfuron	-0.23 ± 0.29	0.432	0.58 ± 0.37	0.132
PCA 2	AMPA	-0.33 ± 0.28	0.246	-0.58 ± 0.5	0.257
	Nicosulfuron	-0.92 ± 0.31	0.004	-0.2 ± 0.49	0.680
Ventral refle	ctance				
PCA 1	AMPA	-0.38 ± 0.2	0.699	-0.07 ± 0.38	0.850
	Nicosulfuron	-0.12 ± 0.3	0.789	-0.28 ± 0.37	0.459
PCA 2	AMPA	0.56 ± 0.33	0.093	-0.23 ± 0.43	0.605
	Nicosulfuron	0.14 ± 0.36	0.692	-0.03 ± 0.42	0.950
Dorsal reflec	ctance				
PCA 1	AMPA	0.73 ± 0.37	0.055	0.19 ± 0.26	0.474
	Nicosulfuron	-0.15 ± 0.41	0.712	0.03 ± 0.26	0.913
PCA 2	AMPA	-0.14 ± 0.27	0.608	-0.21 ± 0.32	0.530
	Nicosulfuron	-0.2 ± 0.3	0.510	0.53 ± 0.32	0.103

Values are estimates obtained from linear regressions with control as a reference level. PC scores were obtained from a PCA including hue, UV-saturation, UV-luminance, total brightness (B1 300–700), yellow-orange saturation, violet-blue saturation, and green saturation. The variables that correlated the most with axis 1 (PCA 1) and axis 2 (PCA 2) are identifiable in Fig. 2 and Appendix B. Achromatic components (UV, luminance) are mainly correlated with PCA 1, while chromatic components (e.g. hue) are mostly correlated with PCA 2. Values in bold are significant at α =0.05. *Represents relationships close to statistical significance (p<0.10)

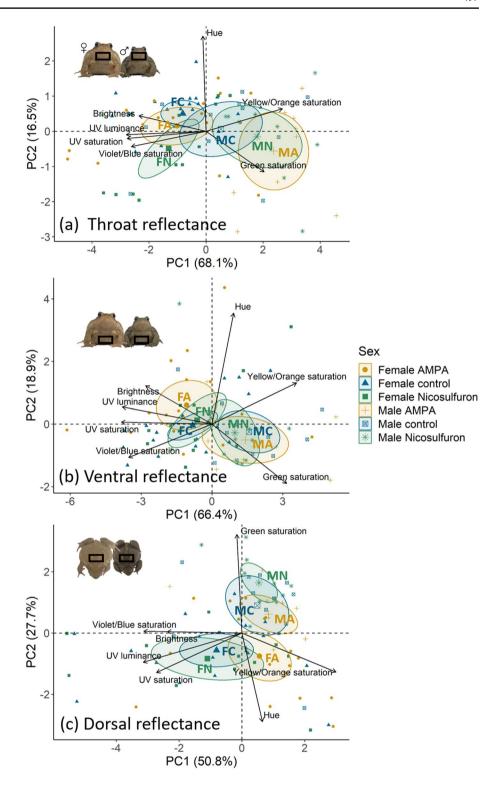
ornate colourations (Portik et al. 2019). To our knowledge, our study is the first to report a female-biased UV reflectance in amphibians, where females expressed throat and ventral reflection in the UV spectrum, while males did not.

UV reflectance has already been shown in males of many species. Diverse taxa possess retinal photopigments that allow UV vision (Jacobs 1992), and UV sensitivity has been suggested to be useful in making crucial visual discriminations, for example allowing to highlight targets against their background and thus aid in their detection (Jacobs 1992). As such, UV colouration in males has been shown to be used in mate choice in birds (Prum 2006), but also in lizards (Pérez i de Lanuza and Font 2007; Bajer et al. 2010) and amphibians (Secondi et al. 2012), the UV reflection allowing to increase sexual dichromatism and male conspicuousness (Pérez i de Lanuza and Font 2007). Here, we highlighted intense UV reflection in female spiny toads, suggesting either that mate choice can also be performed by males, as it has been found in many species (Clutton-Brock 2009), or that it allows males to detect the presence of females once they arrive in the breeding site. As explosive breeders, spiny toad males gather in very large numbers in the breeding ponds days to weeks before the arrival of females, and often place themselves either at the bottom of the pond or close to the shore below the water surface. In both cases they would spot the females from below. The strong ventral and throat UV reflectance of females could allow them to be easily discriminated from the background and allow males to visually distinguish them from other males. These two hypotheses need to be tested.

The mate choice hypotheses can be strengthened by the fact that colouration is often an honest signal of individuals' quality and ability to reproduce (Baatrup and Junge 2001; Clutton-Brock 2009; Shenoy 2012; Blévin et al. 2014). In birds, UV colouration is already known to reflect individual condition and quality (Prum 2006), and to be related to age, sex, and morphology in lizards (Martin et al. 2013). In our study, we show that females with higher UV reflectance were in better body condition, which is consistent with an honest signal of quality in female spiny toads. The relationship between colouration and quality is common in various species, as only individuals in good condition can afford to allocate pigments for trait colouration without compromising other functions (Hamilton and Zuk 1982). UV colouration might be costly to produce (Senar 2006), and its production could be traded off with many other functions. Colouration can be influenced by immunocompetence and parasite load (Molnár et al. 2012; Olsson et al. 2013), testosterone levels (Cox et al. 2005, 2008), and oxidative stress (Simons et al. 2012), which could all be influenced by exposure to agrochemicals (Mann et al. 2009; Brandt et al. 2016; Cheron et al. 2022). Our results thus suggest that UV colour could be an important female-condition signal for males in amphibians. To understand the underlying mechanisms, we



Fig. 3 Ordination plot from reflectance principal component analysis (PCA) scores grouped by sex and agrochemical treatment in spiny toads Bufo spinosus, for a throat reflectance, b ventral reflectance and c dorsal reflectance. PC1 and PC2 values show the contribution of the axes to the total variation. Each combination of sex and treatment is represented by a different symbol. Each dot represents an individual. In each sex, individuals were exposed either to tap water (control, FC = female control, MC = male control), Nicosulfuron (FN = female Nicosulfuron, MN = Male Nicosulfuron) or AMPA (FA = female AMPA, MA = male AMPA). The symbol at the centre of each ellipse represents the centroid of PCA scores for that group



should further investigate if UV colouration in this species is linked to immunity or hormone levels (such as oestrogen), and additionally to fecundity or investment in reproduction, as it has been shown in lizards (Kopena et al. 2020).

We were here able to show that spiny toads are sexually dichromatic, and this dichromatism can be affected by

agrochemicals. Numerous agrochemicals have enzyme- and endocrine-disrupting capabilities (Colborn et al. 1993; Khan and Law 2005; Lifshitz and St Clair 2016), which could lead to alterations of gonadal form and function (McCoy et al. 2008). As colouration is linked to hormone expression (Richards 1982; Rand 1992; Tang et al. 2014) and reproductive



abilities (Clutton-Brock 2009; Kopena et al. 2020), it is also likely to be disrupted by agrochemicals. Pigments involved in trait colouration, such as melanin, may be physiologically costly to produce (Jawor and Breitwisch 2003), and could be traded off with the costs of agrochemical exposure (e.g. reduced growth and increased susceptibility to disease (Baker et al. 2013). Additionally, agrochemicals, including AMPA, can cause damage to organisms by producing free radicals that overwhelm the antioxidant system (Galván and Alonso-Alvarez 2009; Cheron et al. 2022). Thus, pigments may rather be allocated to immune- or antioxidant-system (detoxification mechanisms, (McGraw 2005)), rather than to ornamentation in exposed individuals (Faivre et al. 2003; Arellano-Aguilar and Macías Garcia 2008; Lifshitz and St Clair 2016). Exposure to agrochemicals and other pollutants results in reduced carotenoid colouration (Baatrup and Junge 2001; Alonso-Alvarez and Galván 2011; Shenoy 2012; Lifshitz and St Clair 2016), probably because of the allocation of dietary carotenoids to counteract oxidative stress (Arellano-Aguilar and Macías Garcia 2008), and also to a decrease in the expression of brown melanin and an increase in black melanin (Lifshitz and St Clair 2016).

Interestingly, we show that, in AMPA-exposed males, throat yellow-orange colouration was more vivid than that of control males, probably indicating higher levels of carotenoids (Blount and McGraw 2008). This was unexpected given previous evidence that agrochemicals lead to reduced carotenoid colouration (Baatrup and Junge 2001; Alonso-Alvarez and Galván 2011; Shenoy 2012; Lifshitz and St Clair 2016). This effect either suggests that (1) AMPA concentrations used in our study were too low to induce oxidative damages in adults, or (2) males are not able to mobilize carotenoids to counteract oxidative stress [because carotenoids may have low antioxidant effects as shown in other species (Costantini and Møller 2008)]. These two hypotheses need to be untangled by evaluating oxidative stress concomitantly with colouration. One additional hypothesis could be that the observed pattern results from endocrine disruption. In vitro and in vivo studies have shown that glyphosate-based herbicides and AMPA exhibit estrogenlike properties in various taxa (Uren Webster et al. 2014; Jarrell et al. 2020; Milesi et al. 2021). In zebra fish (Danio rerio) for instance, exposure to glyphosate-based herbicide increases the expression of ovarian aromatase, an enzyme which catalyses the conversion of testosterone to estradiol in the gonads of females (Uren Webster et al. 2014). Estradiol exposition in male African clawed frogs Xenopus laevis led to sub-cellular events indicative of disrupted testicular development and decreased testosterone concentrations (Hecker et al. 2005). Consequently, AMPA exposure could decrease testosterone concentrations in male toads through estrogenic effects. Because a trade-off exists between testosterone and immunity (Peters 2007; Schroderus et al. 2010), if AMPA is related to lower testosterone concentrations, polluted males could afford mobilizing enough carotenoids to counteract oxidative stress and in parallel display a more yellow-orange throat colour. Estradiol, in addition to testosterone levels, need to be further assessed in exposed individuals to test for this last hypothesis and understand these variations.

AMPA and Nicosulfuron were also shown to respectively increase ventral and decrease throat hue in females. Hue represents the colour shade and is produced by a combination of dermal chromatophores, epidermal melanocytes, and epidermal diffraction gratings (McNamara et al. 2016). In toads, throat hue positively correlates with age, body mass and body size, and thus represents an honest signal of condition and age (Zamora-Camacho and Comas 2019). As all experimental groups were balanced in size, this suggests that exposed females have lost this honest signal. Additionally, older toads are darker (Zamora-Camacho and Comas 2019). In this study, we did not evaluate individual's age, which could have been done via skeletochronology. Since age correlates with size in amphibians (Shine 1979), even with some variance (Olsson and Shine 1996; Moreno-Rueda et al. 2021), we hypothesized that age-class might be similar between treatments. This suggests Nicosulfuron, and to a lesser extent AMPA, exposure could mimic or accelerate integument aging in exposed females, probably linked to telomere attrition, which might be a cost for maintaining colouration (Giraudeau et al. 2016). Telomere length should be analysed to test this hypothesis. Lastly, in another toad species, hue has been shown to decrease in intersex individuals (individuals with mixed-sex gonads or gonadal abnormalities; Ujhegyi and Bókony 2020), which effect is often found in response to herbicide (Howe et al. 2004; Lanctôt et al. 2014; Ujhegyi and Bókony 2020). This could indicate that Nicosulfuron has caused gonadal abnormalities in females, which effect might also be found with the exposition to AMPA, leading to a masculinisation of their dorsal reflectance (more saturated in yellow-orange in comparison to control females). Interestingly, the effect of body condition on UV reflectance was only found in control females and disappeared when considering exposed females. This suggests that in female spiny toads, agrochemicals could fade the honest signal carried by UV reflectance. Our results highlight sex-dependent responses of spiny toads to environmentally-relevant agrochemical exposure. By influencing mate attractiveness, these sex-dependent effects are likely to lead to reduced reproductive abilities and breeding success (Shenoy 2012; Ujhegyi and Bókony 2020; Yang et al. 2021).

Conclusion

We here provide the first evidence of UV reflectance in female spiny toads as a significant honest signal of quality for their mates. To go further, we would need to investigate



whether male choice toward females does depend on UV colouration, and if this feature correlates with increased fecundity or investment in reproduction. Additionally, we highlighted that AMPA and Nicosulfuron exposure affected throat colouration in males and females, but in a sex-dependent manner, which can bear consequences on mate attractiveness, and thus individuals' fitness in agricultural landscapes. Further investigations need to be conducted to better understand the underlying mechanisms disrupted by agrochemical exposure.

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Author contribution statement Co-first authors ST and LLS contributed equally to this manuscript, and each has the right to list themselves first in author order on their CVs. MC carried out fieldwork. ST and MC carried out the exposure experiment. AD performed the spectrophotometer measurements, and preliminary statistical analyses. ST and LLS analysed the data and wrote the first draft of the manuscript; all authors read the manuscript and provided critical reviews on previous versions. FB conceptualized the study, acquired the funding and supervised the global project.

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Data availability The data that support the findings of this study are available in the supplementary material of this article.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

Ethical statement All applicable institutional and/or national guidelines for the care and use of animals were followed. This work was approved by the French authorities (COMETHEA ethic committee and Ministère de L'Enseignement Superieur, de la Recherche et de L'innovation) under permits APAFIS#29265i2021012014416948 and DREAL/2020D/8041.

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References

- Adams E, Leeb C, Brühl CA (2021) Pesticide exposure affects reproductive capacity of common toads (*Bufo bufo*) in a viticultural landscape. Ecotoxicology 30:213–223. https://doi.org/10.1007/s10646-020-02335-9
- Alonso-Alvarez C, Galván I (2011) Free radical exposure creates paler carotenoid-based ornaments: a possible interaction in the

- expression of black and red traits. PLoS One 6:e19403. https://doi.org/10.1371/journal.pone.0019403
- Arellano-Aguilar O, Macías Garcia C (2008) Exposure to pesticides impairs the expression of fish ornaments reducing the availability of attractive males. Proc R Soc B Biol Sci 275:1343–1351. https://doi.org/10.1098/rspb.2008.0163
- Baatrup E, Junge M (2001) Antiandrogenic pesticides disrupt sexual characteristics in the adult male guppy *Poecilia reticulata*. Environ Health Perspect 109:1063–1070. https://doi.org/10.1289/ehp.011091063
- Badiane A, Martin M, Meylan S et al (2020) Male ultraviolet reflectance and female mating history influence female mate choice and male mating success in a polyandrous lizard. Biol J Linn Soc 130:586–598. https://doi.org/10.1093/biolinnean/blaa061
- Bagnara JT, Taylor JD, Hadley ME (1968) The dermal chromatophore unit. J Cell Biol 38:67–79. https://doi.org/10.1083/jcb. 38.1.67
- Bajer K, Molnár O, Török J, Herczeg G (2010) Female European green lizards (*Lacerta viridis*) prefer males with high ultraviolet throat reflectance. Behav Ecol Sociobiol 64:2007–2014. https://doi.org/10.1007/s00265-010-1012-2
- Baker NJ, Bancroft BA, Garcia TS (2013) A meta-analysis of the effects of pesticides and fertilizers on survival and growth of amphibians. Sci Total Environ 449:150–156. https://doi.org/10.1016/j.scitotenv.2013.01.056
- Barzaghi B, Melotto A, Cogliati P et al (2022) Factors determining the dorsal coloration pattern of aposematic salamanders. Sci Rep 12:17090. https://doi.org/10.1038/s41598-022-19466-0
- Bell RC, Zamudio KR (2012) Sexual dichromatism in frogs: natural selection, sexual selection and unexpected diversity. Proc R Soc B Biol Sci 279:4687–4693. https://doi.org/10.1098/rspb.2012. 1609
- Bell RC, Webster GN, Whiting MJ (2017) Breeding biology and the evolution of dynamic sexual dichromatism in frogs. J Evol Biol 30:2104–2115. https://doi.org/10.1111/jeb.13170
- Berger G, Graef F, Pfeffer H (2013) Glyphosate applications on arable fields considerably coincide with migrating amphibians. Sci Rep 3:1–5. https://doi.org/10.1038/srep02622
- Blévin P, Tartu S, Angelier F et al (2014) Integument colouration in relation to persistent organic pollutants and body condition in arctic breeding black-legged kittiwakes (*Rissa tridactyla*). Sci Total Environ 470–471:248–254. https://doi.org/10.1016/j.scitotenv.2013.09.049
- Blount JD, McGraw KJ (2008) Signal functions of carotenoid colouration. In: Carotenoids. Springer, pp 213–236
- Bradbury JW, Vehrencamp SL (1998) Principles of animal communication. Sinauer Associates Inc, Sunderland
- Brandt A, Gorenflo A, Siede R et al (2016) The neonicotinoids thiacloprid, imidacloprid, and clothianidin affect the immunocompetence of honey bees (*Apis mellifera* L.). J Insect Physiol 86:40– 47. https://doi.org/10.1016/j.jinsphys.2016.01.001
- Brühl CA, Schmidt T, Pieper S, Alscher A (2013) Terrestrial pesticide exposure of amphibians: an underestimated cause of global decline? Sci Rep 3:1135. https://doi.org/10.1038/srep01135
- Burkhardt D, Finger E (1991) Black, white and uv: How birds see birds. Naturwissenschaften 78:279–280. https://doi.org/10.1007/ BF01134359
- Caro T (2005) The adaptive significance of coloration in mammals. Bioscience 55:125. https://doi.org/10.1641/0006-3568(2005) 055[0125:TASOCI]2.0.CO;2
- Cessna AJ, Donald DB, Bailey J, Waiser M (2015) Persistence of the sulfonylurea herbicides Sulfosulfuron, Rimsulfuron, and Nicosulfuron in farm dugouts (ponds). J Environ Qual 44:1948–1955. https://doi.org/10.2134/jeq2014.11.0503
- Cheron M, Costantini D, Angelier F et al (2022) Aminomethylphosphonic acid (AMPA) alters oxidative status during embryonic



development in an amphibian species. Chemosphere 287:131882. https://doi.org/10.1016/j.chemosphere.2021.131882

- Clusella Trullas S, van Wyk JH, Spotila JR (2007) Thermal melanism in ectotherms. J Therm Biol 32:235–245. https://doi.org/10.1016/j.jtherbio.2007.01.013
- Clutton-Brock T (2009) Sexual selection in females. Anim Behav 77:3–11. https://doi.org/10.1016/j.anbehav.2008.08.026
- Colborn T, vom SFS, Soto AM (1993) Developmental effects of endocrine-disrupting chemicals in wildlife and humans. Environ Health Perspect 101:378–384. https://doi.org/10.1289/ehp. 93101378
- Costantini D, Møller AP (2008) Carotenoids are minor antioxidants for birds. Funct Ecol 22:367–370. https://doi.org/10.1111/j.1365-2435.2007.01366.x
- Côte J, Boniface A, Blanchet S et al (2018) Melanin-based coloration and host–parasite interactions under global change. Proc R Soc B Biol Sci 285:20180285. https://doi.org/10.1098/rspb.2018.0285
- Cox RM, Skelly SL, Leo A, John-Alder HB (2005) Testosterone regulates sexually dimorphic coloration in the eastern fence lizard, *Sceloporus undulatus*. Copeia 2005:597–608. https://doi.org/10.1643/CP-04-313R
- Cox RM, Zilberman V, John-Alder HB (2008) Testosterone stimulates the expression of a social color signal in Yarrow's Spiny Lizard, *Sceloporus jarrovii*. J Exp Zool Part Ecol Genet Physiol 309A:505–514. https://doi.org/10.1002/jez.481
- Delhey K, Peters A, Kempenaers B (2007) Cosmetic coloration in birds: occurrence, function, and evolution. Am Nat 169:S145–S158. https://doi.org/10.1086/510095
- Deutschlander ME, Phillips JB (1995) Characterization of an ultraviolet photoreception mechanism in the retina of an amphibian, the axolotl (*Ambystoma mexicanum*). Neurosci Lett 197:93–96. https://doi.org/10.1016/0304-3940(95)11905-C
- Doucet SM, Mennill DJ (2010) Dynamic sexual dichromatism in an explosively breeding Neotropical toad. Biol Lett 6:63–66. https://doi.org/10.1098/rsbl.2009.0604
- Dray AB, Dufour S (2007) The ade4 package: implementing the duality diagram for ecologists. J Stat Softw 22:1–20
- Duellman WE, Trueb L (1986) Biology of Amphibians. Copeia, New York McGraw-Hill
- Endler JA, Mappes J (2017) The current and future state of animal coloration research. Philos Trans R Soc B Biol Sci 372:20160352
- Faivre B, Grégoire A, Préault M et al (2003) Immune activation rapidly mirrored in a secondary sexual trait. Science 300:103–103. https://doi.org/10.1126/science.1081802
- Fleishman LJ, Loew ER, Leal M (1993) Ultraviolet vision in lizards. Nature 365:397–397. https://doi.org/10.1038/365397a0
- Galván I, Alonso-Alvarez C (2009) The expression of melanin-based plumage is separately modulated by exogenous oxidative stress and a melanocortin. Proc R Soc B Biol Sci 276:3089–3097. https://doi.org/10.1098/rspb.2009.0774
- Giraudeau M, Friesen CR, Sudyka J et al (2016) Ageing and the cost of maintaining coloration in the Australian painted dragon. Biol Lett 12:20160077. https://doi.org/10.1098/rsbl.2016.0077
- Goiran C, Bustamante P, Shine R (2017) Industrial melanism in the seasnake *Emydocephalus annulatus*. Curr Biol 27:2510-2513.e2. https://doi.org/10.1016/j.cub.2017.06.073
- Gomez D, Richardson C, Lengagne T et al (2009) The role of nocturnal vision in mate choice: females prefer conspicuous males in the European tree frog (*Hyla arborea*). Proc R Soc B Biol Sci 276:2351–2358. https://doi.org/10.1098/rspb.2009.0168
- Grandcoin A, Piel S, Baurès E (2017) AminoMethylPhosphonic acid (AMPA) in natural waters: its sources, behavior and environmental fate. Water Res 117:187–197. https://doi.org/10.1016/j. watres.2017.03.055
- Guillot H, Boissinot A, Angelier F et al (2016) Landscape influences the morphology of male common toads (*Bufo bufo*). Agric

- Ecosyst Environ 233:106–110. https://doi.org/10.1016/j.agee. 2016.08.032
- Hamilton WD, Zuk M (1982) Heritable true fitness and bright birds: a role for parasites? Science 218:384–387. https://doi.org/10.1126/science.7123238
- Hayes TB, Khoury V, Narayan A et al (2010) Atrazine induces complete feminization and chemical castration in male African clawed frogs (Xenopus laevis). Proc Natl Acad Sci 107:4612–4617. https://doi.org/10.1073/pnas.0909519107
- Heath J, Cipollini D, Stireman J (2013) The role of carotenoids and their derivatives in mediating interactions between insects and their environment. Arthropod-Plant Interact 7:1–20. https://doi.org/10.1007/s11829-012-9239-7
- Hecker M, Kim WJ, Park J-W et al (2005) Plasma concentrations of estradiol and testosterone, gonadal aromatase activity and ultrastructure of the testis in *Xenopus laevis* exposed to estradiol or atrazine. Aquat Toxicol 72:383–396. https://doi.org/10.1016/j. aquatox.2005.01.008
- Hews DK, Moore MC (1995) Influence of androgens on differentiation of secondary sex characters in tree lizards, *Urosaurus ornatus*. Gen Comp Endocrinol 97:86–102. https://doi.org/10.1006/gcen. 1995.1009
- Höbel G, Feagles O, Ruder E (2022) Diversity and sexual dichromatism in treefrog throat coloration: potential signal function? J Herpetol 56:294–301. https://doi.org/10.1670/21-047
- Howe CM, Berrill M, Pauli BD et al (2004) Toxicity of glyphosatebased pesticides to four North American frog species. Environ Toxicol Chem 23:1928–1938. https://doi.org/10.1897/03-71
- Hunt DM, Wilkie SE, Bowmaker JK, Poopalasundaram S (2001) Vision in the ultraviolet. Cell Mol Life Sci CMLS 58:1583–1598. https://doi.org/10.1007/PL00000798
- Jablonski NG (1998) Ultraviolet light-induced neural tube defects in amphibian larvae and their implications for the evolution of melanized pigmentation and declines in amphibian populations. J Herpetol 32:455–457. https://doi.org/10.2307/1565466
- Jacobs GH (1992) Ultraviolet vision in vertebrates. Am Zool 32:544–554. https://doi.org/10.1093/icb/32.4.544
- Jarrell ZR, Ahammad MU, Benson AP (2020) Glyphosate-based herbicide formulations and reproductive toxicity in animals. Vet Anim Sci 10:100126. https://doi.org/10.1016/j.vas.2020.100126
- Jawor JM, Breitwisch R (2003) Melanin ornaments, honesty, and sexual selection. Auk 120:249–265. https://doi.org/10.1642/0004-8038(2003)120[0249:MOHASS]2.0.CO;2
- Kawamoto A, Le Galliard J-F, Badiane A (2021) The role of social costs as a mechanism enforcing the honesty of ultraviolet-reflecting signals in a lizard. Biol J Linn Soc 133:1126–1138. https:// doi.org/10.1093/biolinnean/blab008
- Khan MZ, Law FCP (2005) Adverse effects of pesticides and related chemicals on enzyme and hormone systems of fish, amphibians and reptiles: a review. Proc Pakistan Acad Sci 42:315–323
- Kopena R, López P, Majlathova V, Martín J (2020) Sexually dichromatic coloration of female Iberian green lizards correlates with health state and reproductive investment. Behav Ecol Sociobiol 74:131. https://doi.org/10.1007/s00265-020-02915-z
- Lanctôt C, Navarro-Martín L, Robertson C et al (2014) Effects of glyphosate-based herbicides on survival, development, growth and sex ratios of wood frog (*Lithobates sylvaticus*) tadpoles. II: Agriculturally relevant exposures to Roundup WeatherMax® and Vision® under laboratory conditions. Aquat Toxicol 154:291–303. https://doi.org/10.1016/j.aquatox.2014.05.025
- Larramendy ML (2017) Ecotoxicology and genotoxicology: non-traditional aquatic models. Royal Society of Chemistry
- Lenhardt PP, Brühl CA, Berger G (2015) Temporal coincidence of amphibian migration and pesticide applications on arable fields in spring. Basic Appl Ecol 16:54–63. https://doi.org/10.1016/j.baae.2014.10.005



- LeNoir JS, McConnell LL, Fellers GM et al (1999) Summertime transport of current-use pesticides from California's Central Valley to the Sierra Nevada Mountain Range, USA. Environ Toxicol Chem 18:2715–2722. https://doi.org/10.1002/etc. 5620181210
- Lifshitz N, St Clair CC (2016) Coloured ornamental traits could be effective and non-invasive indicators of pollution exposure for wildlife. Conserv Physiol 4:cow028. https://doi.org/10.1093/conphys/cow028
- Lin JY, Fisher DE (2007) Melanocyte biology and skin pigmentation. Nature 445:843–850. https://doi.org/10.1038/nature05660
- Mack M, Beaty L (2021) The influence of environmental and physiological factors on variation in American toad (*Anaxyrus americanus*) dorsal coloration. J Herpetol 55:119–126. https://doi.org/10.1670/20-093
- Maia R, Eliason CM, Bitton P-P et al (2013) pavo: an R package for the analysis, visualization and organization of spectral data. Methods Ecol Evol 4:906–913. https://doi.org/10.1111/2041-210X.12069
- Mann RM, Hyne RV, Choung CB, Wilson SP (2009) Amphibians and agricultural chemicals: review of the risks in a complex environment. Environ Pollut 157:2903–2927. https://doi.org/10.1016/j.envpol.2009.05.015
- Martin M, Meylan S, Gomez D, Le Galliard J-F (2013) Ultraviolet and carotenoid-based coloration in the viviparous lizard *Zootoca* vivipara (Squamata: Lacertidae) in relation to age, sex, and morphology. Biol J Linn Soc 110:128–141. https://doi.org/10.1111/ bij.12104
- Martín J, López P (2009) Multiple color signals may reveal multiple messages in male Schreiber's green lizards, *Lacerta schreiberi*. Behav Ecol Sociobiol 63:1743–1755
- McConnell LL, LeNoir JS, Datta S, Seiber JN (1998) Wet deposition of current-use pesticides in the Sierra Nevada mountain range, California, USA. Environ Toxicol Chem 17:1908–1916. https://doi.org/10.1002/etc.5620171003
- McCoy KA, Bortnick LJ, Campbell CM et al (2008) Agriculture alters gonadal form and function in the toad *Bufo marinus*. Environ Health Perspect 116:1526–1532. https://doi.org/10.1289/ehp. 11536
- McGraw KJ (2005) The antioxidant function of many animal pigments: are there consistent health benefits of sexually selected colourants? Anim Behav 69:757–764. https://doi.org/10.1016/j.anbehav.2004.06.022
- McNamara ME, Orr PJ, Kearns SL et al (2016) Reconstructing carotenoid-based and structural coloration in fossil skin. Curr Biol 26:1075–1082. https://doi.org/10.1016/j.cub.2016.02.038
- Milesi MM, Lorenz V, Durando M et al (2021) Glyphosate herbicide: reproductive outcomes and multigenerational effects. Front Endocrinol 12:672532. https://doi.org/10.3389/fendo.2021. 672532/full
- Mirč M, Vukov T, Kijanović A, Kolarov NT (2023) Effects of background color on pigmentation, morphological traits, and behavior in the European tree frog (*Hyla arborea*, Hylidae, Anura) tadpoles. Contrib Zool 92:112–129. https://doi.org/10.1163/18759 866-bja10040
- Molnár O, Bajer K, Török J, Herczeg G (2012) Individual quality and nuptial throat colour in male European green lizards. J Zool 287:233–239. https://doi.org/10.1111/j.1469-7998.2012.00916.x
- Moreno-Rueda G, Reguera S, Zamora-Camacho FJ, Comas M (2021) Inter-individual differences in ornamental colouration in a Mediterranean lizard in relation to altitude, season, sex, age, and body traits. Diversity 13:158. https://doi.org/10.3390/d13040158
- Nilsson Sköld H, Aspengren S, Wallin M (2013) Rapid color change in fish and amphibians–function, regulation, and emerging applications. Pigment Cell Melanoma Res 26:29–38. https://doi.org/10.1111/pcmr.12040

- Norris KS, Lowe CH (1964) An analysis of background color-matching in amphibians and reptiles. Ecology 45:565–580. https://doi.org/10.2307/1936109
- Olsson M, Shine R (1996) Does reproductive success increase with age or with size in species with indeterminate growth? A case study using sand lizards (*Lacerta agilis*). Oecologia 105:175–178. https://doi.org/10.1007/BF00328543
- Olsson M, Stuart-Fox D, Ballen C (2013) Genetics and evolution of colour patterns in reptiles. Semin Cell Dev Biol 27:529–541. https://doi.org/10.1016/j.semcdb.2013.04.001
- Pérez i de Lanuza G, Font E (2007) Ultraviolet reflectance of male nuptial colouration in sand lizards (*Lacerta agilis*) from the Pyrenees. Amphib-Reptil 28:438–443. https://doi.org/10.1163/156853807781374764
- Peters A (2007) Testosterone and carotenoids: an integrated view of trade-offs between immunity and sexual signalling. BioEssays 29:427–430. https://doi.org/10.1002/bies.20563
- Portik DM, Bell RC, Blackburn DC et al (2019) Sexual dichromatism drives diversification within a major radiation of African amphibians. Syst Biol 68:859–875. https://doi.org/10.1093/sysbio/syz023
- Prum RO (2006) Anatomy, physics, and evolution of avian structural colors. In: Hill GE, McGraw KJ (eds) Bird coloration, vol I. Mechanisms and Measurements. Harvard University Press, Cambridge, pp 295–353
- Ptacek MB (2000) The role of mating preferences in shaping interspecific divergence in mating signals in vertebrates. Behav Processes 51:111–134. https://doi.org/10.1016/S0376-6357(00)00123-6
- R Core Team (2019) R: A Language and Environment for Statistical Computing
- Rand MS (1992) Hormonal control of polymorphic and sexually dimorphic coloration in the lizard *Sceloporus undulatus eryth-rocheilus*. Gen Comp Endocrinol 88:461–468. https://doi.org/10. 1016/0016-6480(92)90241-B
- Rashid B, Husnain T, Riazuddin S (2010) Herbicides and pesticides as potential pollutants: a global problem. In: Ashraf M, Ozturk M, Ahmad MSA (eds) Plant adaptation and phytoremediation. Springer, Dordrecht, pp 427–447
- Richards CM (1982) The alteration of chromatophore expression by sex hormones in the kenyan reed frog, *Hyperolius viridiflavus*. Gen Comp Endocrinol 46:59–67. https://doi.org/10.1016/0016-6480(82)90163-0
- Rudh A, Qvarnström A (2013) Adaptive colouration in amphibians.
 In: Seminars in Cell & Developmental Biology. Elsevier, pp 553–561
- Schiedt K (1989) New aspects of carotenoid metabolism in animals. In: Carotenoids: chemistry and biology. Springer, pp 247–268
- Schroderus E, Jokinen I, Koivula M et al (2010) Intra- and inter-sexual trade-offs between testosterone and immune system: implications for sexual and sexually antagonistic selection. Am Nat 176:E90–E97. https://doi.org/10.1086/656264
- Secondi J, Lepetz V, Théry M (2012) Male attractiveness is influenced by UV wavelengths in a newt species but not in its close relative. PLoS One 7:e30391
- Senar JC (2006) Bird colors as intrasexual signals of aggression and dominance. In: Hill GE, McGraw KJ, eds. Bird coloration, Vol.
 2. Function and evolution. MA: Harvard University, Cambridge, pp 93–136
- Sever DM, Staub NL (2011) Chapter 5—hormones, sex accessory structures, and secondary sexual characteristics in amphibians. In: Norris DO, Lopez KH (eds) Hormones and reproduction of vertebrates. Academic Press, London, pp 83–98
- Shenoy K (2012) Environmentally realistic exposure to the herbicide atrazine alters some sexually selected traits in male guppies. PLoS One 7:e30611. https://doi.org/10.1371/journal.pone.00306 11



Simons MJP, Cohen AA, Verhulst S (2012) What does carotenoid-dependent coloration tell? Plasma carotenoid level signals immunocompetence and oxidative stress state in birds—a meta-analysis. PLoS One 7:e43088. https://doi.org/10.1371/journal.pone.0043088

- Speybroeck J, Beukema W, Bok B, Van Der Voort J (2018) Guide Delachaux des amphibiens & reptiles de France et d'Europe. Delachaux et Nieslé
- Stoehr AM, McGraw KJ (2001) Ultraviolet reflectance of color patches in male *Sceloporus undulatus* and *Anolis carolinensis*. J Herpetol 35:168–171. https://doi.org/10.2307/1566045
- Stuart-Fox D, Moussalli A (2009) Camouflage, communication and thermoregulation: lessons from colour changing organisms. Philos Trans R Soc B Biol Sci 364:463–470. https://doi.org/10. 1098/rstb.2008.0254
- Stuart-Fox D, Moussalli A, Whiting MJ (2007) Natural selection on social signals: signal efficacy and the evolution of chameleon display coloration. Am Nat 170:916–930. https://doi.org/10.1086/522835
- Stückler S, Fuxjager MJ, Preininger D (2022) Evidence that catecholaminergic systems mediate dynamic colour change during explosive breeding events in toads. Biol Lett 18:20220337. https://doi.org/10.1098/rsbl.2022.0337
- Svensson PA, Wong BBM (2011) Carotenoid-based signals in behavioural ecology: a review. Behaviour 148:131–189. https://doi.org/10.1163/000579510X548673
- Sztatecsny M, Strondl C, Baierl A et al (2010) Chin up: are the bright throats of male common frogs a condition-independent visual cue? Anim Behav 79:779–786. https://doi.org/10.1016/j.anbehav.2010.01.003
- Sztatecsny M, Preininger D, Freudmann A et al (2012) Don't get the blues: conspicuous nuptial colouration of male moor frogs (*Rana arvalis*) supports visual mate recognition during scramble competition in large breeding aggregations. Behav Ecol Sociobiol 66:1587–1593. https://doi.org/10.1007/s00265-012-1412-6
- Tang Z-J, Lue S-I, Tsai M-J et al (2014) The hormonal regulation of color changes in the sexually dichromatic frog *Buergeria*

- robusta. Physiol Biochem Zool 87:397–410. https://doi.org/10. 1086/675678
- Tartu S, Renoirt M, Cheron M et al (2022) Did decades of glyphosate use have selected for resistant amphibians in agricultural habitats? Environ Pollut 310:119823. https://doi.org/10.1016/j.envpol.2022.119823
- Trudeau VL, Thomson P, Zhang WS et al (2020) Agrochemicals disrupt multiple endocrine axes in amphibians. Mol Cell Endocrinol 513:110861. https://doi.org/10.1016/j.mce.2020.110861
- Ujhegyi N, Bókony V (2020) Skin coloration as a possible non-invasive marker for skewed sex ratios and gonadal abnormalities in immature common toads (*Bufo bufo*). Ecol Indic 113:106175. https:// doi.org/10.1016/j.ecolind.2020.106175
- Uren Webster TM, Laing LV, Florance H, Santos EM (2014) Effects of Glyphosate and its formulation, roundup, on reproduction in Zebrafish (*Danio rerio*). Environ Sci Technol 48:1271–1279. https://doi.org/10.1021/es404258h
- Weaver RJ, Santos ES, Tucker AM et al (2018) Carotenoid metabolism strengthens the link between feather coloration and individual quality. Nat Commun 9:1–9. https://doi.org/10.1038/s41467-017-02649-z
- Wells KD (2007) The Ecology and Behavior of Amphibians. University of Chicago Press
- Yang C, Lim W, Song G (2021) Reproductive toxicity due to herbicide exposure in freshwater organisms. Comp Biochem Physiol Part C Toxicol Pharmacol 248:109103. https://doi.org/10.1016/j.cbpc. 2021.109103
- Zamora-Camacho FJ, Comas M (2019) Beyond sexual dimorphism and habitat boundaries: Coloration correlates with morphology, age, and locomotor performance in a toad. Evol Biol 46:60–70. https://doi.org/10.1007/s11692-018-9466-7

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